

Pathogen Reduction of Red Cell Concentrates

Laurence Corash, MD

Chief Scientific Officer, Cerus Corporation

Professor, University of California, School of Medicine, San Francisco

Disclosures

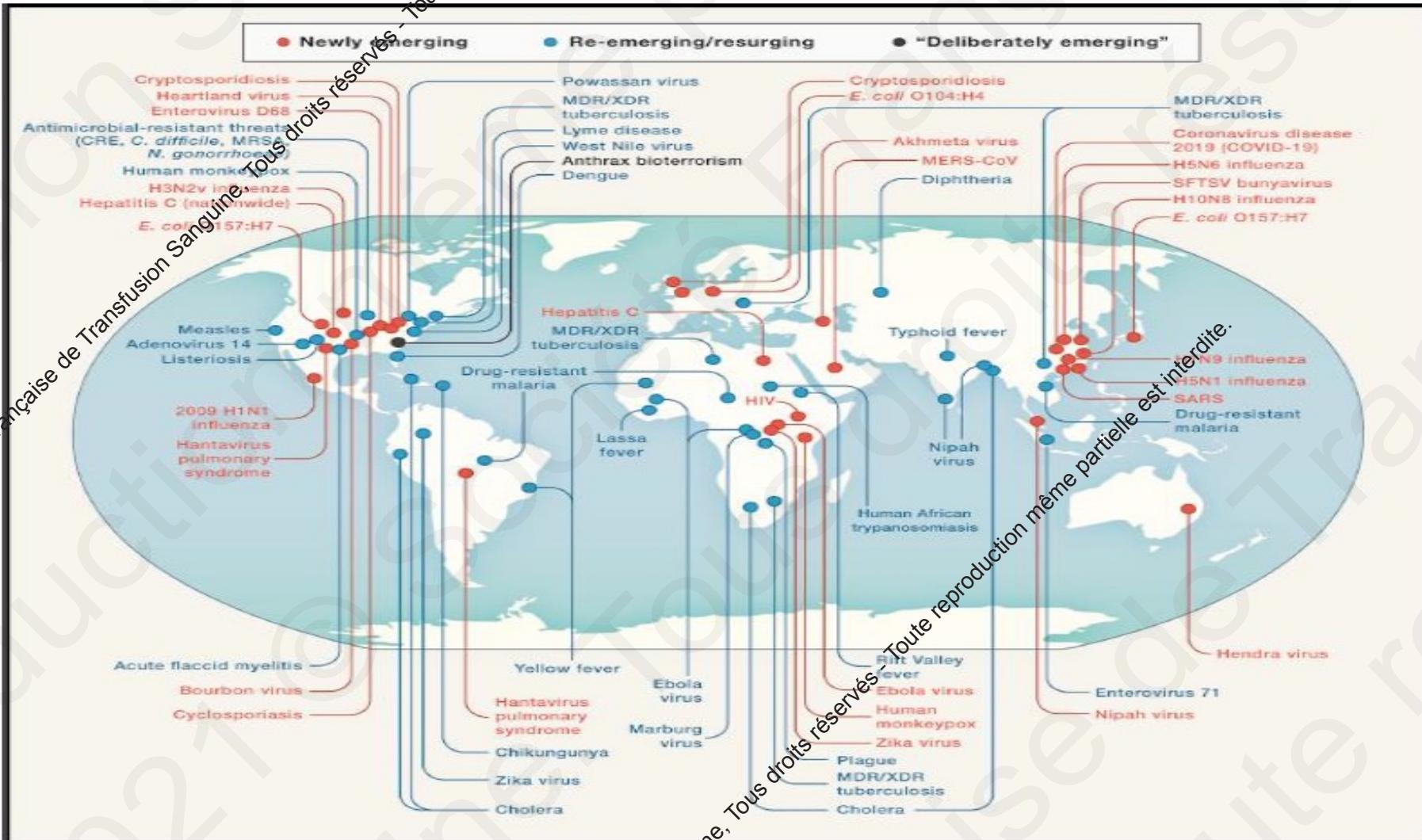
- Employee of Cerus Corporation
- Professor, Laboratory Medicine, University of California, SF
- Principal Investigator HHS/BARDA Contract for Pathogen Reduced RBC
- Principal Investigator HHS/FDA Contract for Development of New Compounds for Whole Blood Pathogen Reduction

Agenda

- Rationale for pathogen reduction, emerging pathogens, and pandemic preparedness
- Technology
 - Riboflavin + UVB
 - Amustaline-GSH
- Clinical experience
- Conclusions

History of Global Emerging and Re-Emerging Pathogens Entering Humans and the Blood Donor Population

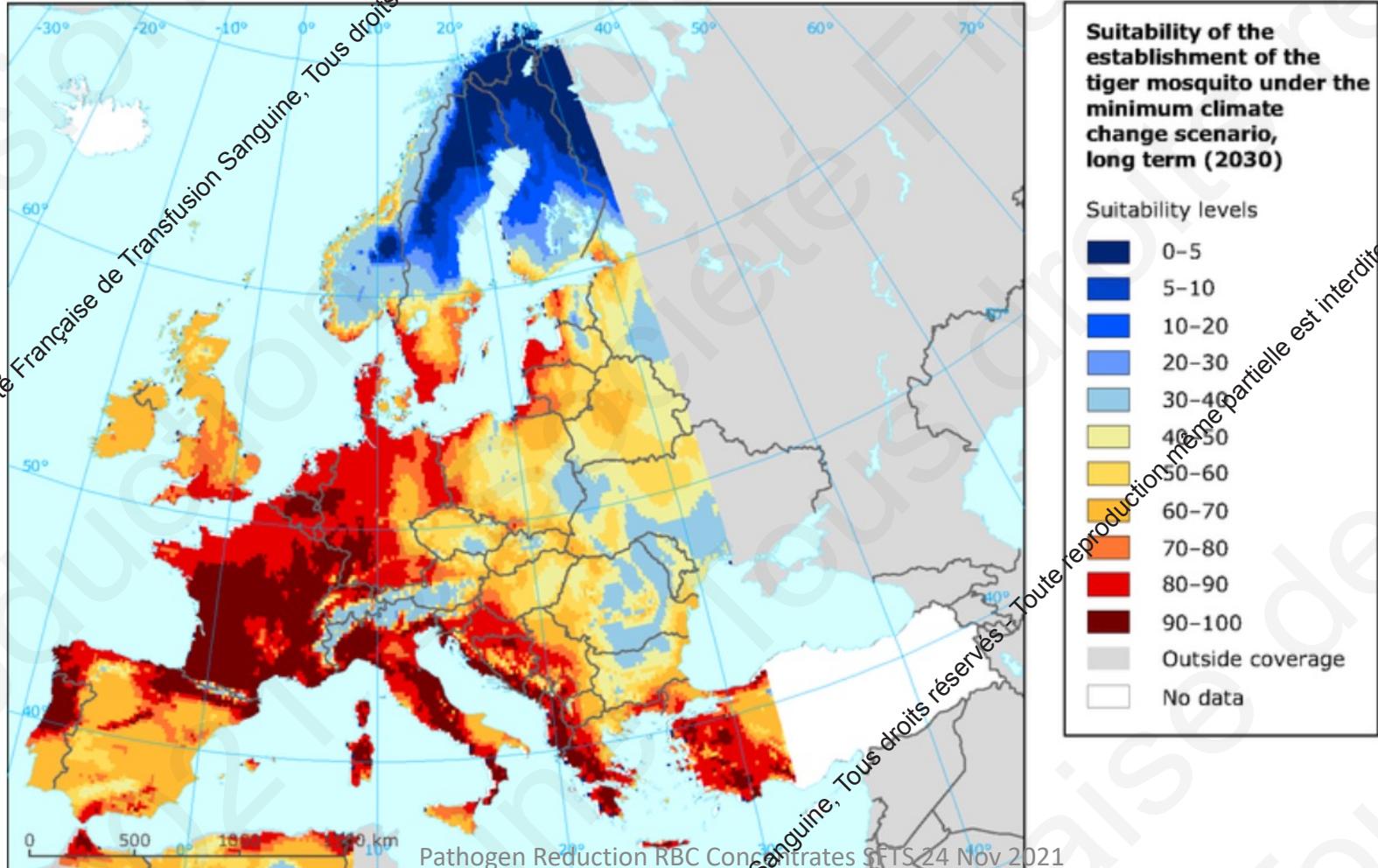
Morens and Fauci 2020 Cell ; 182:1077-1092



