

Bacilles à Gram négatif de résistance émergentes aux antibiotiques : Nouveaux antibiotiques et nouvelles stratégies thérapeutiques

Pr. Tristan Ferry, MD, PhD

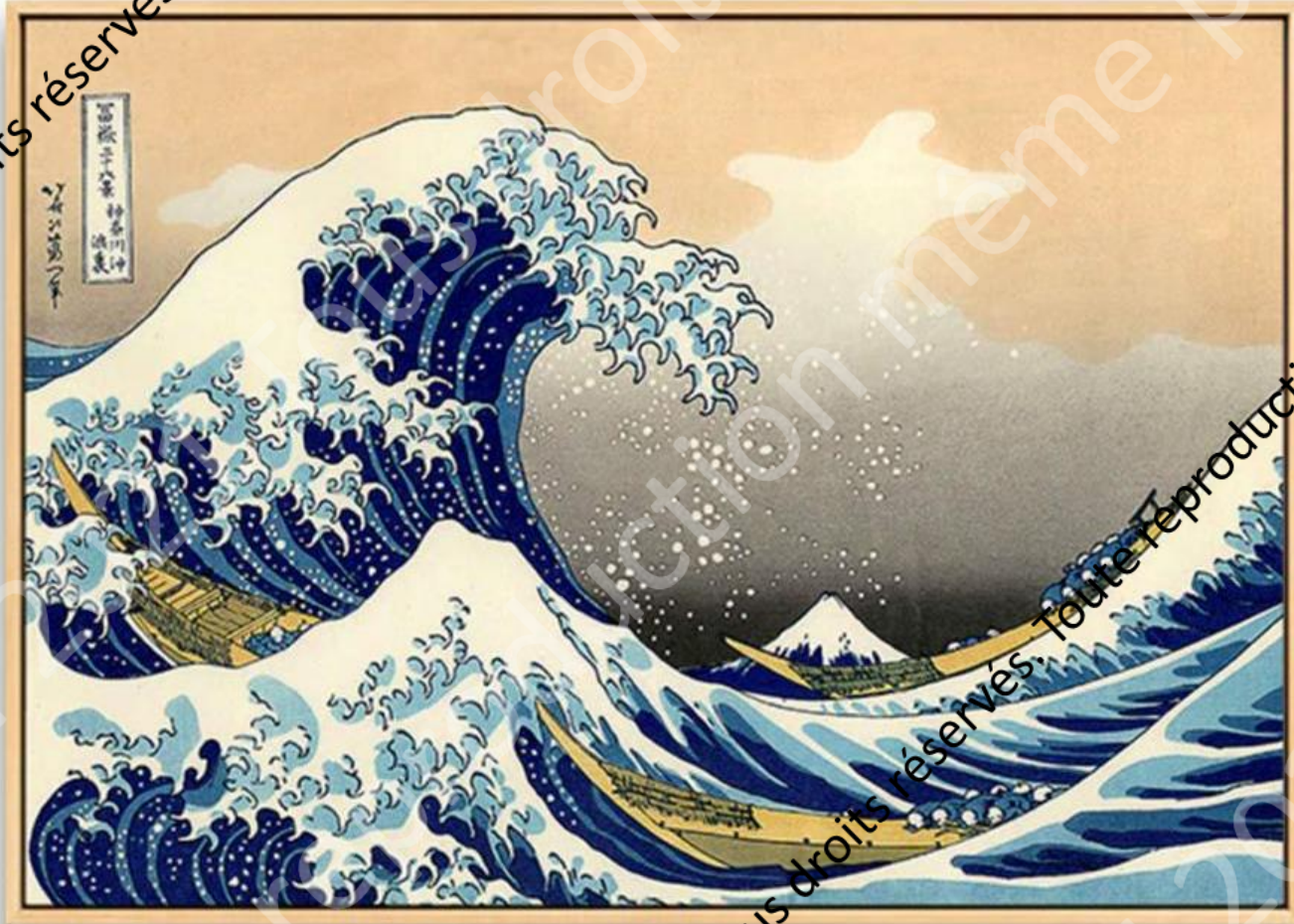
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Regional referral center for the management of complex bone and joint infection
Centre de Référence des IOA complexes de Lyon (CRIOAc Lyon)





Hokusai, 1931

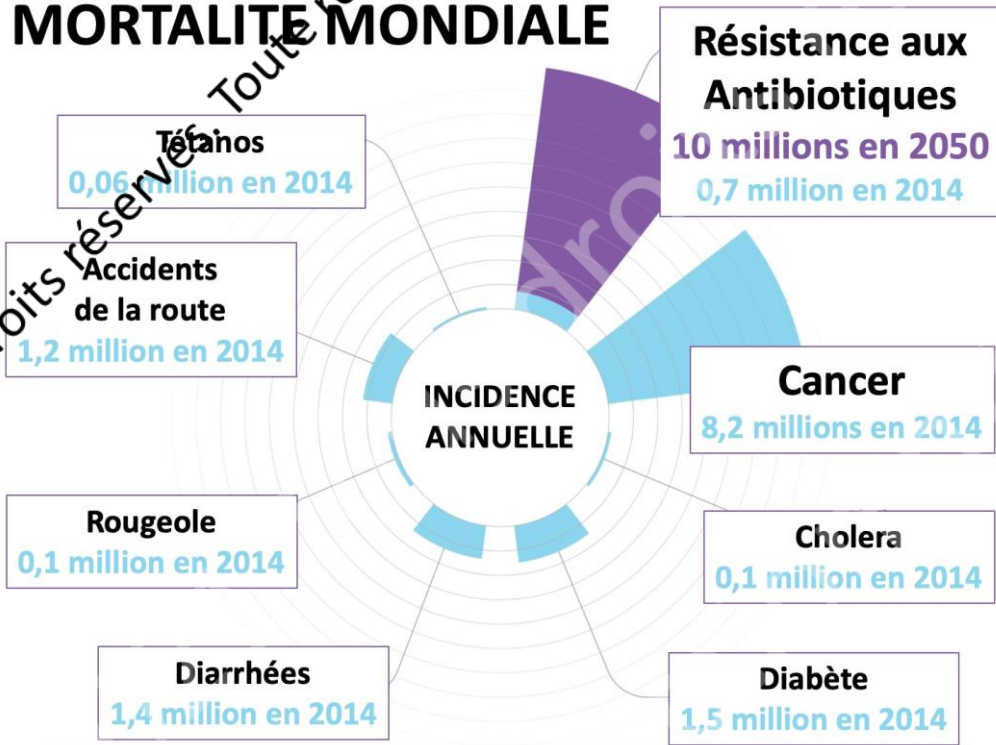
Antimicrobial resistance
is, indeed,
a slow motion tsunami



World Health
Organization

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MORTALITE MONDIALE



Data and statistics

33,000

The number of people who die every year in EU/EEA countries due to an infection with a resistant bacterial strain. Estimate for the whole WHO European Region is currently not available.



World Health Organization
REGIONAL OFFICE FOR Europe

WHO publishes list of bacteria for which new antibiotics are urgently needed

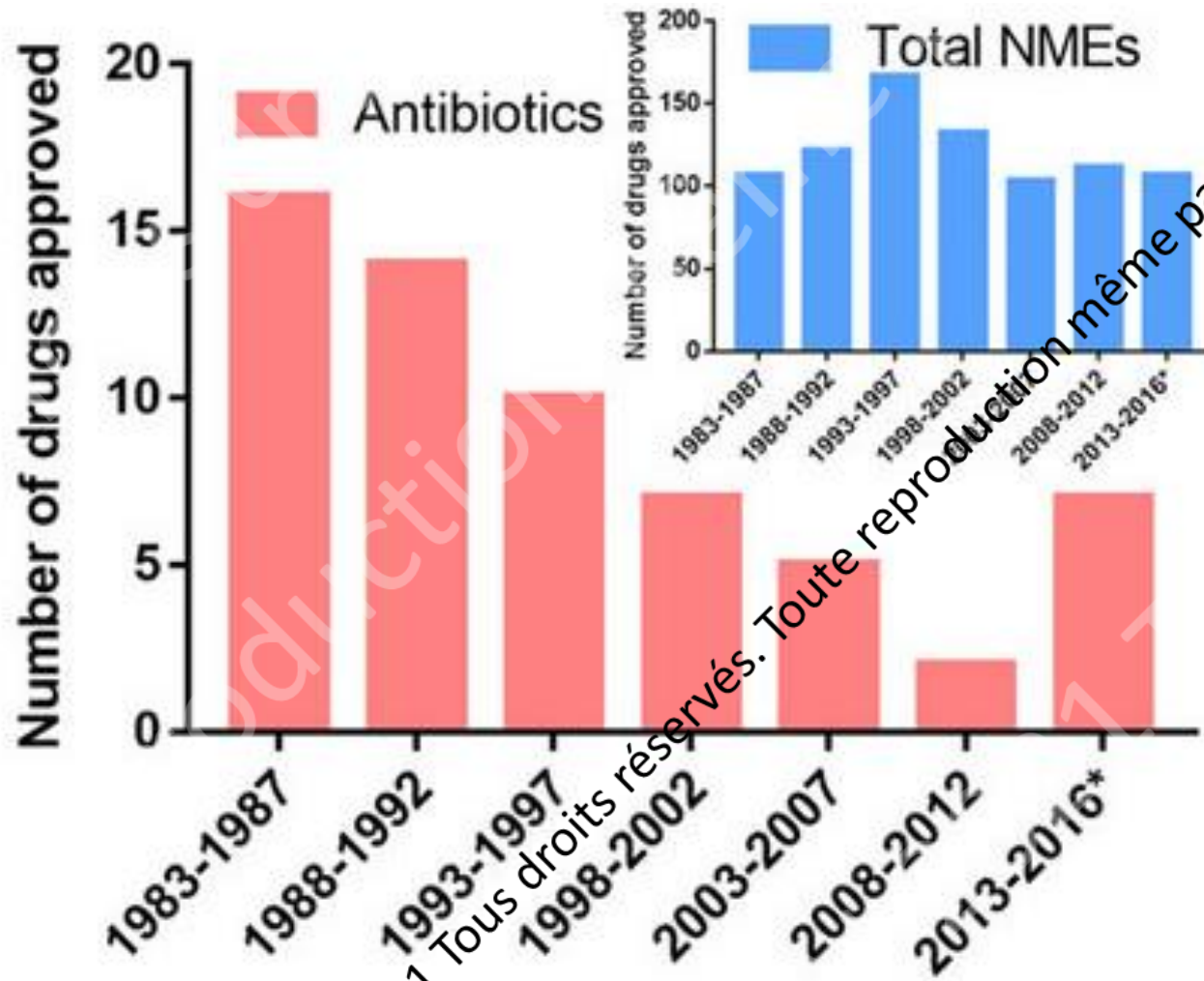
Priority 1: CRITICAL

Acinetobacter baumannii, carbapenem-resistant
Pseudomonas aeruginosa, carbapenem-resistant
Enterobacteriaceae, carbapenem-resistant, ESBL-producing

Priority 2: HIGH

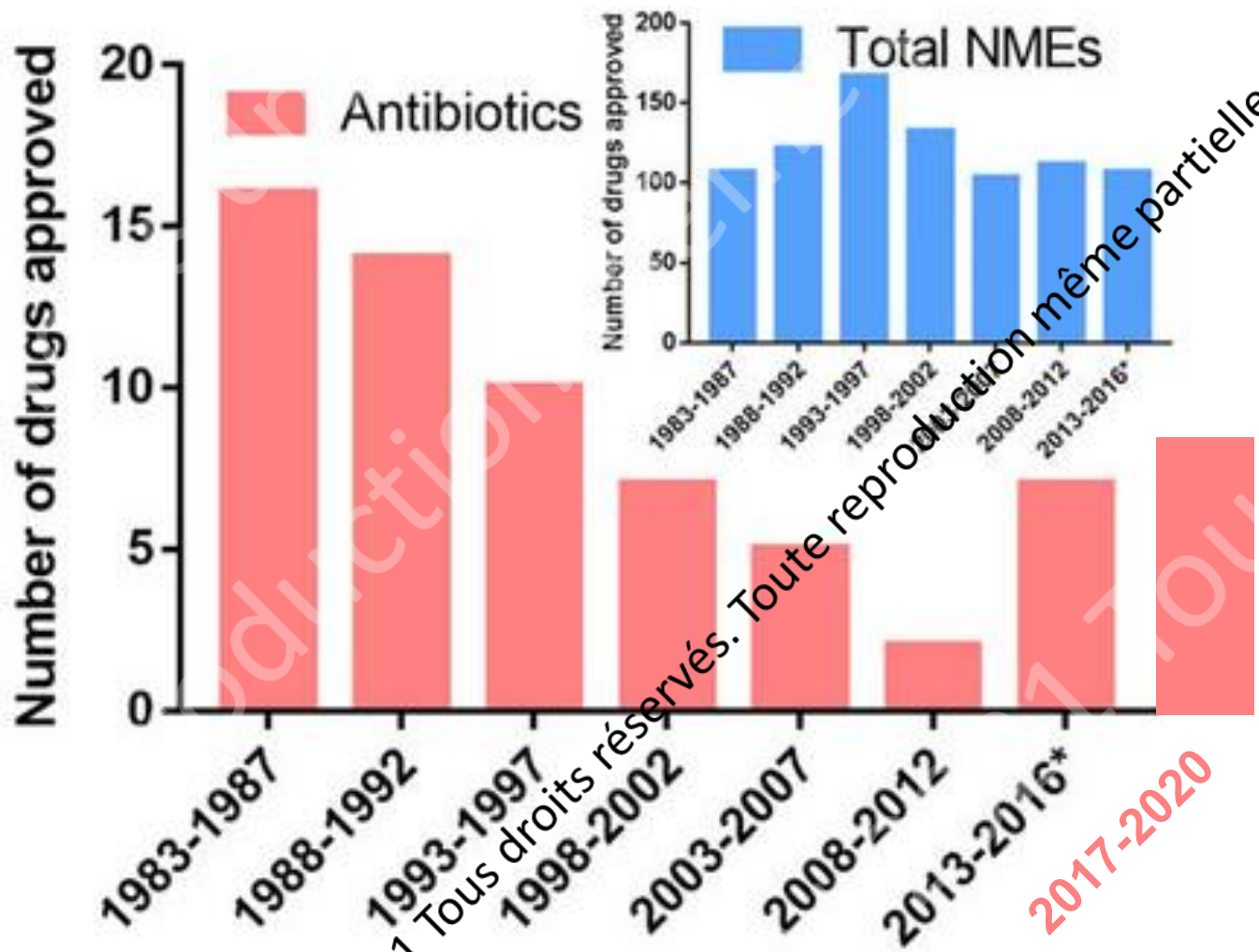
Enterococcus faecium, vancomycin-resistant
Staphylococcus aureus, methicillin-resistant, vancomycin intermediate and resistant
Helicobacter pylori, clarithromycin-resistant
Campylobacter spp., fluoroquinolone-resistant
Salmonellae, fluoroquinolone-resistant
Neisseria gonorrhoeae, cephalosporin-resistant, fluoroquinolone-resistant

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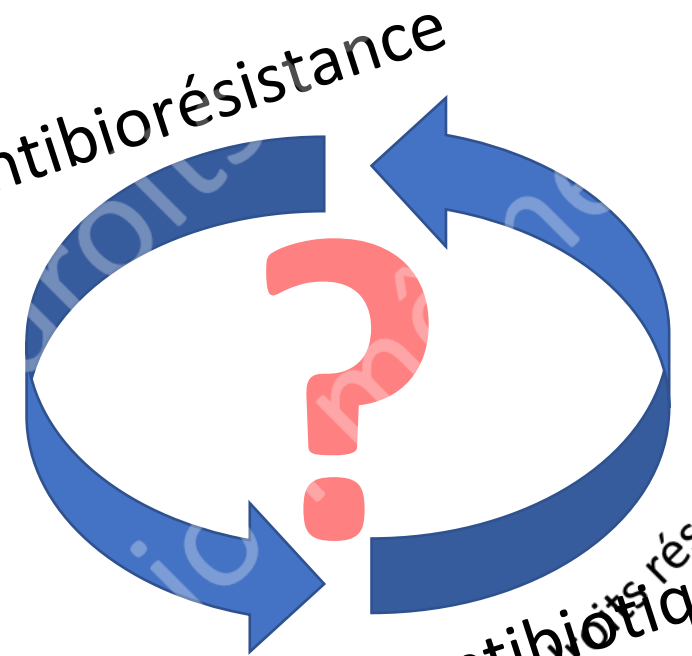