Suitability of Climate in Europe for the Asian Tiger Mosquito

Climate Change is Changing the Range of Insect Vectors

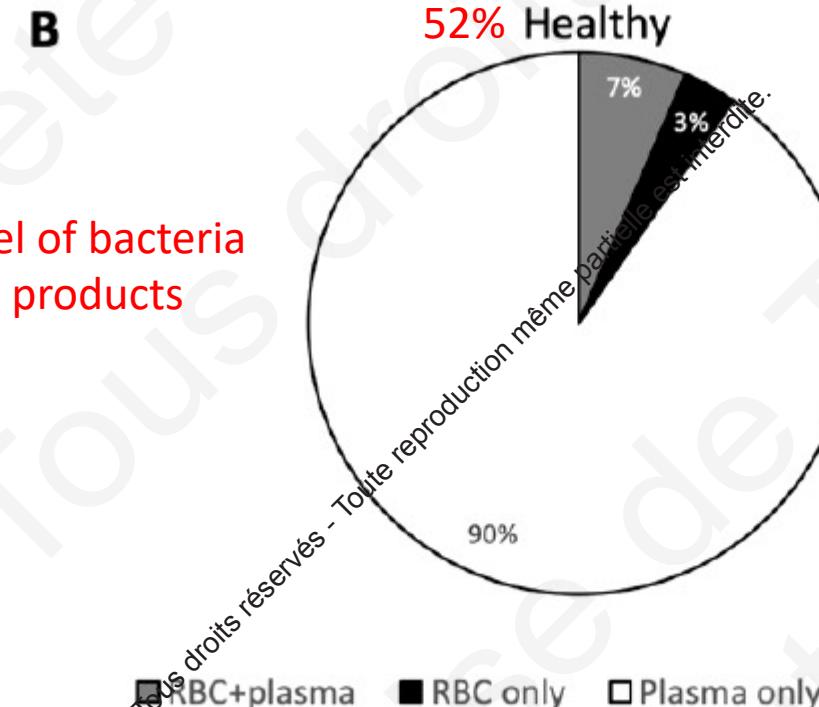
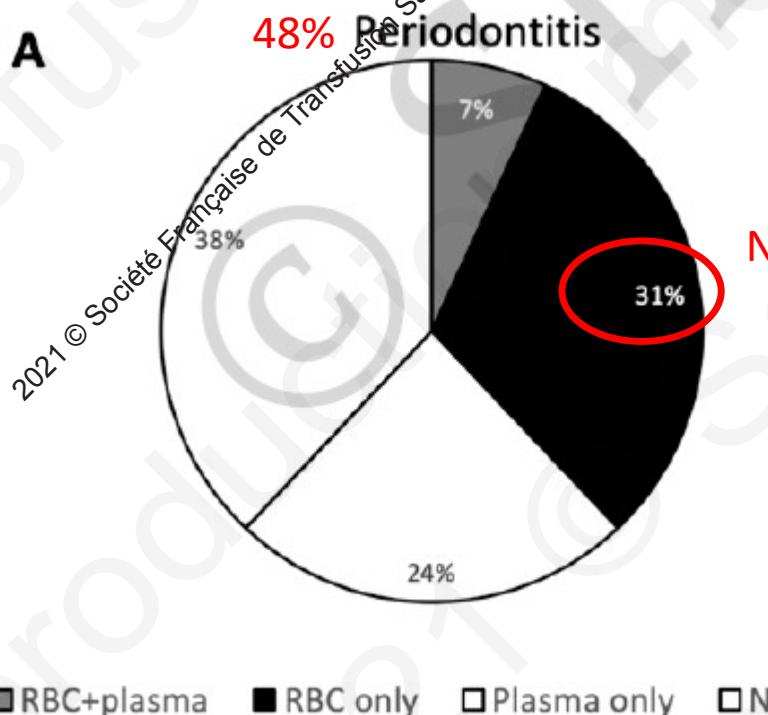
Transmits 25 different arboviruses

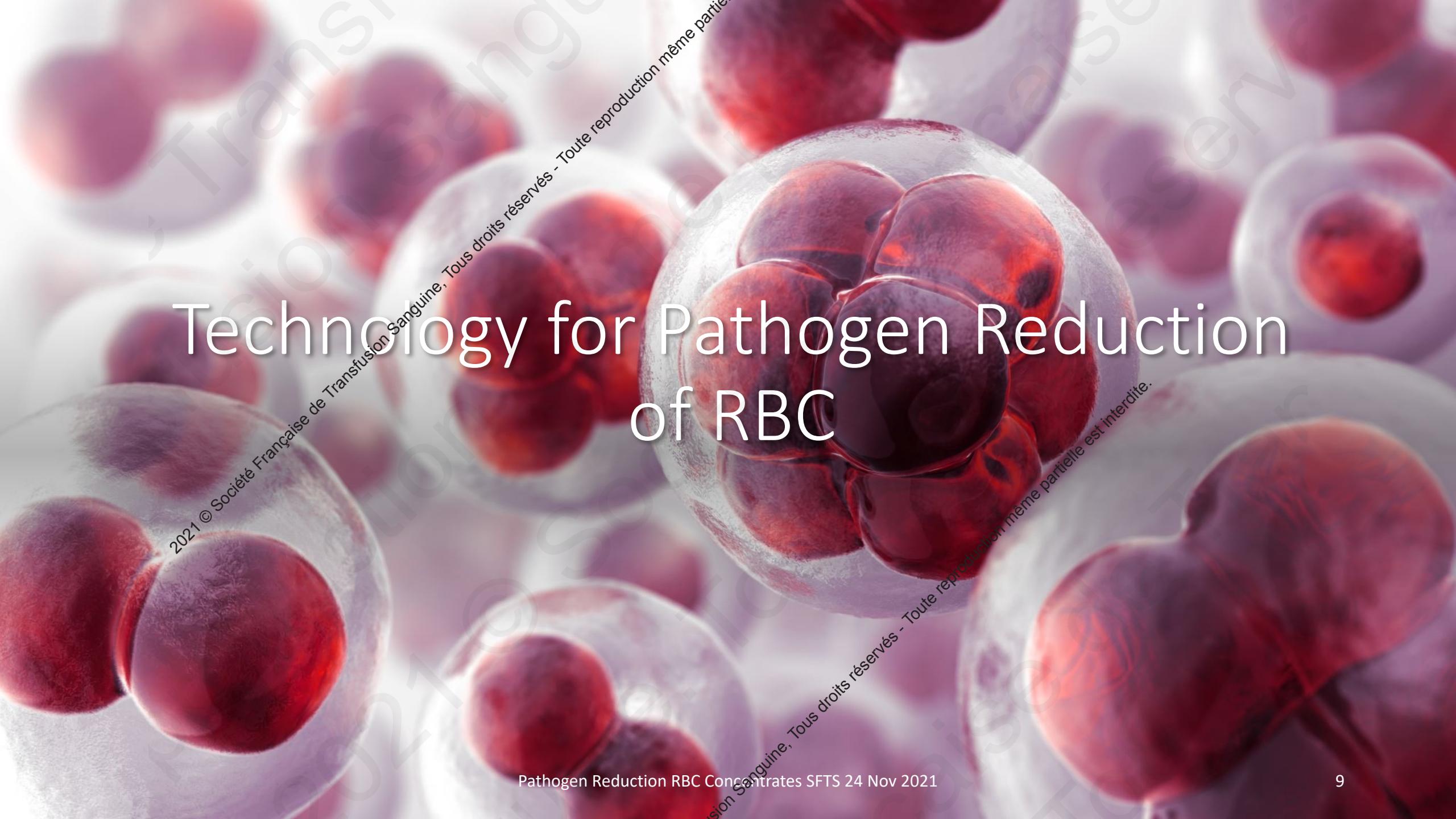


Bacteria in RBC Concentrates- Unrecognized Donor Risk of Periodontitis

Periodontitis increases risk of viable bacteria in freshly drawn blood donations

Christian Damgaard^{1,2}, Susanne G. Sækmose³, Martin Nilsson⁴, Mogens Kilian⁵, Claus H. Nielsen^{1,2}, Palle Holmstrup¹



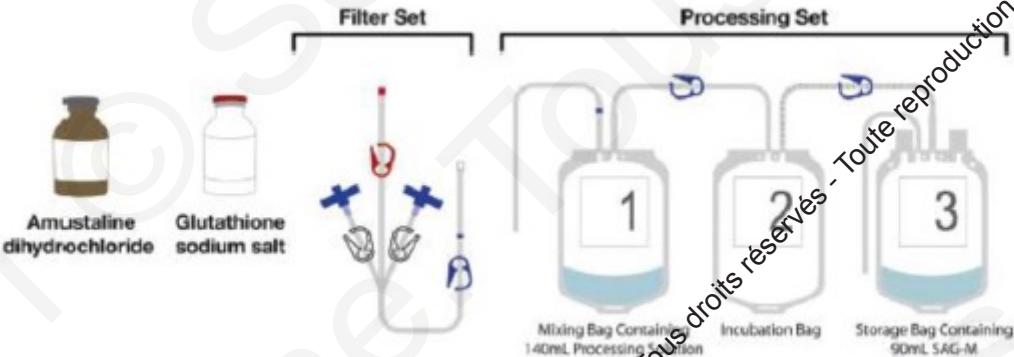


Technology for Pathogen Reduction of RBC

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Amustaline-GSH

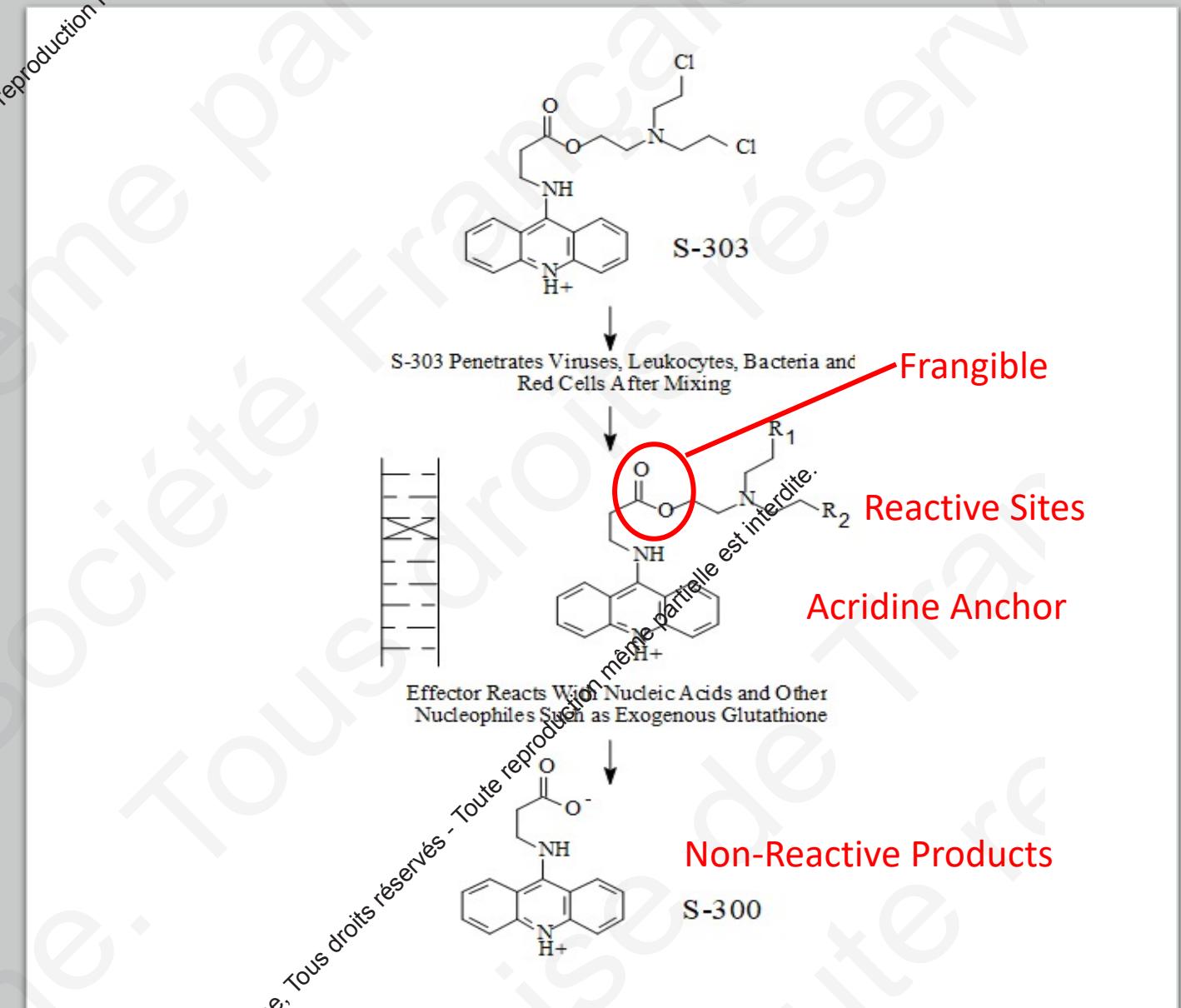
Amustaline- GSH Pathogen Reduction



S-303 MECHANISM OF ACTION

- Specific adduct formation by alkylation
- Does not require reactive oxygen species
- Degradation to inactive by-products

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Riboflavin-UV

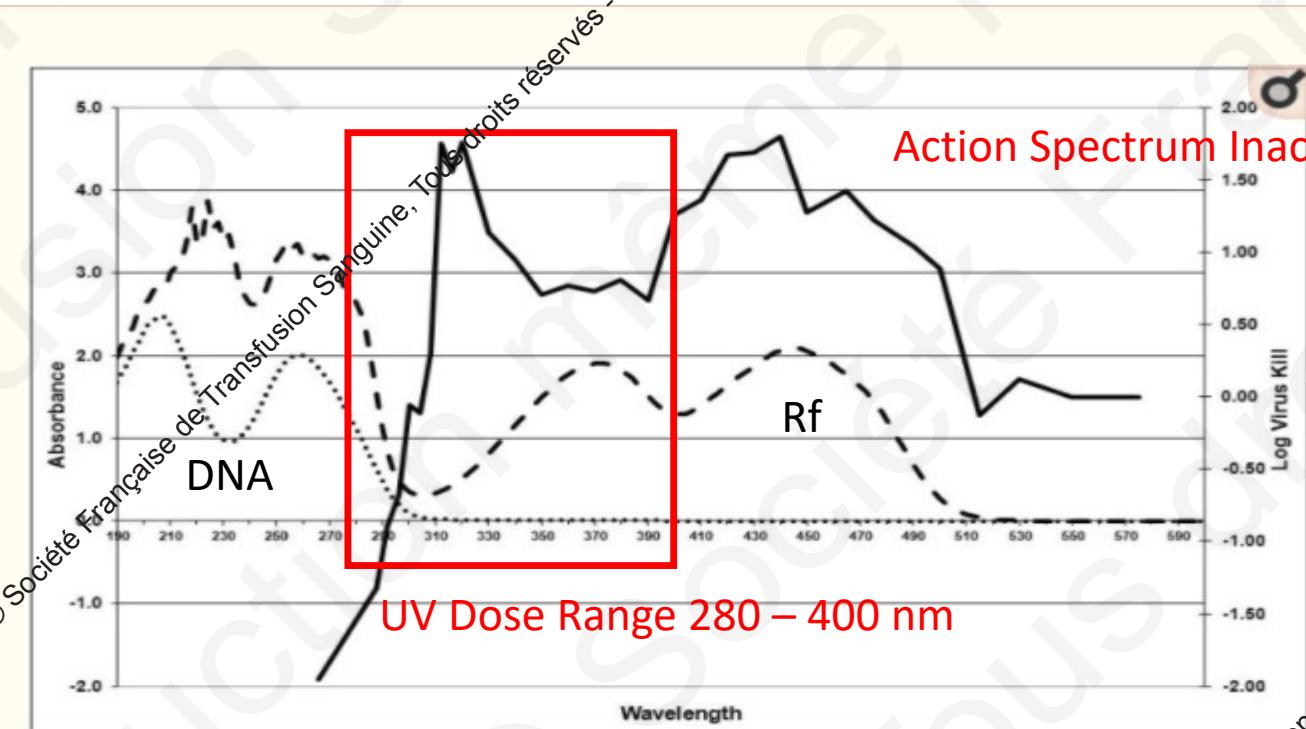
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○ + ●