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British Journal of Pharmacology 2017

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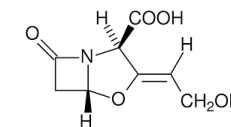
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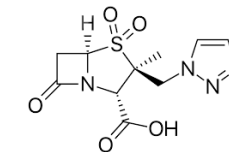
Les nouveaux inhibiteurs de β -lactamase

Inhibiteurs de β -lactamases	β -lactamases						
	BLSE	AmpC entéro	AmpC Pyo	Carbapénémase			
				KPC	MBL	OXA-48	OXA-23 OXA-58
Dérivés de β-lactamines							
Acide clavulanique							
Tazobactam							

Acide clavulanique



Tazobactam



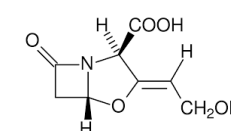
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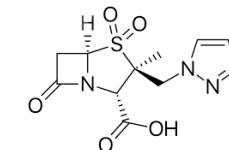
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	BLSE	AmpC entéro	AmpC Pyo	Carbapénémase			
				KPC	MBL	OXA-48	OXA-23 OXA-58
Dérivés de β-lactamines							
Acide clavulanique	Green	Red	Red	Light Red	Red	Red	Red
Tazobactam	Green	Red	Red	Light Red	Red	Red	Red
Diazabicyclooctanes							
Avibactam	Green	Green	Green	Green	Red	Green	Red
Relebactam	Green	Green	Green	Green	Red	Yellow	Red
Dérivés de l'acide boronique							
Vaborbactam	Green	Green	Yellow	Green	Red	Red	Red

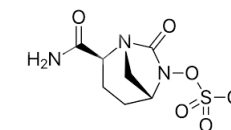
Acide clavulanique



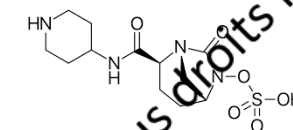
Tazobactam



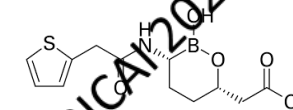
Avibactam



Relebactam



Vaborbactam

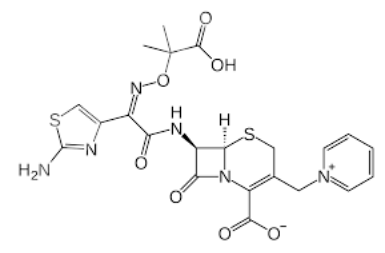


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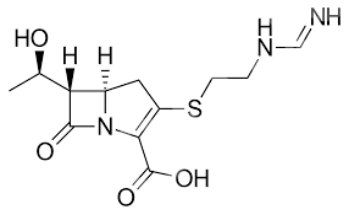
Anciens antibiotiques

Les nouveaux inhibiteurs de β -lactamase

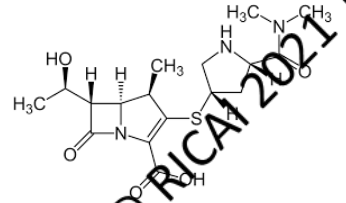
Ceftazidime



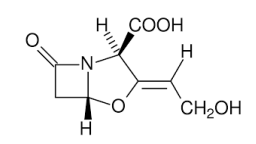
Imipénème cilastatine



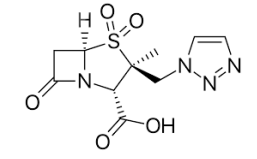
Méropénème



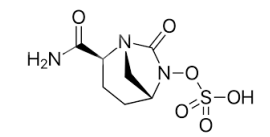
Acide clavulanique



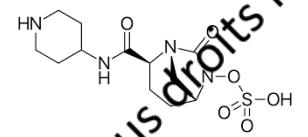
Tazobactam



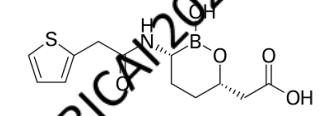
Avibactam



Relebactam



Vaborbactam

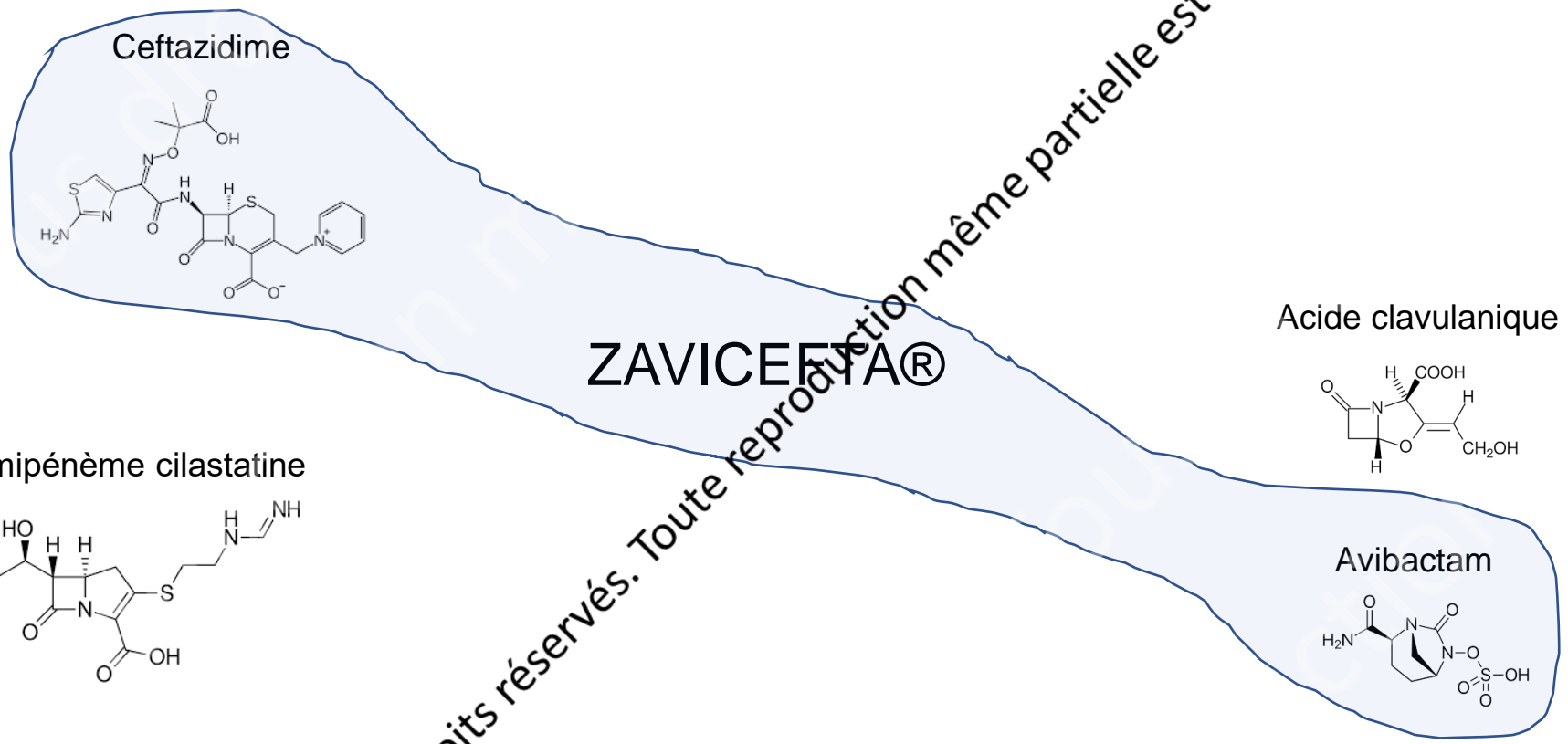


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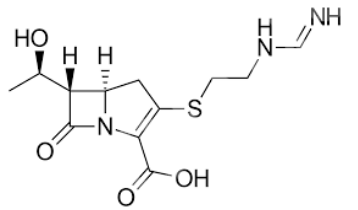
Anciens antibiotiques

Les nouveaux inhibiteurs de β -lactamase

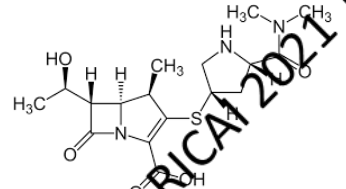


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Imipénème cilastatine



Méropénème



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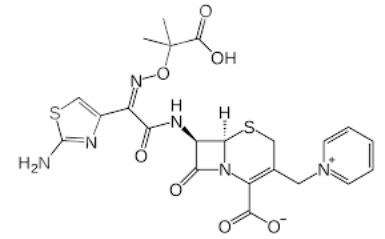
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Anciens antibiotiques

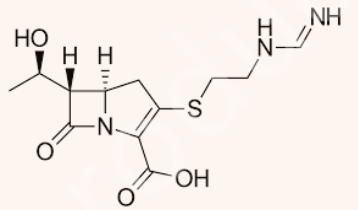
Les nouveaux inhibiteurs de β -lactamase

Ceftazidime



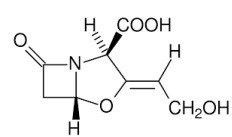
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Imipénème cilastatine

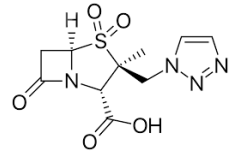


RECARBRIO®

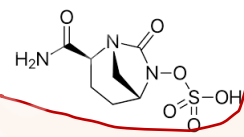
Acide clavulanique



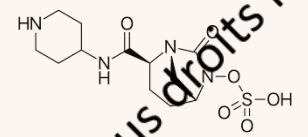
Tazobactam



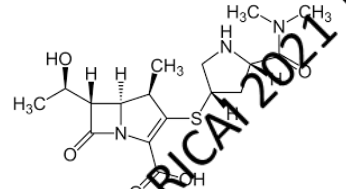
Avibactam



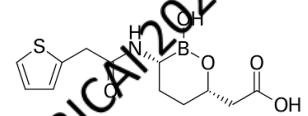
Relebactam



Méropénème



Vaborbactam



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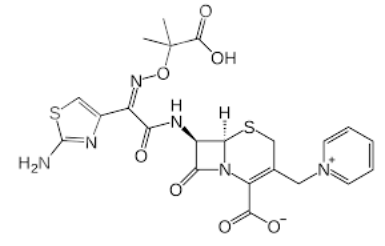
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Anciens antibiotiques

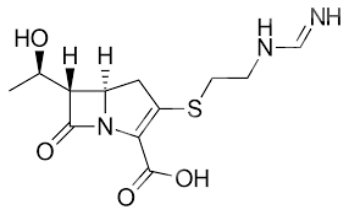
Les nouveaux inhibiteurs de β -lactamase

Ceftazidime



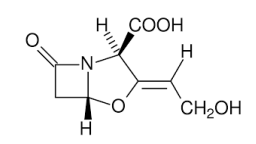
ZAVICEFYA®

Imipénème cilastatine

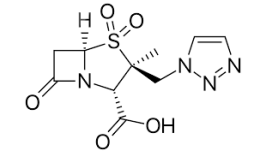


REC CARBRIO®

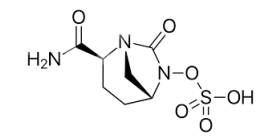
Acide clavulanique



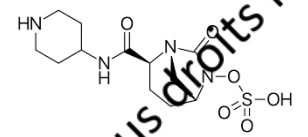
Tazobactam



Avibactam



Relebactam

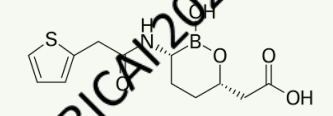


Méropénème



VABOREM®

Vaborbactam

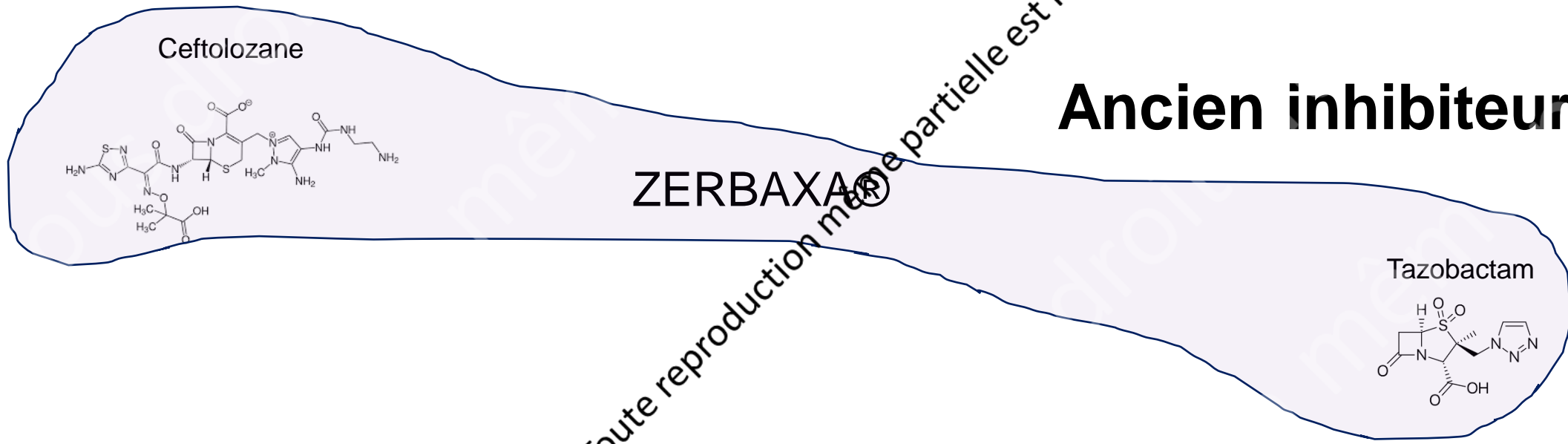


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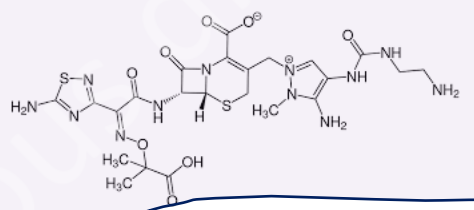
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Nouveaux antibiotiques

Ancien inhibiteur

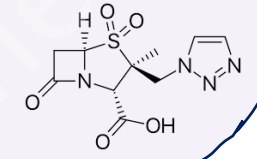


Ceftolozane

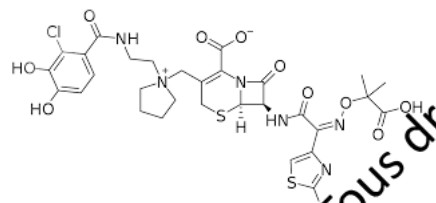


ZERBAXA®

Tazobactam



Cefiderocol



FETCROJA®

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Recommended antibiotic treatment options for carbapenem-resistant Enterobacterales (CRE), assuming *in vitro* susceptibility to agents in table, for URINARY TRACT INFECTIONS