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Improving the safety of whole blood-derived transfusion products with a riboflavin-based pathogen reduction technology

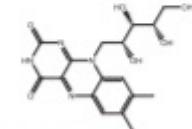
Susan Yonemura,¹ Suzann Doan,¹ Shawn Keil,¹ Raymond Goodrich,^{1,2} Heather Pidcock,¹ and Marcia Cardoso³



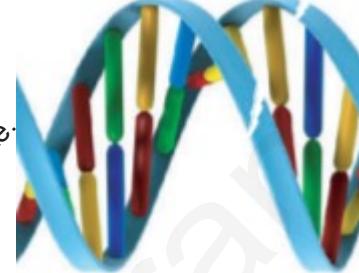
280nm 315nm

Mirasol

280-400nm

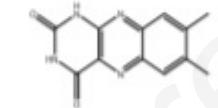


Riboflavin



8-oxodG
strand breaks

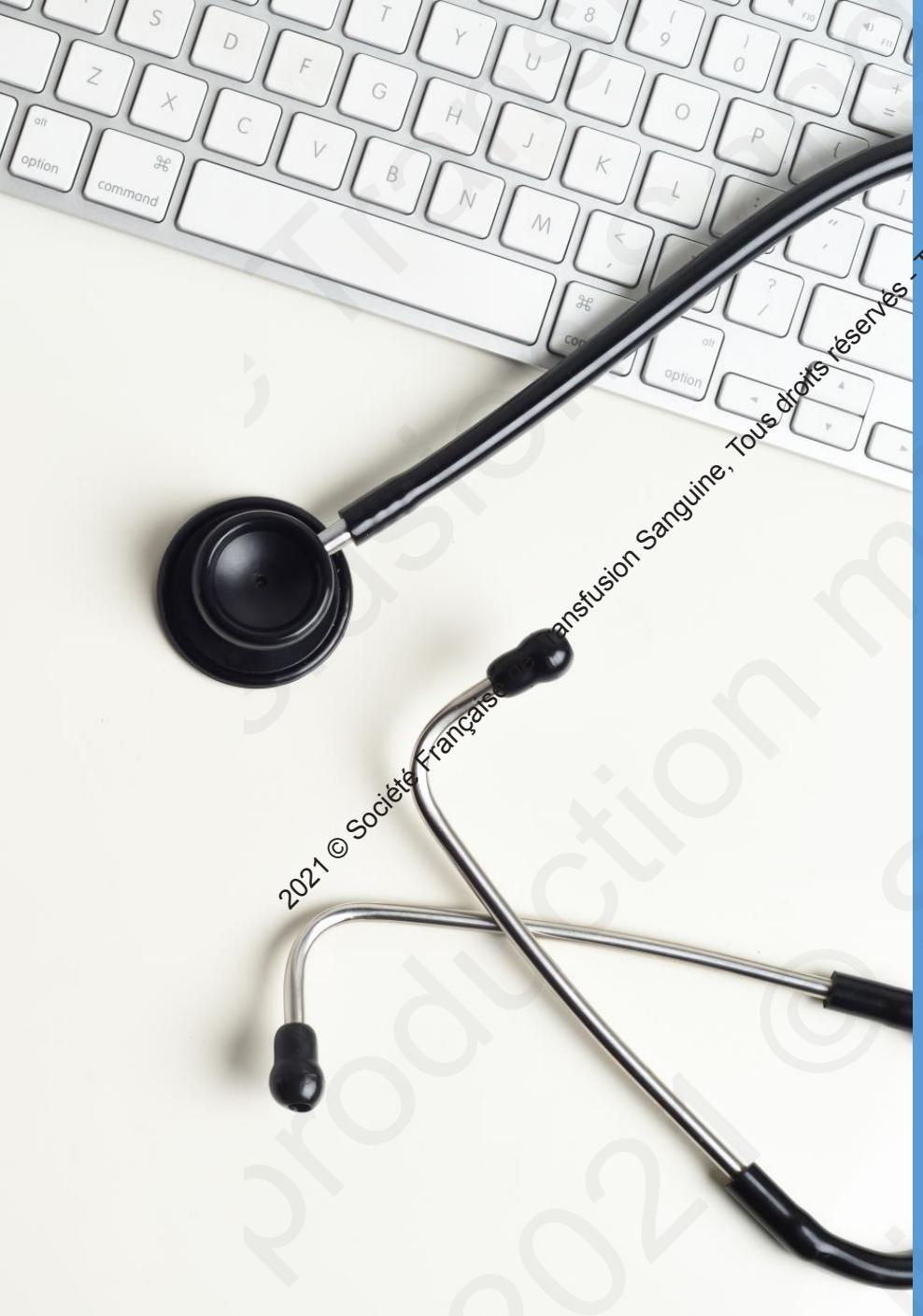
Reactive O₂



Lumichrome

Chemical and Biological Mechanisms of Pathogen Reduction Technologies

Janna M Mundt,¹ Lindsay Rouse,¹ Jeroen Van den Bossche,¹ and Raymond P Goodrich¹



Clinical Data

Pathogen Reduction Technology: Clinical Trials

- Amustaline-GSH
 - Phase 2: radiolabel post transfusion recovery and lifespan 35-day RBC
 - Phase 3
 - EU: Acute anemia – cardiovascular surgery (n = 52)
 - EU: Chronic anemia – transfusion dependent thalassemia (n = 80)
 - US: Acute anemia – all cause of acute anemia ongoing (n = 600)
 - US: Acute anemia – cardiovascular surgery, ongoing (n = 290)
- Riboflavin-UVB
 - Phase 2: radiolabel post transfusion recovery and lifespan 21-day RBC
 - Phase 3: prevention of transfusion transmitted plasmodia by WB

Amustaline-GSH RBC Clinical Data

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Red blood cell concentrates treated with the amustaline (S-303) pathogen reduction system and stored for 35 days retain post-transfusion viability: results of a two-centre study

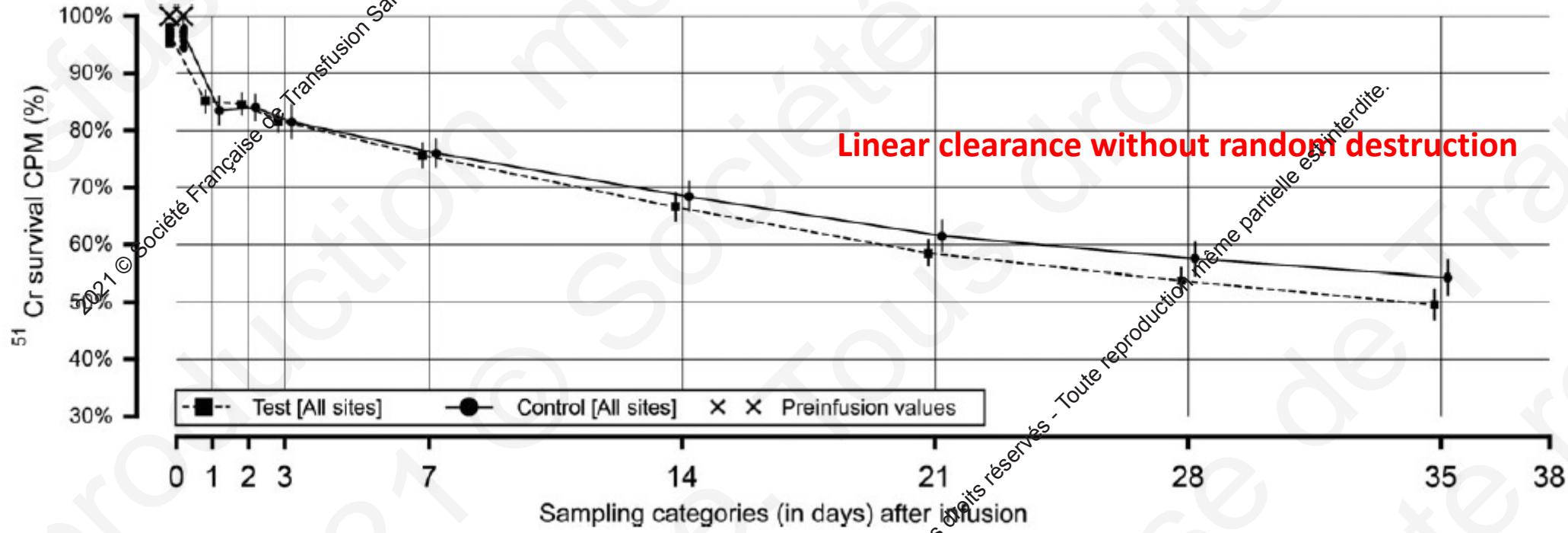
J. A. Canelas,¹ J. L. Gottschall,² N. Rugg,¹ S. Graminske,² M. A. Schott,³ A. North,³ N. Huang,³ N. Mufti,³ A. Erickson,³ S. Rico³ & L. Corash³