Source of Infection	Preferred Treatment	Alternative Treatment (first-line options not available or tolerated)
Cystitis	Oral drug Single-dose of an aminoglycoside Meropenem ₁ (standard-infusion): only if ertapenem resistant, meropenem susceptible, AND carbapenemase testing results are either not available or negative.	Ceftazidime-avibactam Meropenem-vaborbactam Imipenem-cilastatin-relebactam Cefiderocol Colistin (only when no alternative options are available)
Pyelonephritis or cUTI ₂	Ceftazidime-avibactam Meropenem-vaborbactam Imipenem-cilastatin-relebactam Cefiderocol Meropenem ₁ (extended-infusion): only if ertapenem resistant, meropenem susceptible, AND carbapenemase testing results are either not available or negative.	Once-daily aminoglycosides

Recommended antibiotic treatment options for carbapenem-resistant Enterobacterales (CRE), assuming *in vitro* susceptibility to agents in table, for OTHER INFECTIONS

Source of Infection	Preferred Treatment	Alternative Treatment (first-line options not available or tolerated)
Infections outside of the urinary tract Resistant to ertapenem, susceptible to meropenem, AND carbapenemase testing results are either not available or negative	Meropenem ¹ (extended-infusion)	Ceftazidime-avibactam
Infections outside of the urinary tract Resistant to ertapenem, meropenem , AND carbapenemase testing results are either not available or negative	Ceftazidime-avibactam Meropenem-vaborbactam Imipenem-cilastatin-relebactam	Cefiderocol Tigecycline, eravacycline (IAI)
KPC identified (Or carbapenemase positive but identity of carbapenemase unknown ³)	Ceftazidime-avibactam Meropenem-vaborbactam Imipenem-cilastatin-relebactam	Cefiderocol Tigecycline, eravacycline (IAI)
Metallo-β-lactamase (i.e., NDM, VIM, or IMP) carbapenemase identified	Ceftazidime-avibactam + aztreonam Cefiderocol	Tigecycline, eravacycline (IAI)
OXA-48-like carbapenemase identified	Ceftazidime-avibactam	Cefiderocol Tigecycline, eravacycline (IAI)

Recommended antibiotic treatment options for difficult-to-treat (DTR) *Pseudomonas aeruginosa*, assuming *in vitro* susceptibility to agents in table

Source of Infection	Preferred Treatment	Alternative Treatment (when first-line options not available/tolerated)
Cystitis	<p>Ceftolozane-tazobactam Ceftazidime- avibactam Imipenem-cilastatin-relebactam Cefiderocol</p> <p>or a single-dose of an aminoglycoside</p>	Colistin
Pyelonephritis or cUTI ₁	<p>Ceftolozane-tazobactam Ceftazidime- avibactam Imipenem-cilastatin-relebactam Cefiderocol</p>	Once-daily aminoglycosides
Infections outside of the urinary tract	<p>Ceftolozane-tazobactam Ceftazidime- avibactam Imipenem-cilastatin-relebactam</p>	<p>Cefiderocol</p> <p>Aminoglycoside monotherapy: limited to uncomplicated bloodstream infections with complete source control₂</p>

Cas clinique

- Patiente de 70 ans, 45 kg
- Cancer du sein en 2011, HTA
- Fracture humérale gauche lors d'un voyage en Ouzbékistan (AVP)
- Ostéosynthèse en Turquie
- Séjour en réanimation avec pneumonie nosocomiale (traitement par colistine-tigécycline)

Rapatriment à Lyon 1 mois plus tard

- Ecoulement de la cicatrice humérale
- Reprise chirurgicale, ablation du matériel
- Absès, pus franc, fracture encore non consolidée
- Antibiothérapie post-opératoire par céfépime-vancomycine

Au lit

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	Klebsiella pneumoniae
	CMI (mg/l)
Ampicilline	R
Amoxicilline	R
Amoxicilline + Ac. Clavulanique	R
Ticarcilline	R
Ticarcilline + Ac. Clav	R
Penicilline	R
Pipéracilline	R
Pipéracilline + Tazobactam	R
Céfoxitine	R
Céfotaxime	R
Ceftazidime	R
Céfépime	R
Aztréonam	R
Ertapénème	R
Imipénème	S
Meropenème	R
Gentamicine	R
Tobramycine	R
Amikacine	R
Tigécycline	S
Norfloxacine	R
Ciprofloxacine	R
Moxifloxacine	R
Acide Nalidixique	R
Cotrimoxazole	R

R

Fosfomycine (Etest)	S CMI : 64
Aztreonam (Etest)	R CMI : > 256
Cefepime (Etest)	R CMI : 64
Imipenème (Etest)	S CMI : 0.380
Meropenème (Etest)	R CMI : > 32
Tobramycine (Etest)	S CMI : 0.250
Tigécycline (Etest)	S CMI : 0.250
Ceftolozane-tazobactam (Etest)	R CMI : > 256
Ceftazidime-Avibactam (Etest)	R CMI : > 256
Colistine (UMIC)	R CMI : 64

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CMI par E-test Aztréonam + Ceftazidime/Avibactam : 0.19 mg/L.

Adaptation thérapeutique

- Aztréonam 2g/8h – Ceftazidime/avibactam 2g/8h – Daptomycine
- Aztréonam 2g/8h – Ceftazidime/avibactam 2g/8h – Fosfomycine 3g en 3h/8h



Adaptation thérapeutique

- Aztréonam 2g/8h – Ceftazidime/avibactam 2g/8h – Daptomycine
 - Aztréonam 2g/8h – Ceftazidime/avibactam 2g/8h – Fosfomycine 3g en 3h/8h
-
- Aztréonam 3g dans un diffuseur sur 12h – Ceftazidime/avibactam 3g dans un diffuseur sur 12h



Aztréonam + Ceftazidime/avibactam ?

Can Ceftazidime-Avibactam and Aztreonam Overcome β -Lactam Resistance Conferred by Metallo- β -Lactamases in *Enterobacteriaceae*?

Steven Marshall,^a Andrea M. Hujer,^{a,b} Laura J. Rojas,^{a,b,c} Krisztina M. Papp-Wallace,^a Romney M. Humphries,^d Brad Spellberg,^e Kristine M. Hujer,^{a,b} Emma K. Marshall,^a Susan D. Rudin,^{a,b} Federico Perez,^f Brigid M. Wilson,^a Ronald B. Wasserman,^f Linda Chikowski,^g David L. Pater,^h Alejandro J. Vila,ⁱ David van Duin,^j Barry N. Kreiswirth,^k Henry F. Chambers,^l Vance G. Fowler, Jr.,^m Michael R. Jacobs,ⁿ Mark E. Pulse,^o William J. Weiss,^c Robert A. Bonomo^{a,b,c,p}

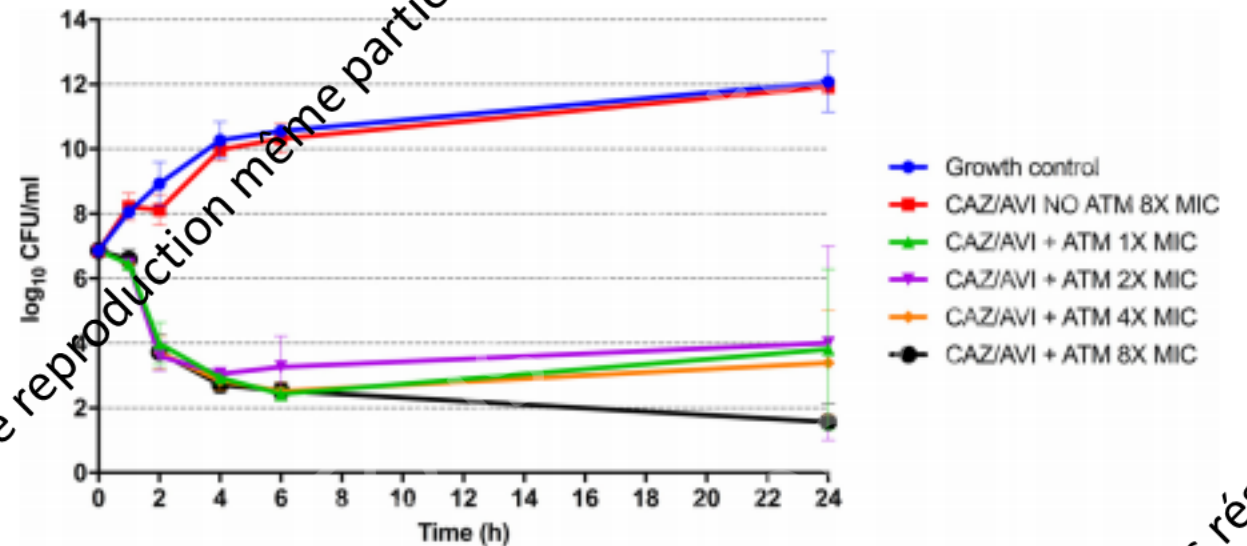


FIG 2 Time-kill curve for *K. pneumoniae* isolate 1.41. ATM concentrations were held constant at 8 $\mu\text{g}/\text{ml}$ for all combinations, with two exceptions: (i) the growth control (no antibiotics added) and (ii) CAZ/AVI with no ATM at 8 \times the MIC. Various ceftazidime-avibactam (CAZ-AVI) concentrations were added corresponding to 1 \times (1 $\mu\text{g}/\text{ml}$ CAZ plus 0.25 $\mu\text{g}/\text{ml}$ AVI), 2 \times (2 $\mu\text{g}/\text{ml}$ CAZ plus 0.5 $\mu\text{g}/\text{ml}$ AVI), 4 \times (4 $\mu\text{g}/\text{ml}$ CAZ plus 1 $\mu\text{g}/\text{ml}$ AVI), and 8 \times (8 $\mu\text{g}/\text{ml}$ CAZ plus 2 $\mu\text{g}/\text{ml}$ AVI) the MIC of the combination CAZ/AVI plus ATM obtained by agar dilution (1 $\mu\text{g}/\text{ml}$). Three replicates were conducted for each of the conditions reported in the time-kill assay.

K. Pneumoniae NDM-1, CTX-M-15, OHA, SHV, TEM

Aztréonam +

Aztreonam plus Clavulanate, Tazobactam Treatment of Infections Caused by Producing Gram-Negative Bacteria

Cécile Emeraud,^{a,b,c,d} Lelia Escaut,^a Athénaïs Boucly,^{d,f,g} Nicolas Fortin
Laurent Dortet^{a,b,c,d}



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TABLE 2 MICs and categorization according to CLSI breakpoints for antimicrobials from producing *Enterobacteriaceae*, MBL-producing *P. aeruginosa*, and *S. maltophilia*

<i>Enterobacteriaceae</i> sp.	β-Lactamases	MICs (mg/liter) by treatment ^a						
		ATM	CZA	C/T	ATM+ CZA	ATM+ C/T	ATM+ AMC	
<i>E. coli</i>	NDM-1 + OXA-1 + OXA-10 + CMY-16 + TEM-1	32	>256	>256	16	0.125	24	8
<i>E. coli</i>	NDM-1 + CTX-M-15 + TEM-1	>256	>256	>256	12	1	>256	2
<i>E. coli</i>	NDM-1 + OXA-1 + OXA-2 + CTX-M-15 + TEM-1	>256	>256	>256	24	2	>256	8
<i>E. coli</i>	NDM-1 + CTX-M-15 + TEM-1	>256	>256	>256	32	6	>256	8
<i>E. coli</i>	NDM-4 + CTX-M-15 + OXA-1	>256	>256	>256	96	6	>256	4
<i>E. coli</i>	NDM-4 + CTX-M-15 + CMY-6	>256	>256	>256	>256	6	>256	24
<i>E. coli</i>	NDM-5 + TEM-1 + CTX-M-15	>256	>256	>256	96	8	>256	64
<i>E. coli</i>	NDM-6 + CTX-M-15 + OXA-1	>256	>256	>256	16	1	>256	2
<i>E. coli</i>	NDM-7 + ESBL	>256	>256	>256	96	4	>256	32
<i>K. pneumoniae</i>	NDM-1 + CTX-M-15 + SHV-11 + OXA-1	>256	>256	>256	12	0.125	24	0.38
<i>K. pneumoniae</i>	NDM-1 + CTX-M-15 + CMY-4 + OXA-1	>256	>256	>256	32	0.75	>256	16
<i>K. pneumoniae</i>	NDM-1 + CTX-M-15 + OXA-1 + OXA-9 + TEM-1 + SHV-28 + SHV-11	>256	>256	>256	32	0.25	>256	3
<i>K. pneumoniae</i>	NDM-1 + OXA-1 + SHV-11	>256	>256	>256	12	0.047	0.094	0.094
<i>K. pneumoniae</i>	NDM-1 + OXA-1 + CTX-M-15 + TEM-1 + SHV-28 + OXA-9 + CMY-6	>256	>256	>256	16	0.047	3	0.25
<i>K. pneumoniae</i>	NDM-1 + TEM-1 + CTX-M-15 + SHV-12 + OXA-9	>256	>256	>256	12	0.125	96	1
<i>K. pneumoniae</i>	NDM-1 + TEM-1 + CTX-M-15 + SHV-12 + OXA-9	>256	>256	>256	12	0.125	96	0.5
<i>K. pneumoniae</i>	NDM-1 + TEM-1 + CTX-M-15 + SHV-11 + OXA-1	>256	>256	>256	12	0.064	8	0.38
<i>Salmonella enterica</i>	NDM-1 + CTX-M-15 + TEM-1 + OXA-1 + OXA-9 + OXA-10	>256	>256	>256	16	0.125	16	0.5
<i>E. coli</i>	VIM-1 + CTX-M-3	>256	>256	>256	16	0.125	24	0.5
<i>E. coli</i>	VIM-4 + ESBL	16	>256	>256	24	1.5	24	16
<i>K. pneumoniae</i>	VIM-1 + SHV-5	>256	>256	>256	>256	0.25	192	1.5
<i>K. pneumoniae</i>	VIM-1 + SHV-12	>256	>256	>256	16	0.125	4	0.25
<i>K. pneumoniae</i>	VIM-1 + ESBL	>256	>256	>256	>256	12	16	12
<i>K. pneumoniae</i>	VIM-1 + SHV-5	16	>256	>256	>256	6	12	32
<i>K. pneumoniae</i>	VIM-1 + TEM-1 + SHV-5	96	>256	>256	>256	96	64	48
<i>K. pneumoniae</i>	VIM-1 + SHV-5	>286	>256	>256	24	0.25	8	0.75
<i>K. pneumoniae</i>	VIM-1 + SHV-5	>256	>256	>256	12	0.125	2	0.38
<i>K. pneumoniae</i>	VIM-19 + CTX-M-3 + TEM-1 + SHV-1	6	32	>256	16	0.047	2	1.5
<i>Enterobacter cloacae</i>	VIM-1 + SHV-70	256	128	>256	48	0.094	0.25	0.19
<i>E. cloacae</i>	VIM-1 + CTX-M-15 + TEM-1 + SHV-31	64	>256	>256	64	1	64	32
<i>Citrobacter freundii</i>	VIM-2 + TEM-1 + ESBL	16	16	>256	32	0.25	2	24
<i>C. freundii</i>	VIM-2 + TEM-1 + OXA-9 + OXA-10	32	24	>256	32	1.5	16	24
<i>E. coli</i>	IMP-8 + SHV-12	128	>256	>256	24	0.19	2	0.38
<i>K. pneumoniae</i>	IMP-8 + SHV-12	>256	48	>256	12	0.094	32	0.25
<i>E. cloacae</i>	IMP-8 + SHV-12	12	>256	>256	24	0.032	0.064	0.094
<i>E. cloacae</i>	GIM-1 + ESBL	12	>256	48	24	0.5	8	16
<i>Enterobacter aerogenes</i>	TMB-1 + overexpressed Case ^b	64	64	32	32	0.5	12	12
<i>C. freundii</i>	TMB-1 + overexpressed Case	64	96	32	12	0.125	12	12
<i>K. pneumoniae</i>	NDM-1 + OXA-181 + SHV-11 + TEM-1 + CTX-M-15 + OXA-1	64	>256	>256	48	0.094	8	2
<i>K. pneumoniae</i>	NDM-1 + OXA-181 + SHV-27 + CTX-M-15 + TEM-1 + OXA-1	128	>256	>256	96	0.25	16	3
<i>K. pneumoniae</i>	NDM-1 + OXA-181 + SHV-11 + CTX-M-15 + OXA-1	256	>256	>256	>256	0.19	32	3
<i>K. pneumoniae</i>	NDM-1 + OXA-181 + SHV-11 + TEM-1 + CTX-M-15 + OXA-9	>256	>256	>256	>256	0.19	>256	12
<i>K. pneumoniae</i>	NDM-1 + OXA-181 + SHV-2 + CTX-M-15 + OXA-1	>256	>256	>256	32	0.125	32	1.5
<i>C. freundii</i>	NDM-1 + OXA-181 + OXA-1 + OXA-9 + OXA-10 + CTX-M-15 + TEM-1	>256	>256	>256	64	0.75	>256	12
<i>E. coli</i>	NDM-1 + OXA-48 + ESBL	32	>256	>256	48	0.094	12	8
<i>E. coli</i>	NDM-1 + OXA-48 + ESBL	>256	>256	>256	>256	0.75	>256	4
<i>E. coli</i>	NDM-1 + OXA-48 + ESBL	>256	>256	>256	>256	1	>256	4
<i>K. pneumoniae</i>	NDM-1 + OXA-232 + ESBL	64	>256	>256	>256	0.094	24	4
<i>E. coli</i>	NDM-1 + OXA-232 + ESBL	>256	>256	>256	>256	1	>256	64
<i>E. coli</i>	NDM-5 + OXA-232 + ESBL	>256	>256	>256	96	1	>256	64
<i>S. maltophilia</i>		>256	>256	>256	32	2	12	2
<i>S. maltophilia</i>		>256	>256	6	96	1.5	6	2
<i>S. maltophilia</i>		>256	>256	>256	>256	4	>256	4
<i>S. maltophilia</i>		>256	16	72	16	1	8	2
<i>S. maltophilia</i>		>256	>256	>256	>256	0.75	24	0.75
<i>P. aeruginosa</i>	VIM-2 + overexpressed cephalosporinase	16	24	>256	>256	8	12	16
<i>P. aeruginosa</i>	IMP-2 + overexpressed cephalosporinase	12	>256	>256	>256	6	12	24
<i>P. aeruginosa</i>	IMP-1 + overexpressed cephalosporinase	128	>256	>256	>256	96	48	64

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1 ?