	Test mean (SD) (n = 26)	Control mean (SD) (n = 26)	Mean difference (Test–Control)	95% CI of Mean treatment Difference (Test–Control) ^a
24-h post-transfusion recovery (%)	83.2 (5.2)	84.9 (5.9)	-1.8	-3.6, 0.0
Life span (days)	62.8 (10.6) ^b	75.1 (13.7)	-12.3	-17.4, -7.2
T ₅₀ (days)	33.5 (7.1) ^b	39.7 (10.2)	-6.2	-9.7, -2.6
AUC _{0–last} (surviving cells × days)	22.6 (1.9)	23.1 (2.2)	-0.6	-1.4, 0.3

- No difference in 24-hour post transfusion recovery
- Difference in survival, but within physiologic ranges
- No difference in area under the curve (AUC)

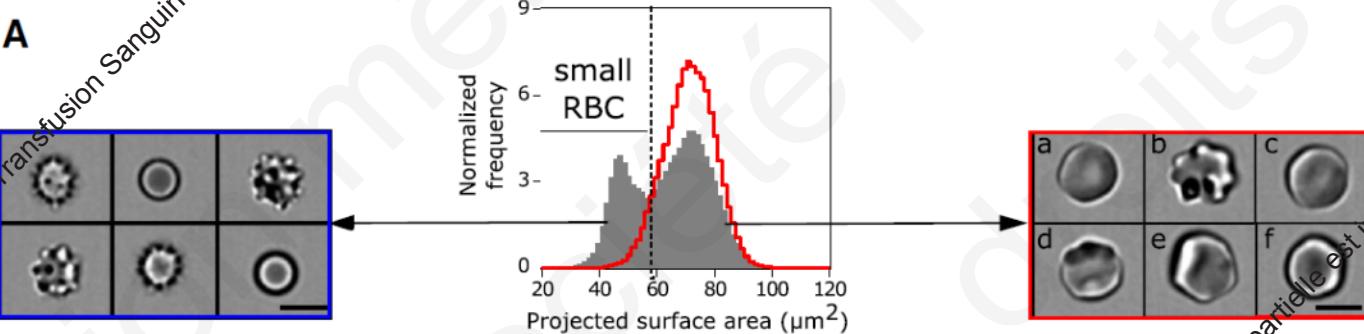
Red blood cell concentrates treated with the amustaline (S-303) pathogen reduction system and stored for 35 days retain post-transfusion viability: results of a two-centre study

J. A. Cancelas,¹ J. L. Gottschall,¹ N. Rugg,¹ S. Graminske,² M. A. Schott,³ A. North,³ N. Huang,³ N. Mufti,³ A. Erickson,³ S. Rico³ & L. Corash³



Linear clearance without random destruction

Spherocytic shift of red blood cells during storage provides a quantitative whole cell-based marker of the storage lesion

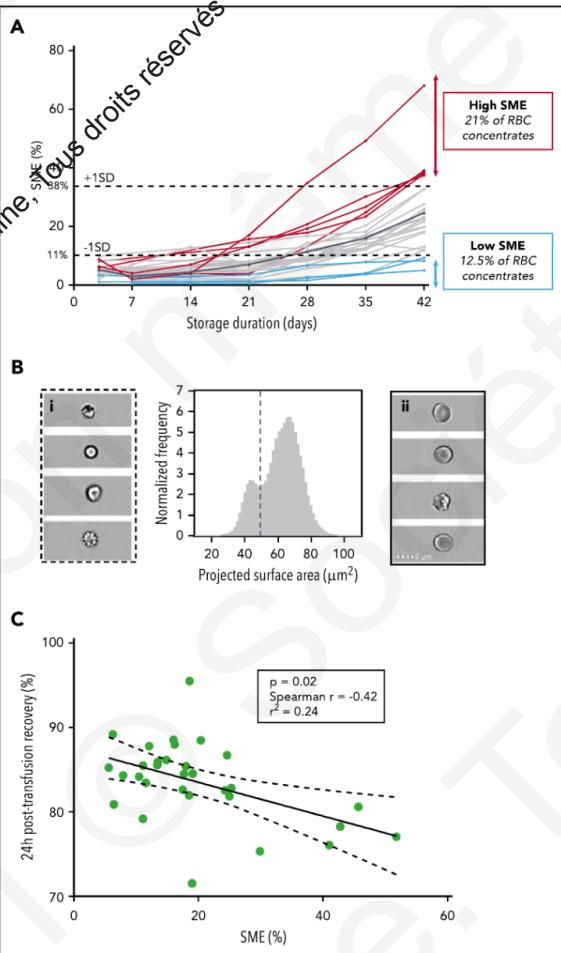


Rapid clearance of storage-induced microerythrocytes alters transfusion recovery

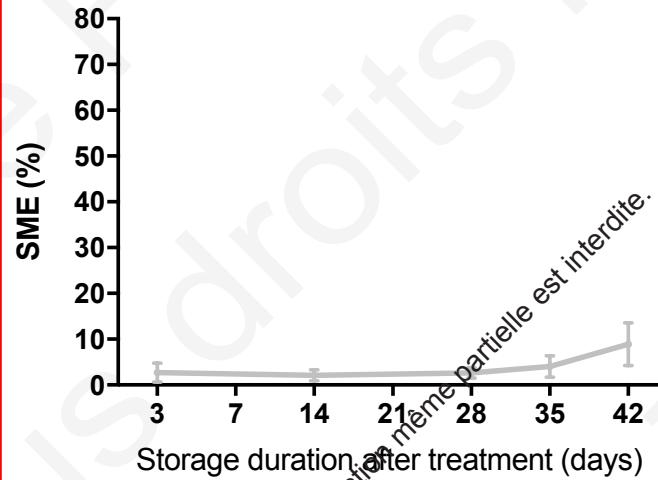
Camille Roussel,^{1-4,*} Alexandre Morel,^{1,2,5,*} Michaël Dussiot,^{1,2,*} Mickaël Marin,^{2,4} Martin Colard,^{1,2} Aurélie Fricot-Monsinjon,^{2,4} Anaïs Martinez,^{1,2} Charlotte Chambrion,^{2,4} Benoît Henry,^{2,4} Madeleine Casimir,^{1,2} Geoffroy Velle,^{2,4} Mallorie Dépond,^{2,4} Safi Dokmak,⁶ François Paye,⁷ Alain Sauvanet,⁶ Caroline Le Van Kim,^{2,4} Yves Colin,^{2,4} Sonia Georgeault,⁸ Philippe Roingeard,^{8,9} Steven L. Spitalnik,¹⁰ Papa Alioune Ndour,^{2,4} Olivier Hermine,^{1,2,5} Eldad A. Hod,¹⁰ Pierre A. Buffet,^{2,4,11,†} and Pascal Amireault^{1,4,†}

Rapid clearance of storage-induced microerythrocytes alters transfusion recovery

Figure 2. Proportion of SMEs at the end of storage correlates with 24-hour posttransfusion recovery in healthy human volunteers. (A) Quantification of SMEs upon storage of RBC concentrates in SAGM solution ($n = 24$) between days 3 and 42 (mean value in solid black line). Low (blue lines) and high proportions of SMEs (red lines) defined by proportions of SME $< -1\text{ SD}$ (11%) and $> +1\text{ SD}$ (38%) at the end of storage, respectively. (B) Representative normalized frequency plot for RBC concentrate at the end of storage in AS-3 showing a well-demarcated subpopulation of SMEs. Subpopulation of SMEs contains spherocytes, spherochinocytes, and type III echinocytes (i), whereas normal-sized RBCs (ii) contain discocytes and type I and II echinocytes. (C) Correlation between 24-hour posttransfusion recovery and proportions of SMEs quantified by imaging flow cytometry at the end of storage ($n = 31$; $P = .02$; Spearman $r = -0.42$; $r^2 = 0.24$).