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Aztréonam + Ceftazidime/avibactam ?

TABLE 2 MICs and categorization according to CLSI breakpoints for antimicrobials on *NDM*-producing *Enterobacteriaceae*, MBL-producing *P. aeruginosa*, and *S. maltophilia*

<i>Enterobacteriaceae</i> sp.	β -Lactamases	MICs (mg/liter) by treatment ^a						
		ATM	CZA	C/T	AMC	ATM+ CZA	ATM+ C/T	ATM+ AMC
<i>K. pneumoniae</i>	NDM-1 + CTX-M-15 + SHV-11 + OXA-1	>256	>256	>256	12	0.125	24	0.38
<i>K. pneumoniae</i>	NDM-1 + CTX-M-15 + CMY-4 + OXA-1	>256	>256	>256	32	0.75	>256	16
<i>K. pneumoniae</i>	NDM-1 + CTX-M-15 + OXA-1 + OXA-9 + TEM-1 SHV-28 + SHV-11	>256	>256	>256	32	0.25	>256	3
<i>K. pneumoniae</i>	NDM-1 + OXA-1 + SHV-11	>256	>256	>256	12	0.047	0.094	0.094
<i>K. pneumoniae</i>	NDM-1 + OXA-1 + CTX-M-15 + TEM-1 + SHV-28 + OXA-9 + CMY-6	>256	>256	>256	16	0.047	3	0.25
<i>K. pneumoniae</i>	NDM-1 + TEM-1 + CTX-M-15 + SHV-12 + OXA-9	>256	>256	>256	12	0.125	96	1
<i>K. pneumoniae</i>	NDM-1 + TEM-1 + CTX-M-15 + SHV-12 + OXA-9	>256	>256	>256	12	0.125	96	0.5
<i>K. pneumoniae</i>	NDM-1 + TEM-1 + CTX-M-15 + SHV-11 + OXA-1	>256	>256	>256	12	0.064	8	0.38
<i>Salmonella enterica</i>	NDM-1 + CTX-M-15 + TEM-1 + OXA-1 + OXA-9 + OXA-10	>256	>256	>256	16	0.125	16	0.38
<i>E. coli</i>	VIM-1 + CTX-M-3	>256	>256	>256	16	0.125	24	0.5
<i>E. coli</i>	VIM-4 + ESBL	16	>256	>256	24	1.5	24	16
<i>K. pneumoniae</i>	VIM-1 + SHV-5	>256	>256	>256	>256	0.25	192	1.5
<i>K. pneumoniae</i>	VIM-1 + SHV-5	>256	>256	>256	16	0.125	4	0.25
<i>K. pneumoniae</i>	VIM-1 + ESBL	>256	>256	>256	>256	12	16	12
<i>K. pneumoniae</i>	VIM-1 + SHV-5	16	>256	>256	>256	6	12	32
<i>K. pneumoniae</i>	VIM-1 + TEM-1 + SHV-5	96	>256	>256	>256	96	64	48
<i>K. pneumoniae</i>	VIM-1 + SHV-5	>256	>256	>256	24	0.25	8	0.75
<i>K. pneumoniae</i>	VIM-1 + SHV-5	>256	>256	>256	12	0.125	2	0.38
<i>K. pneumoniae</i>	VIM-19 + CTX-M-3 + TEM-1 + SHV-1	6	32	>256	16	0.047	2	1.5

ATM, aztreonam; CZA, ceftazidime-avibactam; C/T, ceftolozane-tazobactam; AMC, amoxicillin-clavulanate

Aztréonam + Ceftazidime/avibactam ?

Ceftazidime-Avibactam and Aztreonam, an Interesting Strategy To Overcome β -Lactam Resistance Conferred by Metallo- β -Lactamases in *Enterobacteriaceae* and *Pseudomonas aeruginosa*

Benjamin Davido,^a Lesly Fellous,^b Christine Lawrence,^{c,d} Virginie Maxime,^e Martin Rottman,^{d,f} Aurélien Dinh^a



Antimicrobial Agents and Chemotherapy

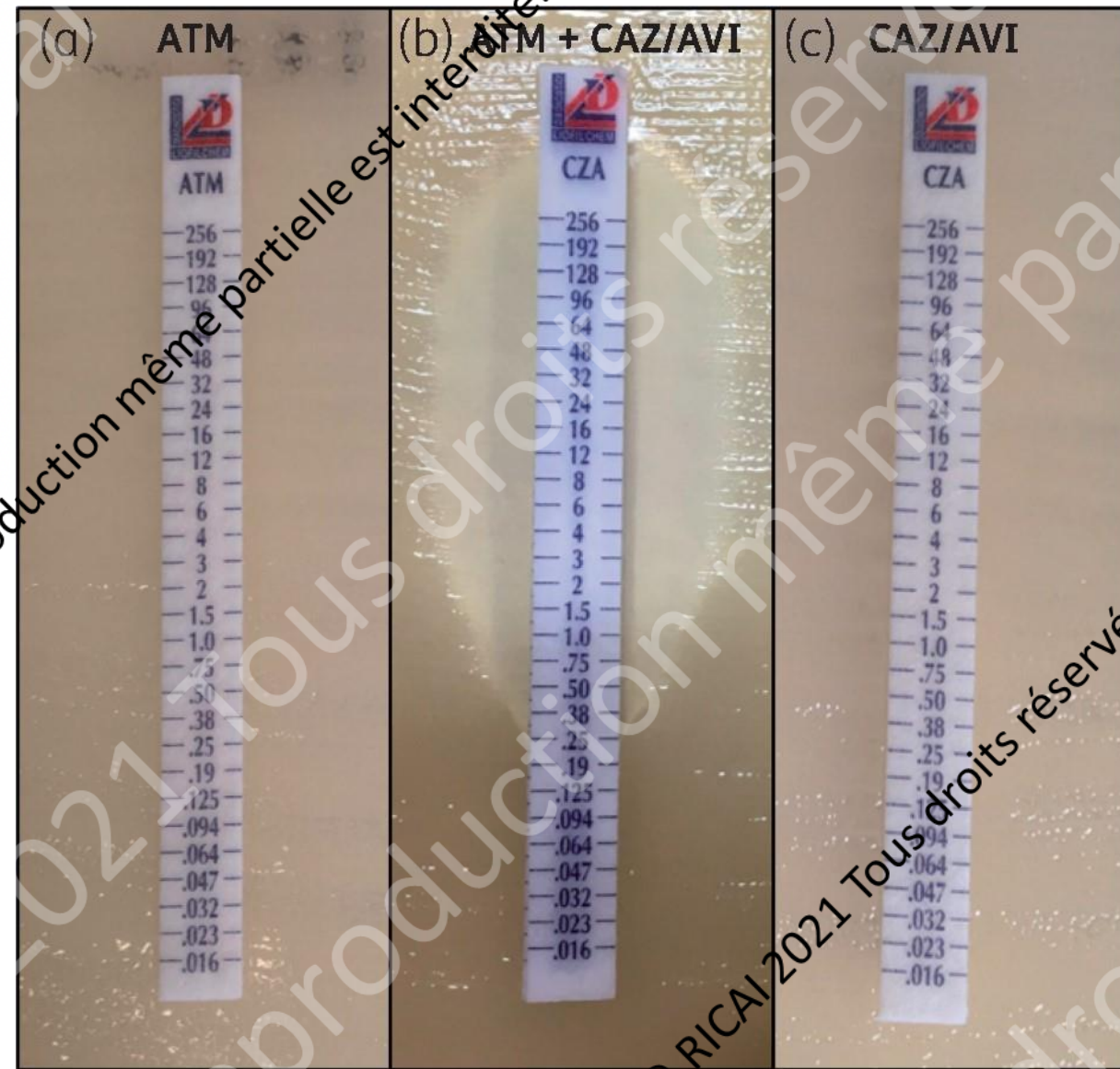


Susceptibility testing showing with ellipsometry the effect of the synergistic combination of CAZ-AVI and ATM. The combination (middle strip) was tested by first applying an ATM strip to the Mueller-Hinton (MH) agar, removing it after 5 min, and then applying a CAZ-AVI strip on the exact same location and placing back the ATM strip to read the susceptibility to ATM in the presence of AVI (and CAZ). *K. pneumoniae* NDM-1/OXA-48 from patient 1.

Ceftazidime/avibactam alone or in combination with aztreonam against colistin-resistant and carbapenemase-producing *Klebsiella pneumoniae*

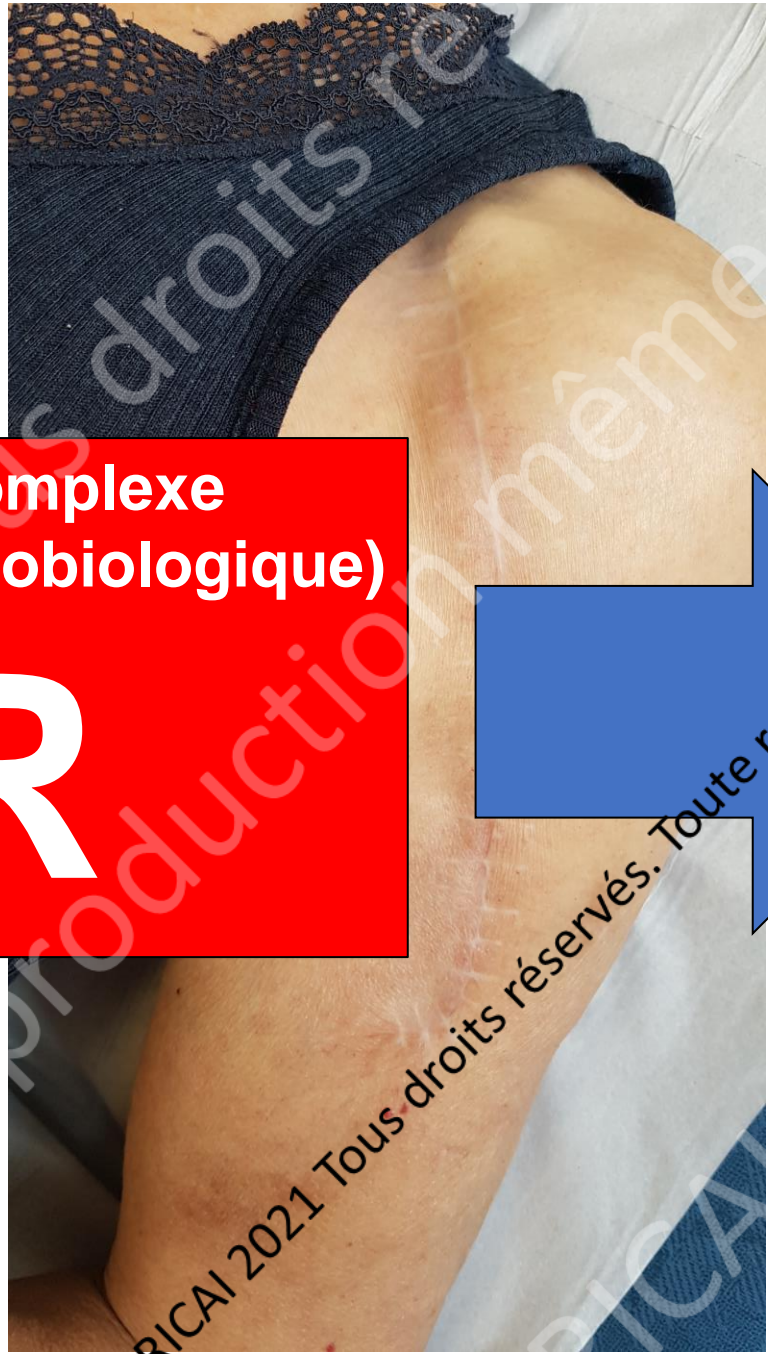
Aurélie Jayol¹⁻⁴, Patrice Nordmann^{1-3,5},
Laurent Poirel^{1-3*} and Véronique Dubois^{4,6}

Figure 1 Example of synergistic combination of ceftazidime/avibactam (CAZ/AVI) and aztreonam (ATM) for an NDM + ESBL²-producing *K. pneumoniae*. Susceptibility testing of ATM alone (a), combination of CAZ/AVI with ATM (b) and CAZ/AVI alone (c).



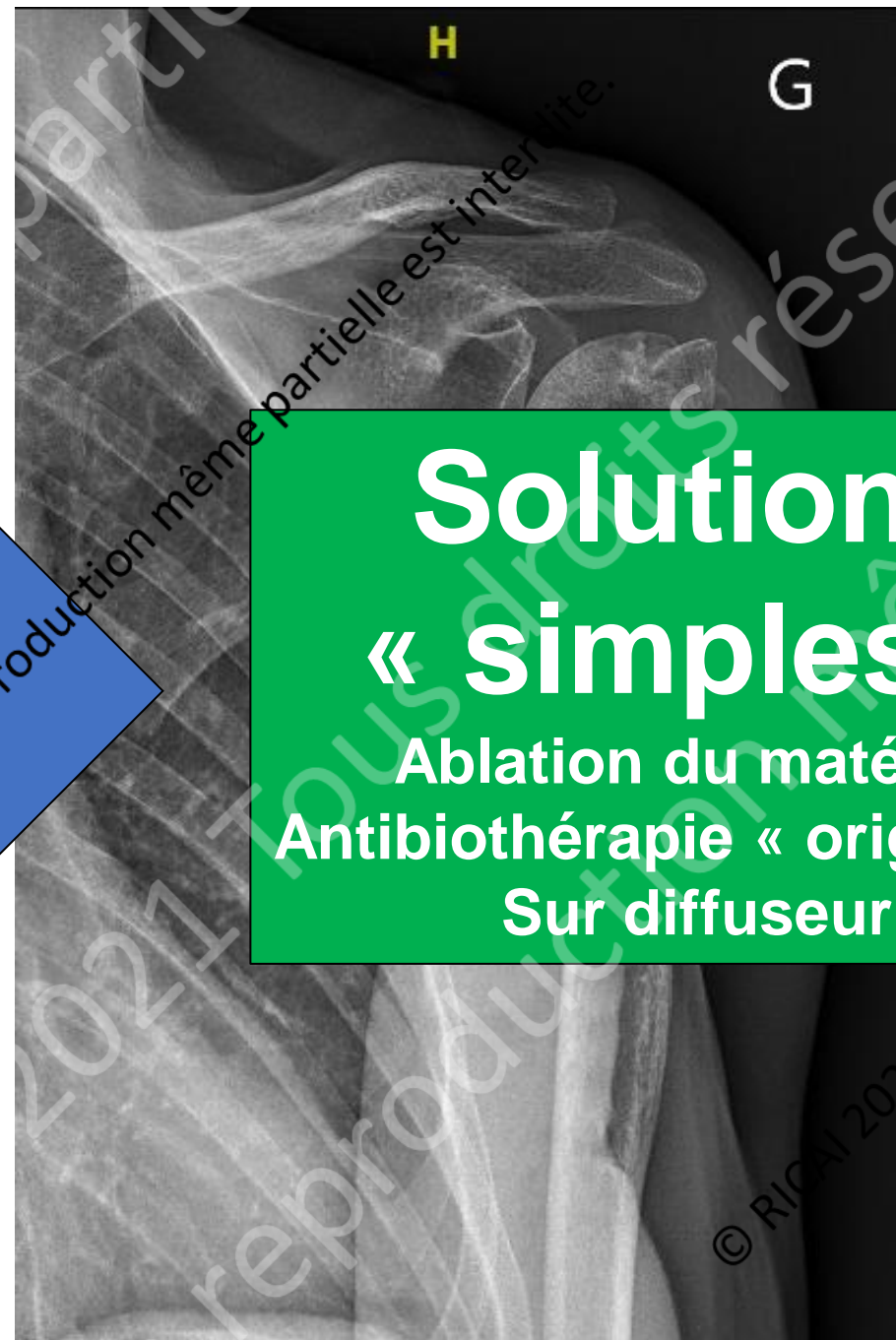
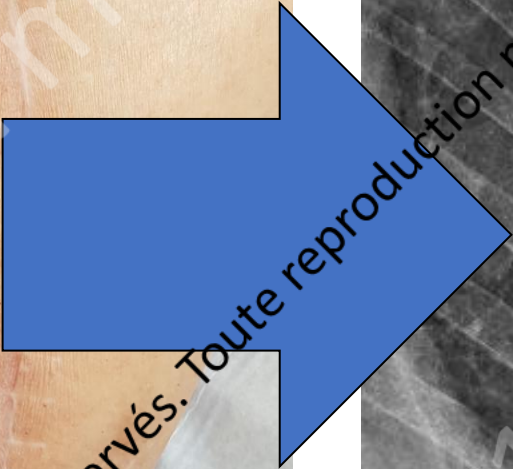
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**IOA Complexe
(critère microbiologique)**

R



Solutions

« simples »

Ablation du matériel
Antibiothérapie « originale »
Sur diffuseur

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Bacilles à Gram négatif de résistance émergentes aux antibiotiques : Nouveaux antibiotiques et nouvelles stratégies thérapeutiques

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Claude Bernard Lyon1 University, Lyon

Centre International de Recherche en Infectiologie, CIRI, Inserm U1111, CNRS
UMR5308, ENS de Lyon, UCBL1, Lyon, France

Regional referral center for the management of complex bone and joint infection
Centre de Référence des IOA complexes de Lyon (CRIOAc Lyon)



2020 ANTIBACTERIAL AGENTS IN CLINICAL AND PRECLINICAL DEVELOPMENT

an overview and analysis



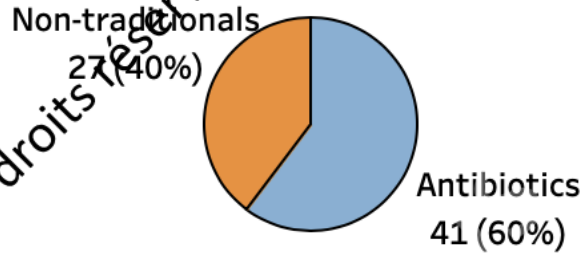
World Health
Organization

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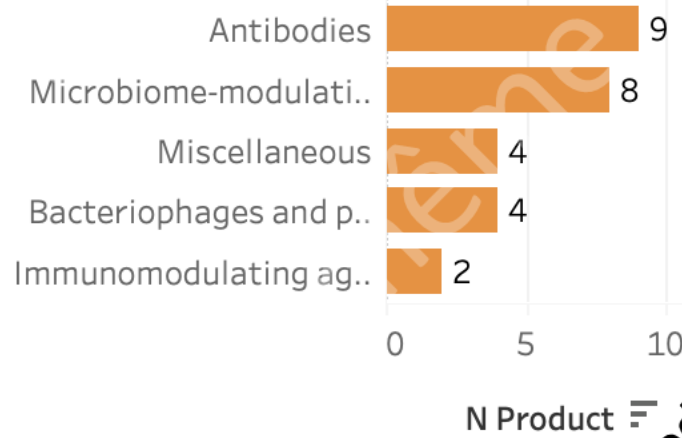
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A.1. Products by type



A.2. No. of non traditional products by category



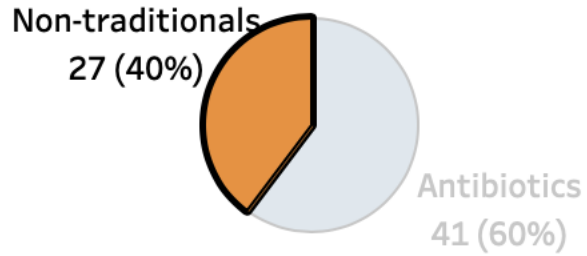
A.3. Products by pathogen category and phase

Pathogen category	Phase I	Phase II	Phase	Unkno..	Total
Priority pathogens	18	15	9	1	43
Mycobacterium tuberculosis	3	9			12
Clostridium difficile	3	8	2		13
Total	24	32	11	1	68

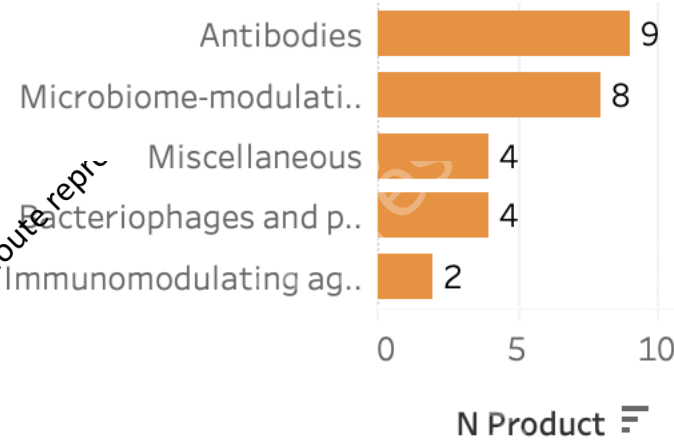
B. Expected activity against priority pathogens

Active?	Critical priority pathogens					Other priority pathogens							Subtotal	Total
	Acinetobac baumannii	Pseudomor aeruginosa	Enteroba..	All critical priority pathogens	Subtotal	Gram-positive priority p..	Neisseria gonorhoei	Helicobact pylori	Staphyloco aureus	Enterococc faecium	Streptococ pneumonia	Campyloba spp.		
Yes	7	7	14	3	31	17	3	2	17	3	2	2	21	38
Possibly	3	3	3	2	6	1	1	1	1	1			2	8
No	12	17	10	17	18	3	7	8	3	7	7	8	10	20

A.1. Products by type



A.2. No. of non traditional products by category



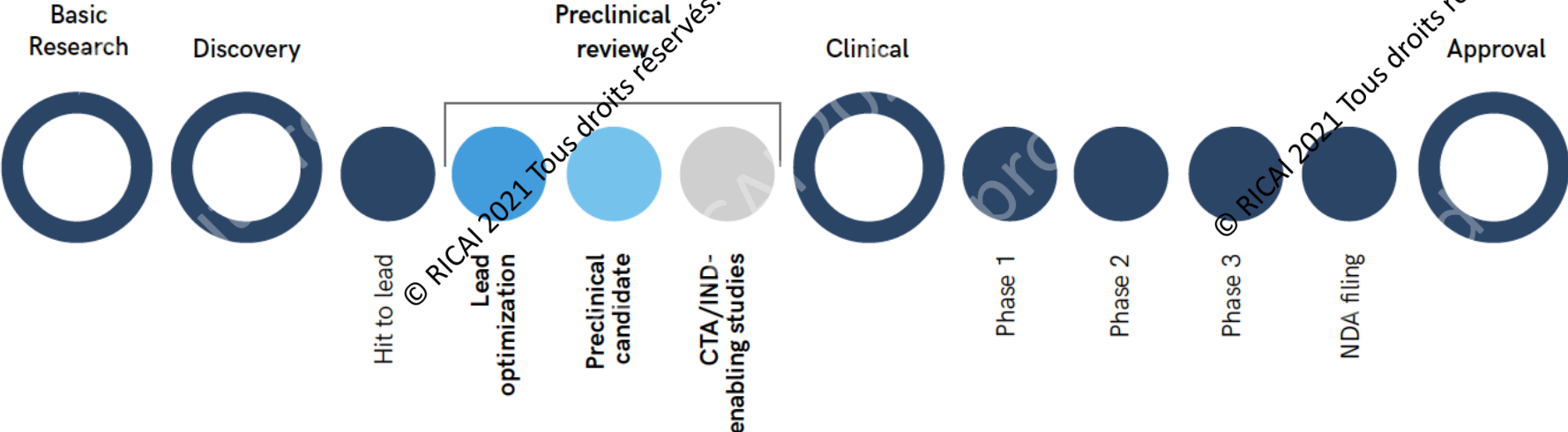
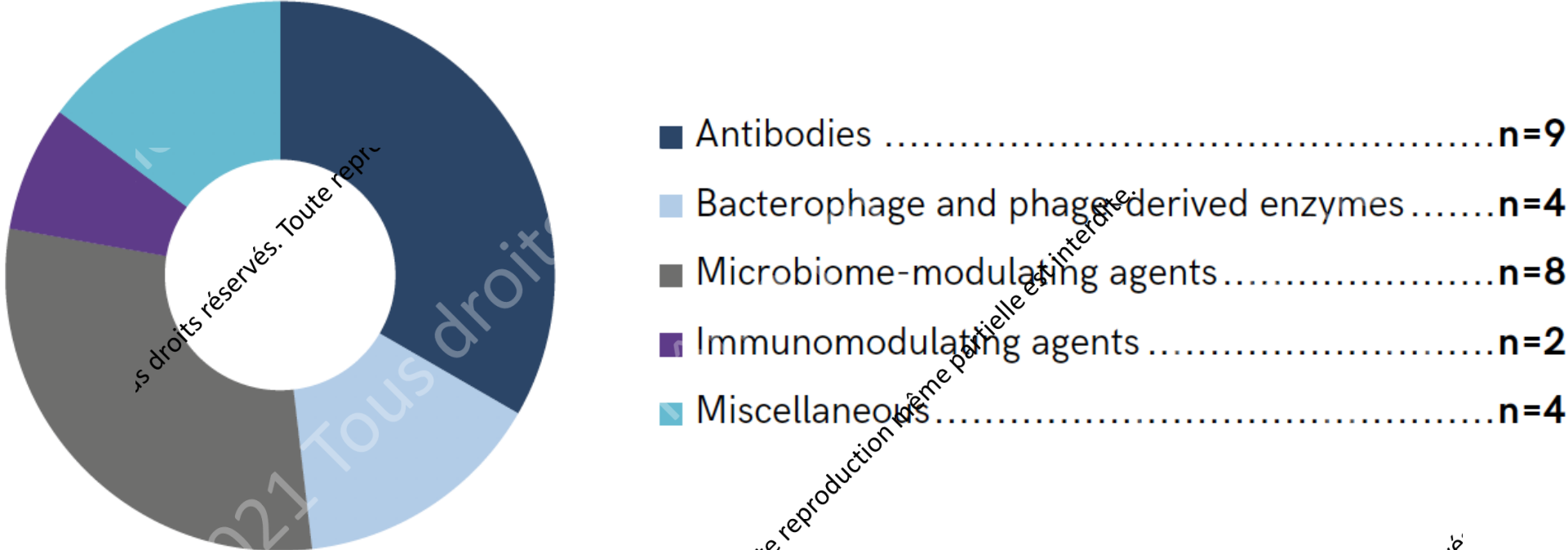
A.3. Products by pathogen category and phase

Pathogen category	Phase I	Phase II	Phase	Unkno..	Total
Priority pathogens	4	11	3	1	19
Clostridium difficile	2	5	1		8
Total	6	16	4	1	27

B. Expected activity against priority pathogens

Active?	Critical priority pathogens					Other priority pathogens							Total	
	Acinetobac baumannii	Pseudomonas aeruginosa	Enteroba..	All critical priority pathogens	Subtotal	Gram-positive priority p..	Neisseria gonorhoei	Helicobact pylori	Staphyloco aureus	Enterococc faecium	Streptococ pneumoniae	Campyloba spp.		Subtotal
Yes	1	5	6	1	4	10	1	1	10	1	1	2	12	19

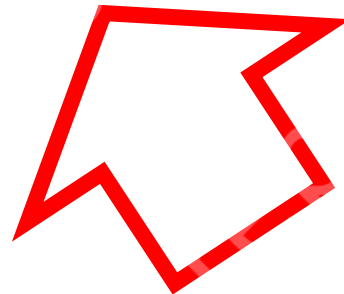
Fig. 7. Number of non-traditional antibacterials in the clinical pipeline.