Amustaline GSH RBCs show lower SME's than conventional stored RBC



Red blood cells treated with the amustaline (S-303) pathogen reduction system: a transfusion study in cardiac surgery

Veronika Brixner ,¹ Arnd-Holger Kiessling,² Katharina Madlener,³ Markus M. Müller,¹ Johannes Leibacher,¹ Sarah Dombos,¹ Iuliia Weber,¹ Hans-Ulrich Pfeiffer,¹ Christof Geisen,¹ Michael Schmidt,¹ Reinhard Henschler,^{4,5} Anne North,⁶ Norman Huang,⁶ Nina Mufti,⁶ Anna Erickson,⁶ Christine Ernst,⁶ Salvador Rico,⁶ Richard J. Benjamin,⁶ Laurence M. Corash,⁶ and Erhard Seifried¹

- Population: Elective adult cardiovascular surgery patients
- Intervention: Amustaline-GSH RBC Concentrates in SAGM
- Comparison: Conventional RBC Concentrates in SAGM
- Primary Outcome: Hemoglobin content of RBC Concentrates
- Timing: 7 days of RBC transfusion support, immune surveillance through day 90

In Vitro Efficacy

Parameter	TEST (n)	CONTROL (n)	Treatment Difference - P
Hemoglobin Content (g)	53.6 ± 5.6 (389)	56.3 ± 6.0 (365)	-2.6, -1.9: p < 0.001
Hemoglobin EOS QC (g)	53.1 ± 5.7 (301)	55.8 ± 5.9 (261)	-2.8, -1.92: p < 0.001
Hemolysis EOS QC (%)	0.28 ± 0.12 (301)	0.35 ± 0.16 (261)	-0.09, -0.04: p < 0.001
ATP EOS (umol/g)	2.8 ± 0.9 (294)	2.4 ± 0.7 (262)	0.33, 0.59: p < 0.001
Plasma free HB EOS (g/dL)	1.42 ± 0.64 (263)	1.79 ± 0.88 (235)	-0.49, -0.23: p < 0.001
Plasma Protein (mg/dL)	68.0 ± 25.6 (301)	228.8 ± 34.7 (298)	-166, 156: p < 0.001

Clinical Outcomes

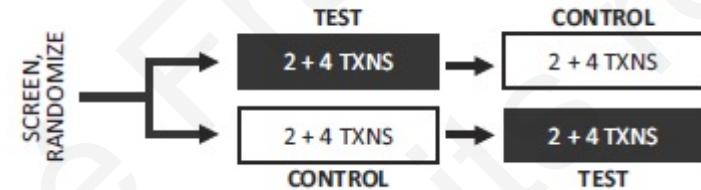
OUTCOME	TEST (25)	CONTROL (26)	P-VALUE
Renal Failure	5 (20%)	3 (11.5%)	0.412
Hepatic Failure	1 (4%)	0	0.371
6 MWT – day 1 (m)	44.8	53.1	0.472
6 MWT – day 13 (m)	95.5	97.7	0.626

Yesim Aydinok,¹ Antonio Piga,²
Raffaella Origa,³ Nina Mufti,⁴ Anna
Erickson,⁴ Anne North,⁴ Katie
Waldhausen,⁴ Christine Ernst,⁴ Jin-Syng
Lin,⁴ Norman Huang,⁴ Richard J.
Benjamin⁴ and Laurence Corash⁴ 

Amustaline-glutathione pathogen-reduced red blood cell concentrates for transfusion-dependent thalassaemia

STUDY DESIGN: Randomized Crossover 12 transfusions 80 Patients TDT

- Population: Transfusion Dependent Thalassemia
- Intervention: Amustaline-GSH RBC
- Comparison: Conventional RBC
- Outcome: Hemoglobin Use Over 6 months
- Timing: 6 Months Transfusion Support



VARIABLE HEMOGLOBIN CONTENT OF RBC COMPONENTS

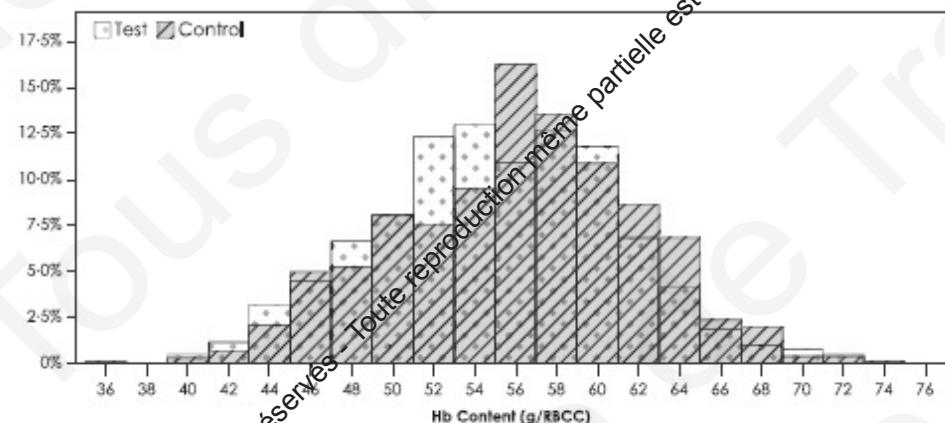


Fig 3. Haemoglobin content of test and control RBCC. The proportional (%) distributions of total haemoglobin (Hb) content (g) for 1024 Test (dots) and 1008 Control (diagonal) red blood cell concentrates (RBCC) are shown.

Primary Endpoint: Hemoglobin Use per Day per Kg

1 g of hemoglobin = 3.7 mg Fe

Hemoglobin Use of A-GSH RBC was not-inferior

Haemoglobin use (g/kg/day) for the ITT populations in the efficacy evaluation period (transfusions 3, 4, 5 and 6).*

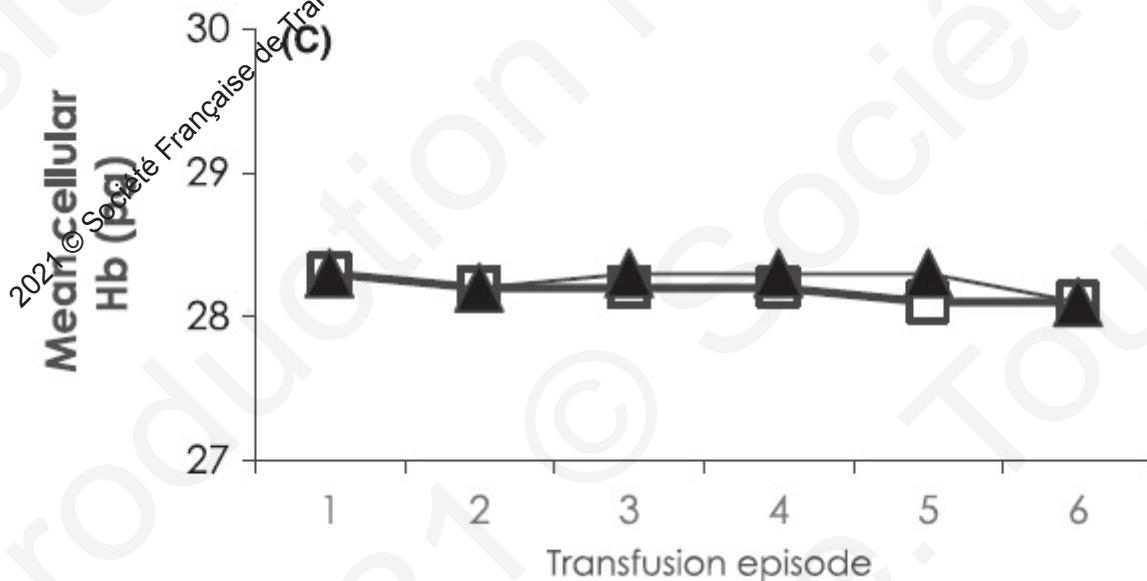
	Non-splenectomized		Splenectomized		ITT population§§		<i>P</i>
	Test	Control	Test	Control	Test	Control	
ITT (n)	40	40	40	40	80	80	
Dose [†]	471 (8)	461 (96)	451 (47)	461 (59)	461 (67)	461 (79)	0.950
Interval [†]	17.7 (3.0)	18.4 (3.4)	20.7 (3.5)	21.0 (3.5)	19.2 (3.6)	19.7 (3.8)	0.032
Hb use [†]	0.135 (0.03)	0.130 (0.03)	0.091 (0.03)	0.092 (0.03)	0.113 (0.04)	0.111 (0.04)	0.373