Name (synonym)	Phase	Antibiotic class	Route of administration (developer)	Expected activity against priority pathogens
CF-301 (exebacase)	3	Phage endolysin	iv (ContraFect)	<i>S. aureus</i>
SAL-200 (tonabacase)	2a	Phage endolysin	iv (iNtRON Biotechnology, Roivant Sciences)	<i>S. aureus</i>
PhageBank	1/2	Phage bank (process)	oral (Adaptive Phage Therapeutics and US Department of Defense)	<i>E. coli</i> , <i>K. pneumoniae</i>
LBP-EC01	1b	CRISPR-Cas3 enhanced phage	iv (Locus Bioscience)	<i>E. coli</i> , <i>K. pneumoniae</i>

Traitement adjuvant dans les bactériémies à *S. aureus*

Name (synonym)	Phase	Antibiotic class	Route of administration (developer)	Expected activity against priority pathogens
CF-301 (exebacase)	3	Phage endolysin	iv (ContraFect)	<i>S. aureus</i>
SAL-200 (tonabacase)	2a	Phage endolysin	iv (iNtRON Biotechnology, Roivant Sciences)	<i>S. aureus</i>
PhageBank	1/2	Phage bank (process)	oral (Adaptive Phage Therapeutics and US Department of Defense)	<i>E. coli</i> , <i>K. pneumoniae</i>
LBP-EC01	1b	CRISPR-Cas3 enhanced phage	iv (Locus Bioscience)	<i>E. coli</i> , <i>K. pneumoniae</i>



Traitement vs placebo dans les colonisations ou infections urinaires

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draft

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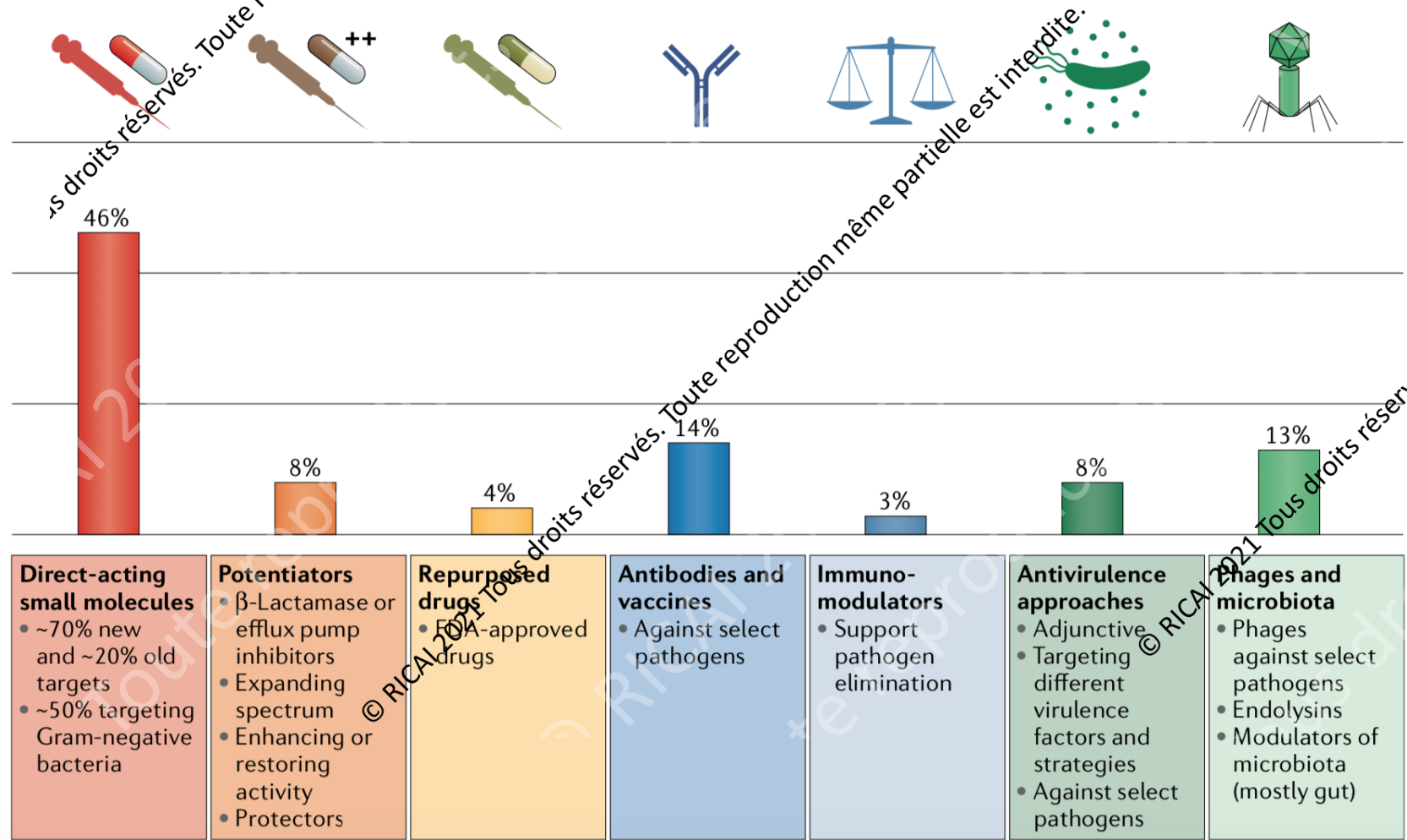
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Name (synonym)	Phase	Antibiotic class	Route of administration (developer)	Expected activity against priority pathogens
CF-301 (exebacase)	3	Phage endolysin	iv (ContraFect)	<i>S. aureus</i>
SAL-200 (tonabacase)	2a	Phage endolysin	iv (iNtRON Biotechnology, Roivant Sciences)	<i>S. aureus</i>
PhageBank	1/2	Phage bank (process)	oral (Adaptive Phage Therapeutics and US Department of Defense)	<i>E. coli</i> , <i>K. pneumoniae</i>
LBP-EC01	1b	CRISPR-Cas3 enhanced phage	iv (Locus Bioscience)	<i>E. coli</i> , <i>K. pneumoniae</i>
+				
Phagos (PHRC 2015)	1/2	Bacteriophage cocktail	Local (Pherecydes)	<i>S. aureus</i>
PhagoPied (PHRC 2015)	1/2	Bacteriophage cocktail	Local (Pherecydes)	<i>S. aureus</i>
PhagoDAIR	2	Bacteriophage cocktail	Local (Pherecydes)	<i>S. aureus</i>

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The global preclinical antibacterial pipeline

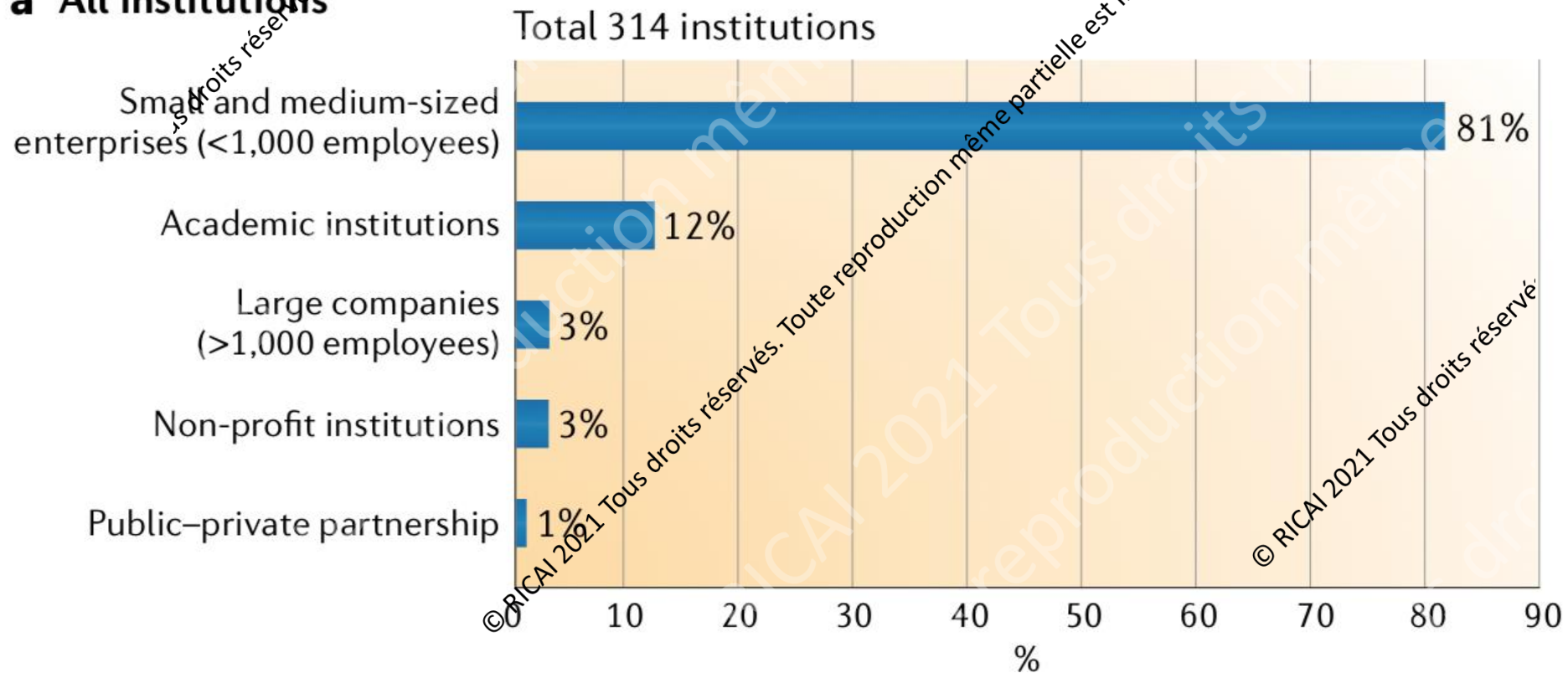
Ursula Theuretzbacher ^{1*}, Kevin Outterson ^{2,3}, Aleks Engel ⁴ and Anders Karlén ⁵



The global preclinical antibacterial pipeline

Ursula Theuretzbacher ^{1*}, Kevin Outterson ^{2,3}, Aleks Engel ⁴ and Anders Karlén ⁵

a All institutions



The global preclinical antibacterial pipeline

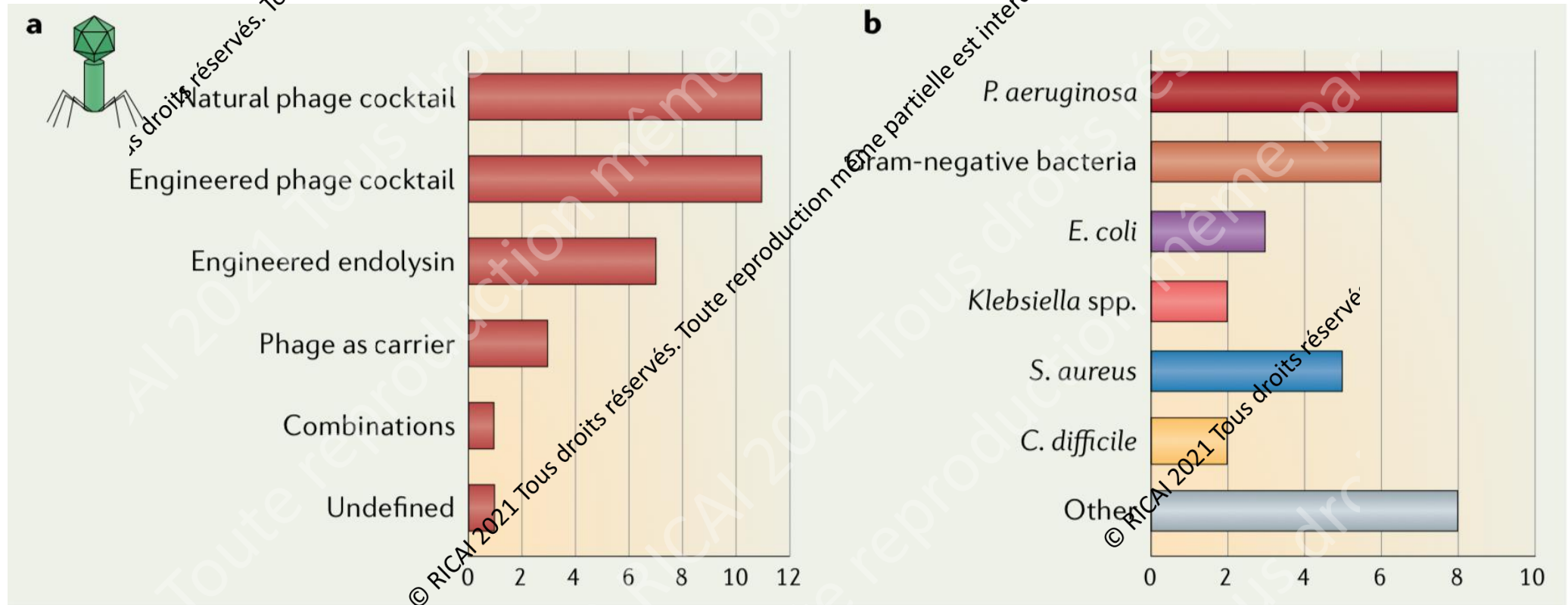
Ursula Theuretzbacher ^{1*}, Kevin Outterson ^{2,3}, Aleks Engel ⁴ and Anders Karlén ⁵

b Small and medium-sized enterprises



The global preclinical antibacterial pipeline

Ursula Theuretzbacher ^{1*}, Kevin Outterson ^{2,3}, Aleks Engel ⁴ and Anders Karlén ⁵



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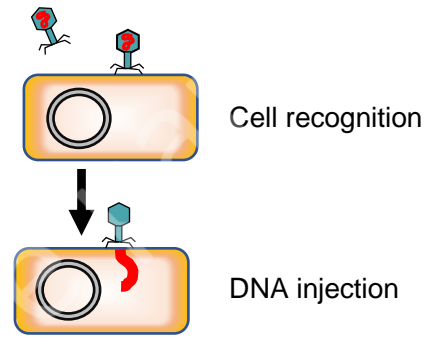


- Creating standards
- Support knowledge exchange
- Designing, initiating and supporting clinical trials
- Discuss with EMA about monography for phages
- Creating an register & database for phage therapy
- ...

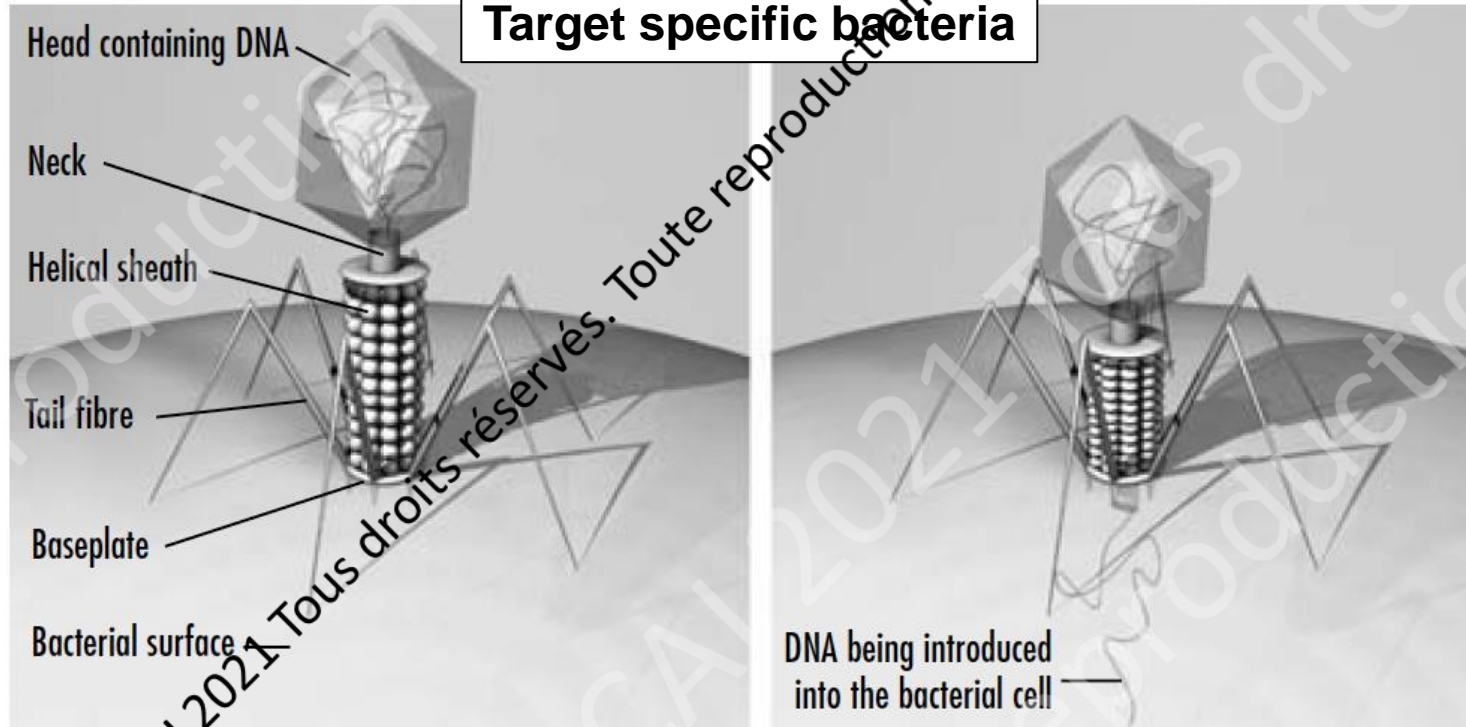
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The phage life



Environmental viruses
Target specific bacteria



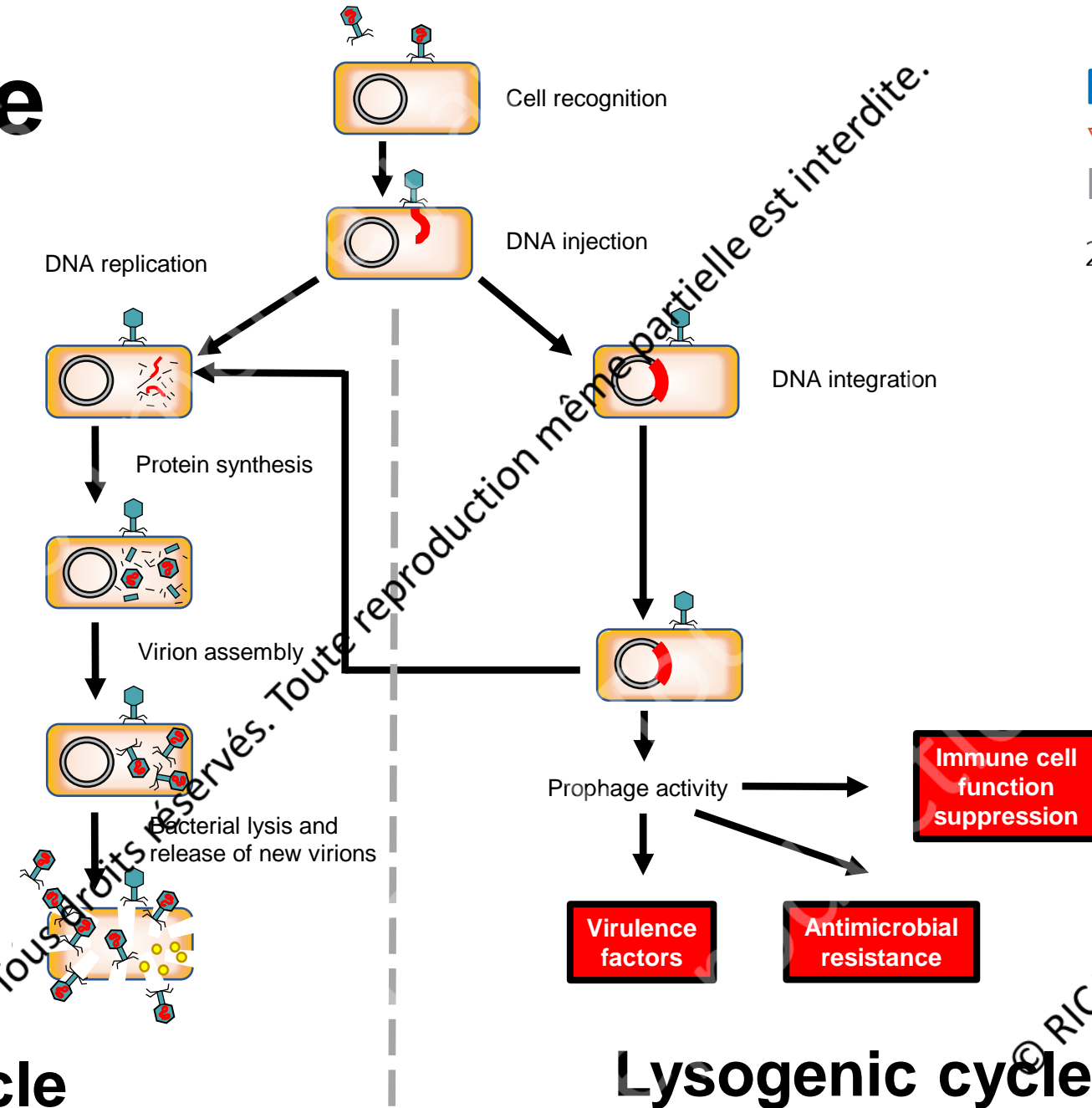
Ferry T. et al.

Virologie

2020;24(1):49-56

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The phage life

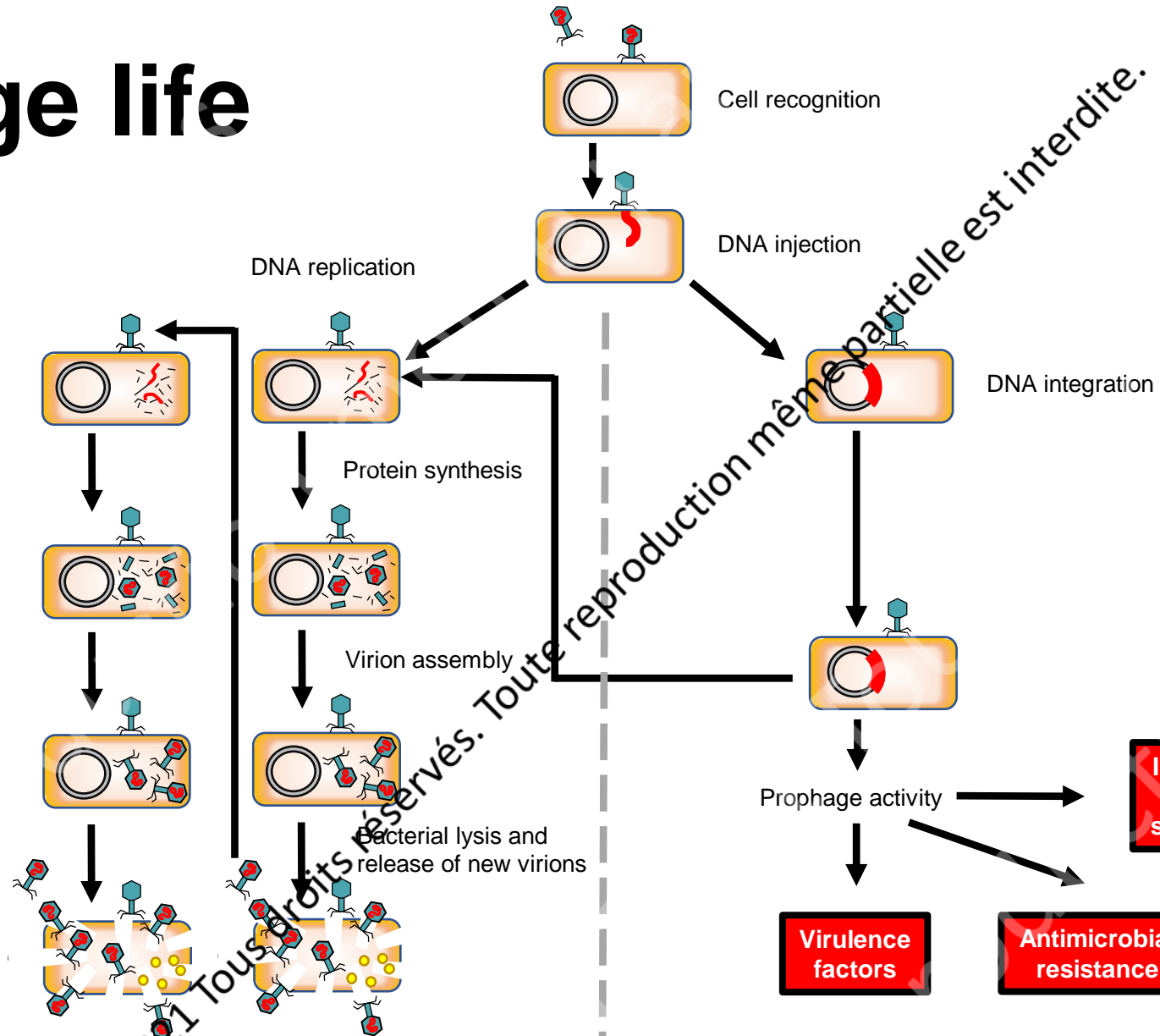


Lytic cycle
Self-maintained bacterial lysis

Lysogenic cycle
Bacterial genetic remodeling

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The phage life



Lytic cycle

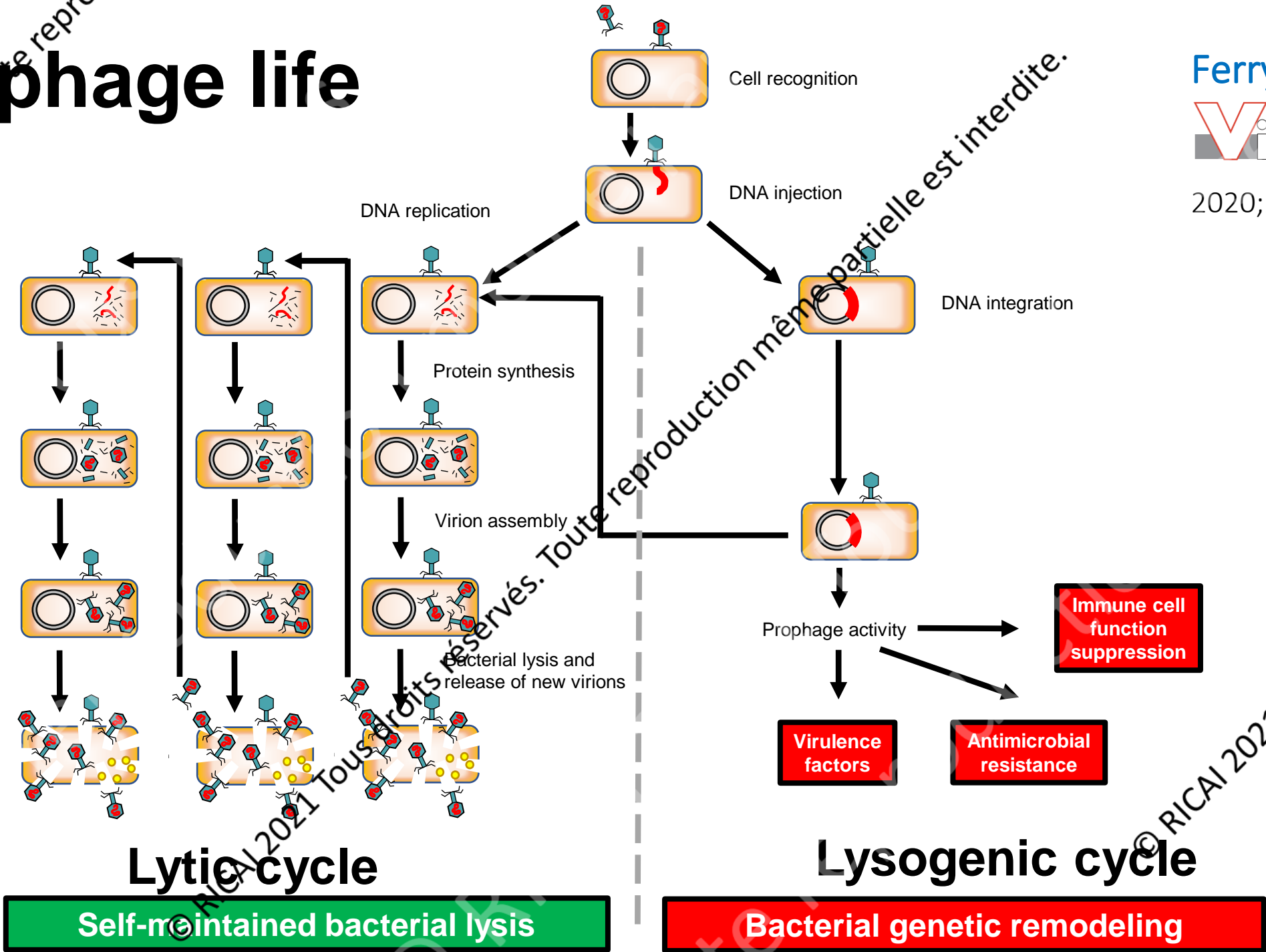
Self-maintained bacterial lysis

Lysogenic cycle

Bacterial genetic remodeling

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The phage life



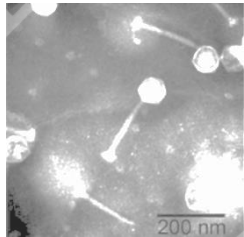
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Only lytic phages have to be used

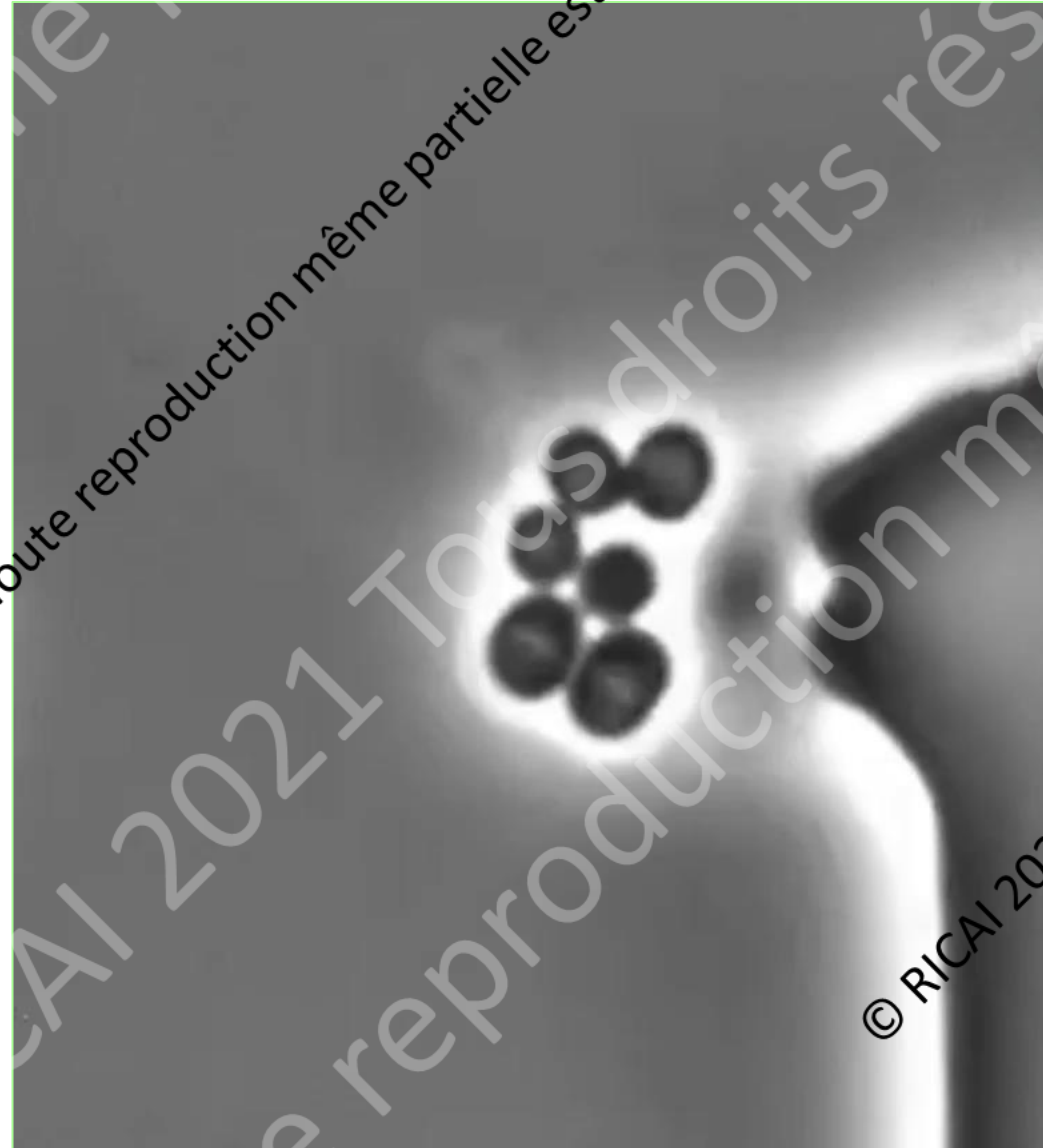
***S. aureus* being lysed
by the Sa2 phage**