Parameter	Test (n = 1024)	Control (n = 1008)	Difference (CI) [†]	<i>P</i> [‡]
RBCC volume (ml)	271.4 (19.0)	278.9 (22.2)	-7.5 (-9.3, 5.7)	<0.001
RBCC Hct (%)	60.5 (2.4)	59.0 (2.8)	1.5 (1.7, 1.7)	<0.001
RBCC Hb content (g)	54.6 (5.9)	55.6 (5.9)	-1.0 (-1.5, -0.5)	<0.001

MCH as an Index of RBC Production Suppression

Decrease Erythroid Hyperplasia

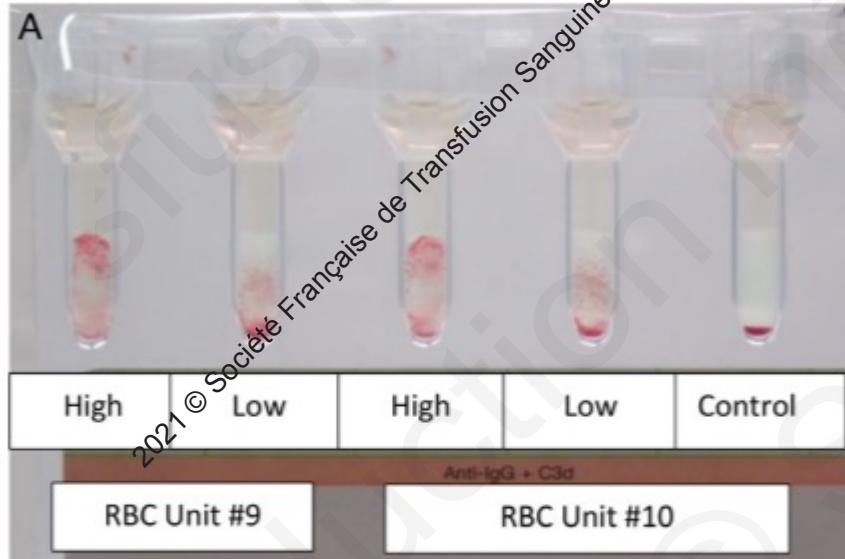
- MCH of Endogenous Thalassemia RBCs is < 25 pg
- Endogenous RBC Production was effectively suppressed



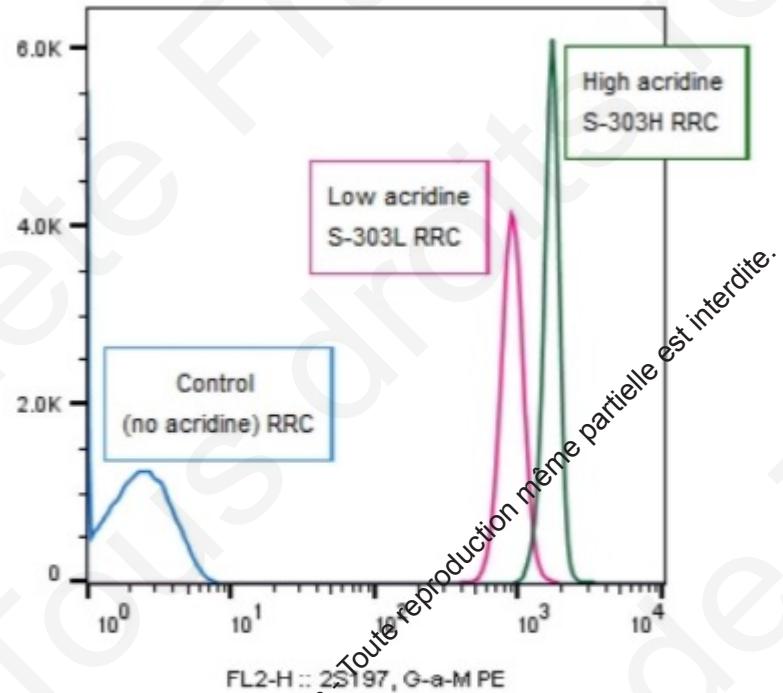
Pre-transfusion patient mean corpuscular haemoglobin (MCH, pg) are indicated for all Test (\blacktriangle) and Control (\square)

Prevalence of natural and acquired antibodies to amustaline/glutathione pathogen reduced red blood cells

Christof Geisen¹  | Anne North² | Lisa Becker¹ | Veronika Brixner¹  |
Melissa von Goetz² | Laurence Corash²  | Richard J. Benjamin²  |
Nina Mufti² | Erhard Seifried¹ 



Cell panels with varying levels of S-303 adducts to detect antibodies to S-303 RBC



SAMPLE ID	PE Molecules/Cell
Control RRC	42
S-303L RRC	15643
S-303H RRC	29721

Patient	Reactivity, initial screen		Antibody class/subclass	Antibody titer		Specificity by inhibition assay
	S-303H RRC	S-303L RRC		S-303H RRC	S-303L RRC	
General hospitalized patients						
1	++	-	IgG, non G _{1,3} /non G ₄	4	neg	Acridine
2	-	++	IgG, non G _{1,3}	neg	8	Unidentified
3	++	-	IgG, non G _{1,3} /non G ₄ ^a	4	neg	Acridine
4	++++	+++	IgG, non G _{1,3} /non G ₄	8	8	Acridine
5	+	-	IgG, non G _{1,3}	1	neg	Acridine
6	+++	+	IgG, non G _{1,3} /non G ₄	4	neg	Acridine
7	++	+	IgG, non G _{1,3}	2	neg	Acridine
8	+/-	-	IgG, non G _{1,3}	2	neg	Acridine
9	+	++	IgG, non G _{1,3} /non G ₄	4	4	Acridine
10	+++	-	IgM	16	neg	Acridine
11	+/-	+	IgG, non G _{1,3} / possible G ₄	4	nt	Acridine
12	+/-	-	IgG, non G _{1,3} /non G ₄	4	neg	Acridine
Hemoglobinopathy patients						
13	++++	+++	possible IgM	8	neg	Acridine
14	+++	++	probable IgM	8	neg	Acridine
15	-	++	IgG, non G _{1,3}	neg	1	unidentified
16	++++	+++	IgG, non G _{1,3}	32	neg	acridine
			IgM	16	neg	
17	+/-	+++	IgG, non G _{1,3}	nd	nt	unidentified

- 10,721 sera from transfused hospitalized patients
- 998 Chronically transfused patients
- 17 positive sera detected. primarily non-IgG_{1,3} all low titer
- 0.15% incidence of native Abs
- Primarily anti-acridine specific

TABLE 3 Patient characteristics in clinical studies using the current generation of amustaline/GSH RBC concentrates

Diagnosis	STARS ⁶ Cardiac surgery	SPARC ⁵ Thalassemia	Total
Number screened for natural S-303 antibody	87	86	173
Patients transfused with amustaline/GSH RBC Concentrates in test arm	25	81	106
Mean (range) exposure to amustaline/GSH RBC (# of concentrates)	2.9 (1-7)	12.5 (3-18)	(1-18)
Mean (SD) S-303 epitopes/RBC (n = 10) on Day 2 post manufacture ^a	9064 (1005)	13151 (919)	-
Mean (SD) S-303 epitopes/RBC (n = 12) on Day 35 post manufacture	5033 (841)	7035 (639)	-
Patients with natural S-303 specific antibodies at screening	0	0	0
Patients with S-303-specific antibodies at Day ~90 post transfusion	0	0	0

- Patients Exposed to S-303 RBC in Clinical Trials: No Immune Responses Observed
- 81 transfusion dependent thalassemia patients transfused for six months
- Large phase registry study for chronically transfused patients is expected as part of post marketing surveillance

Riboflavin UV Clinical Data

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Red blood cells derived from whole blood treated with riboflavin and ultraviolet light maintain adequate survival *in vivo* after 21 days of storage

Jose A. Cancelas,¹ Sherrill J. Slichter,^{2,3} Neeta Rugg,¹ P. Gayle Pratt,¹ Shawnagay Nestheide,¹ Jill Corsini,¹ Esther Pellham,² Marty Huntington,⁴ and Raymond P. Goodrich⁵

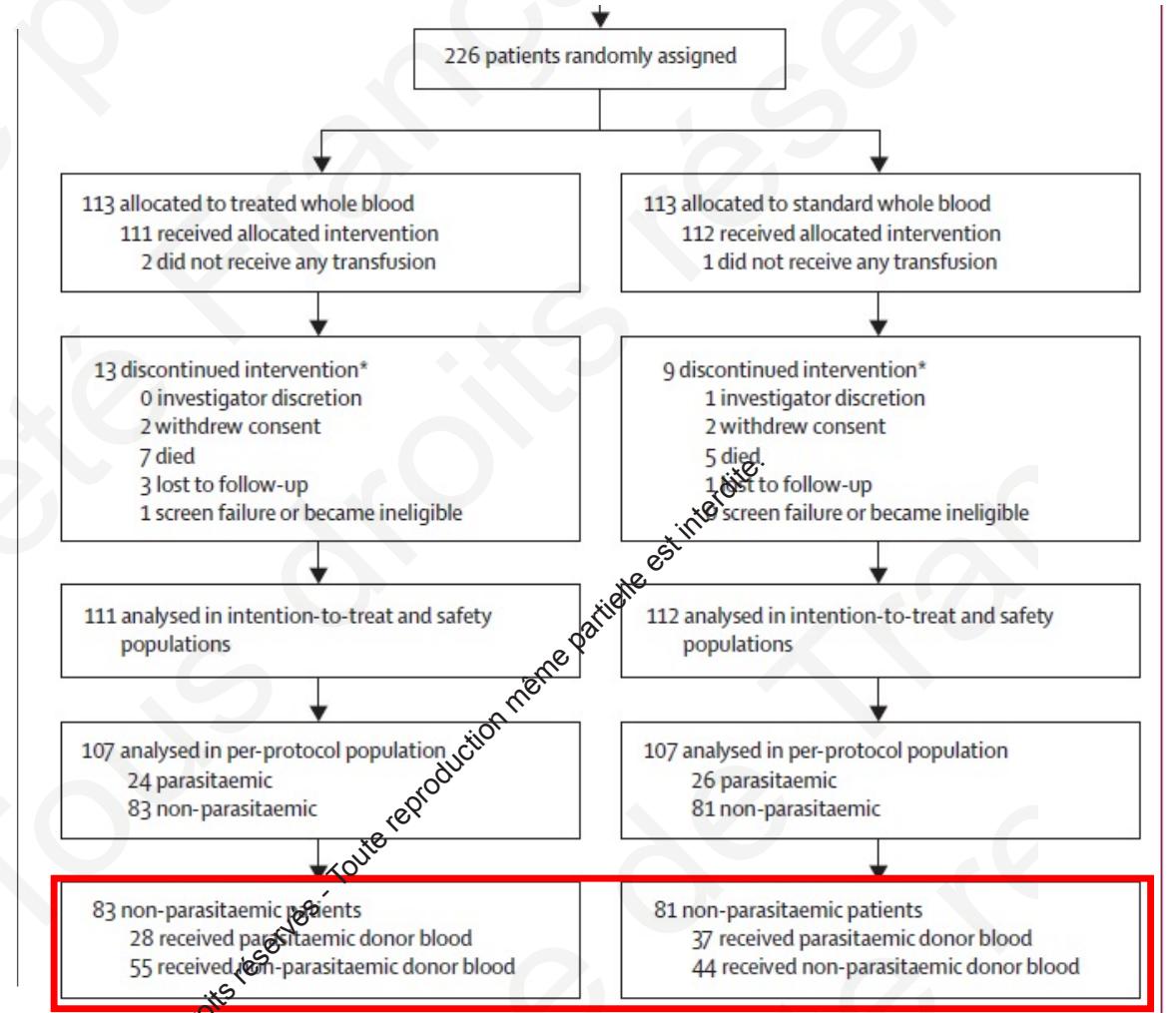
TABLE 1. 24-hour RBC recoveries and linear and exponential survivals

Collection order	All subjects		
	Mirasol treated	Untreated control	Difference
Number	24	24	24
24-hr RBC recovery (%)			
Mean (\pm SD)	82.5 (\pm 3.9)	91.7 (\pm 6.8)	-9.2 (\pm 6.6)
95% CI [†]	80.9 to 84.2	88.8 to 94.6	12.0 to -6.4
Subjects with RBC recovery $\geq 75\%^{\ddagger}$, n (%)	23 (95.8)	24 (100.0)	
RBC survival (days)			
Mean (\pm SD)	60.5 (\pm 5.6)	81.6 (\pm 15.5)	-21.1 (\pm 12.1)
95% CI [†]	58.1 to 62.8	75.0 to 88.1	-26.2 to -16.0
T ₅₀ (days)			
Mean (\pm SD)	22.6 (\pm 4.3)	35.8 (\pm 7.9)	-13.2 (\pm 7.0)
95% CI [†]	20.8 to 24.4	32.5 to 39.1	-16.2 to -10.3

Effect of Plasmodium inactivation in whole blood on the incidence of blood transfusion-transmitted malaria in endemic regions: the African Investigation of the Mirasol System (AIMS) randomised controlled trial

Jean-Pierre Allain, Alex K Owusu-Ofori, Sonny Michael Assennato, Susanne Marquie, Raymond P Goodrich, Shirley Owusu-Ofori

- Population: Plasmodia negative + Anemia
- Intervention: Riboflavin-UV WB Plasmodia + Donors
- Comparison: Conventional WB Plasmodia + Donors
- Outcome: Transfusion Transmitted Plasmodia
- Timing: Transfusion with 28 day follow up
- Design: Randomized, Controlled



Prevention of Transfusion Transmitted Malaria

	Parasitaemic whole blood transfused (exposed patients; n=65)*		Non-parasitaemic whole blood transfused (non-exposed patients; n=99)*		p value
	Treated patients (n=28)	Untreated patients (n=37)	Treated patients (n=54)	Untreated patients (n=45)	
Transfusion-transmitted malaria (≥2 parasitaemic samples, with allelic matching criteria)	1 (3·6%; 95% CI 0·1-28·2)	8 (21·6%†; 95% CI 9·8-38·2)	NA	NA	0·039‡
Transfusion-transmitted malaria (≥2 parasitaemic samples, no allelic matching criteria)	3 (10·7%; 95% CI 2·3-28·3)	13 (35·1%†; 95% CI 20·2-52·5)	1	0	0·024‡
Single parasitaemic samples	11	10	3	4	0·0049§
No parasitaemic samples	14	14	50	41	<0·0001§

The results of this primary endpoint analysis are for the total of 164 non-parasitaemic patients at day 0. NA=not applicable. *37 (56%) of 65 samples from exposed patients were parasitaemic post-transfusion vs eight (8%) of 99 in non-exposed patients ($p<0·0001$ for comparison between exposed and non-exposed patients receiving treated or untreated whole blood). †Absolute risk difference estimate: -18·1% (95% CI -33·0 to -3·1) for transfusion-transmitted malaria defined with allelic matching and -24·4% (-43·6 to -5·2) for transfusion-transmitted malaria defined without allelic matching. ‡p value for comparison between exposed patients transfused with treated vs untreated whole blood. §p value for exposed vs non-exposed patients receiving treated or untreated whole blood.

Table 2: Distribution of parasitaemic samples in exposed and non-exposed patients receiving treated or untreated whole blood

The incidence of TT Malaria was reduced by Riboflavin-UVB Treatment of Whole Blood

Summary and Conclusions

- Riboflavin-UV RBC is in clinical development, no ongoing trials at present
- Amustaline-GSH RBC completed clinical trials for CE mark registration
 - Dossier submitted to TUV and Competent Authority for MDR review
 - Initial indication is for anemia due to ineffective RBC production
 - Pathogen and leukocyte inactivation are effective
 - Post-transfusion RBC viability is within therapeutic range for 35-day RBC
 - RBC components contain adequate levels of hemoglobin
 - Amustaline-GSH RBC were non-inferior for support of transfusion dependent thalassemia in a blinded RCT
 - Immune responses are infrequent, and thus far not physiologically active
 - Post marketing immune surveillance and registry are required

THANK YOU, TOGETHER WE CAN MAKE A DIFFERENCE
EFS is an important collaborative partner