Bacterial DNA
appeared in green

Courtesy Pascal Maugin
Luciano Marraffin Lab

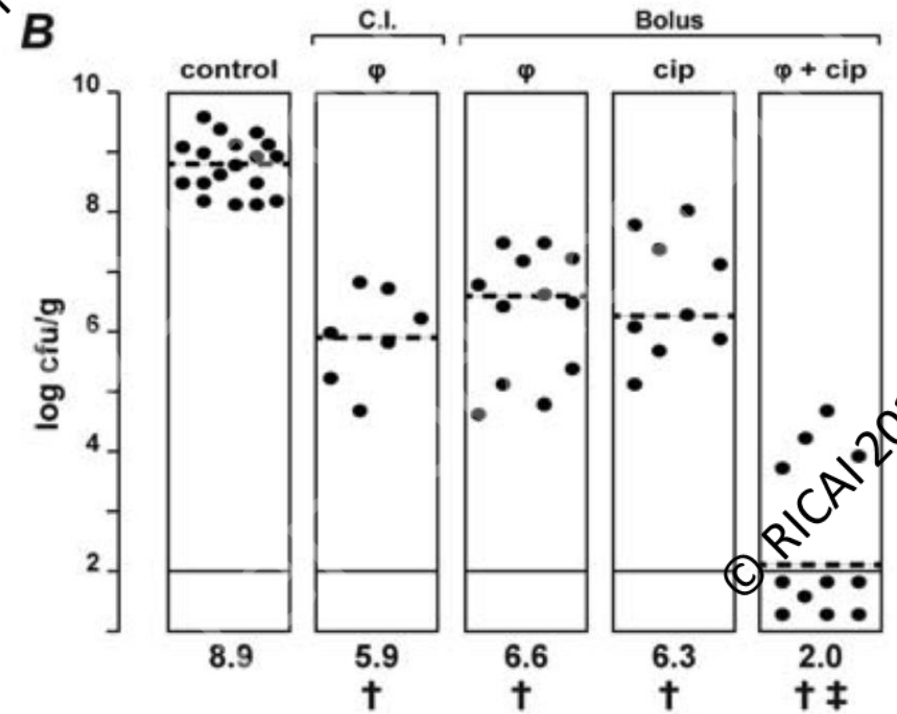
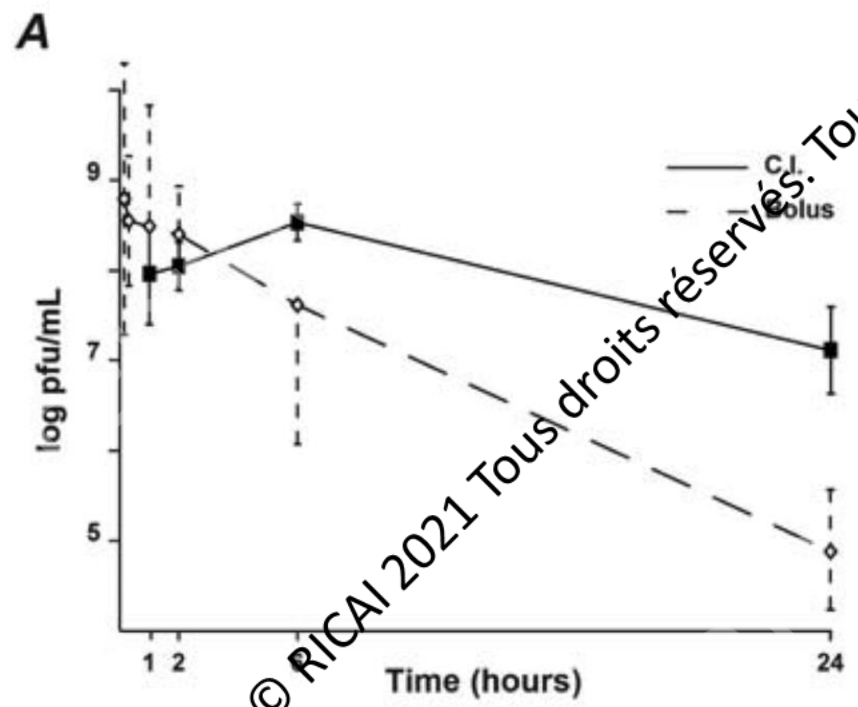
THE ROCKEFELLER UNIVERSITY



Synergistic Interaction Between Phage Therapy and Antibiotics Clears *Pseudomonas Aeruginosa* Infection in Endocarditis and Reduces Virulence

Frank Oechslin,¹ Philippe Piccardi,¹ Stefano Mancini,¹ Jérôme Gabard,³ Philippe Moreillon,¹ José M. Entenza,¹ Gregory Resch,¹ and Yok-Ai Que² 2015

¹Department of Fundamental Microbiology, University of Lausanne, and ²Department of Intensive Care Medicine, Bern University Hospital, Switzerland; and ³Pherecydes Pharma, Romainville, France

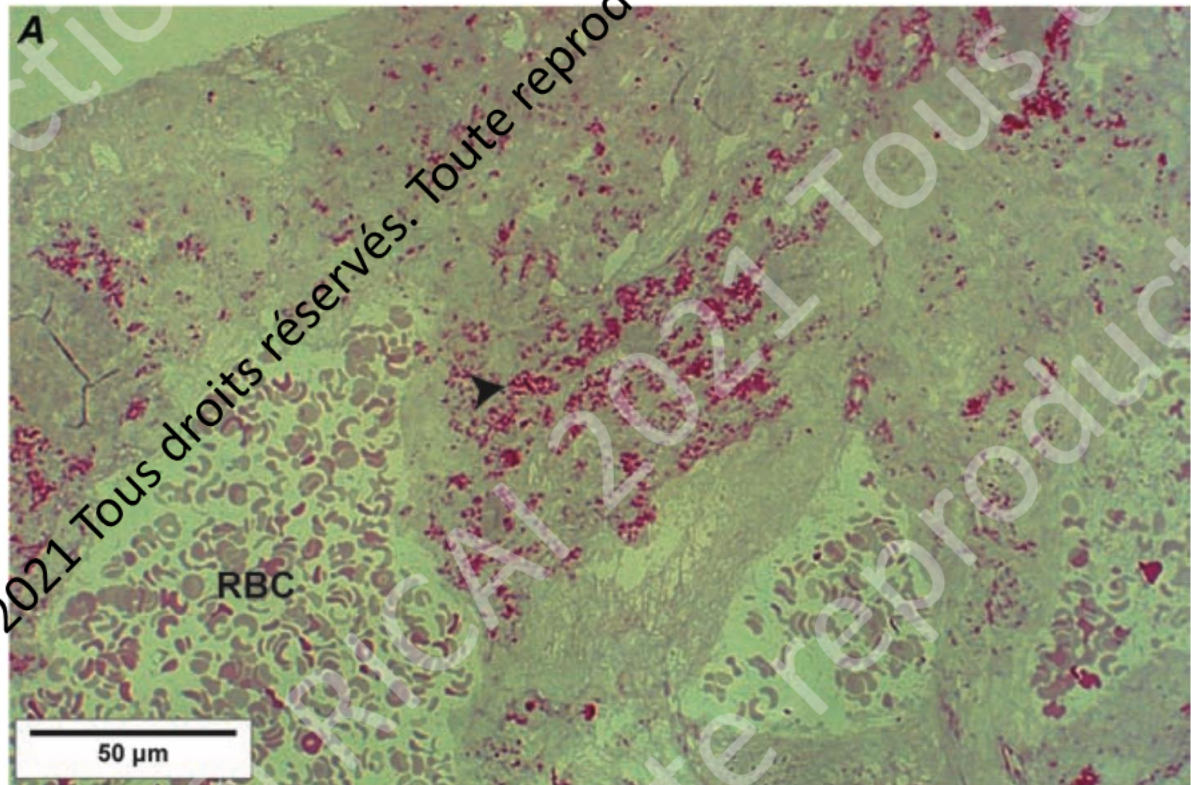


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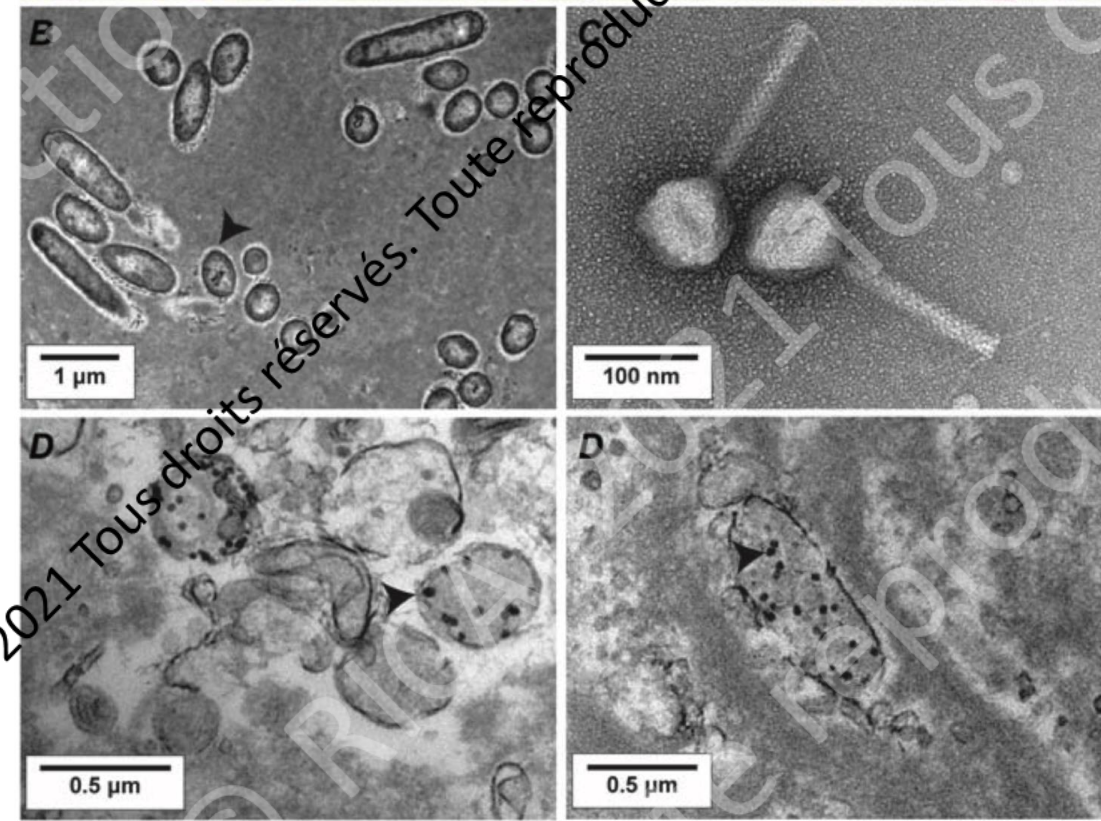
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Implementation of a Phage Therapy Center in a CRIOAc

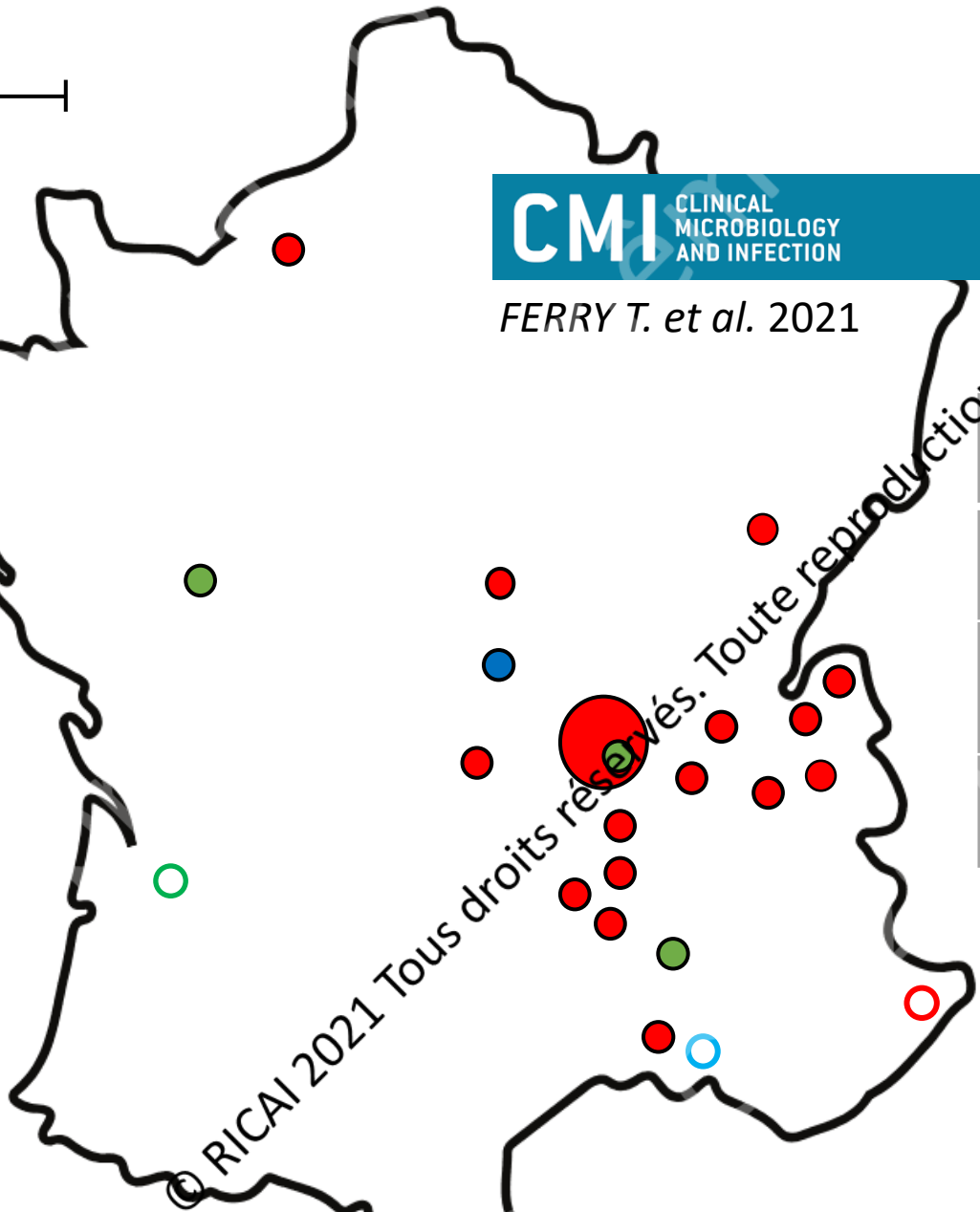
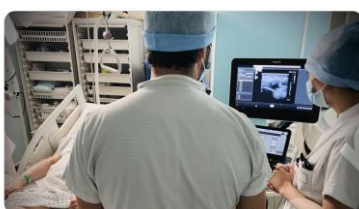
250 km



CMI CLINICAL MICROBIOLOGY AND INFECTION

FERRY T. et al. 2021

Number of patients	2017	2018	2019	2020	Total
Managed in CRIAC Lyon	557	594	647	520	2318
For whom a phagogram was performed	7 (1.2%)	10 (1.7%)	17 (2.6%)	23 (4.4%)	57 (2.4%)
For whom phage therapy was done	4 (0.7%)	2 (0.3%)	8 (1.2%)	7 (1.3%)	21 (0.9%)



Under the supervision of **ansm**

●○ BJI ●○ Endocarditis ●○ Pneumonia

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Implementation of a Phage Therapy Center in a CRIOAc

250 km



PHAGEinLYON



CMI CLINICAL MICROBIOLOGY AND INFECTION

FERRY T. et al. 2021

30 patients in Lyon since 2017

- 27 with phages from



- 3 with phages from

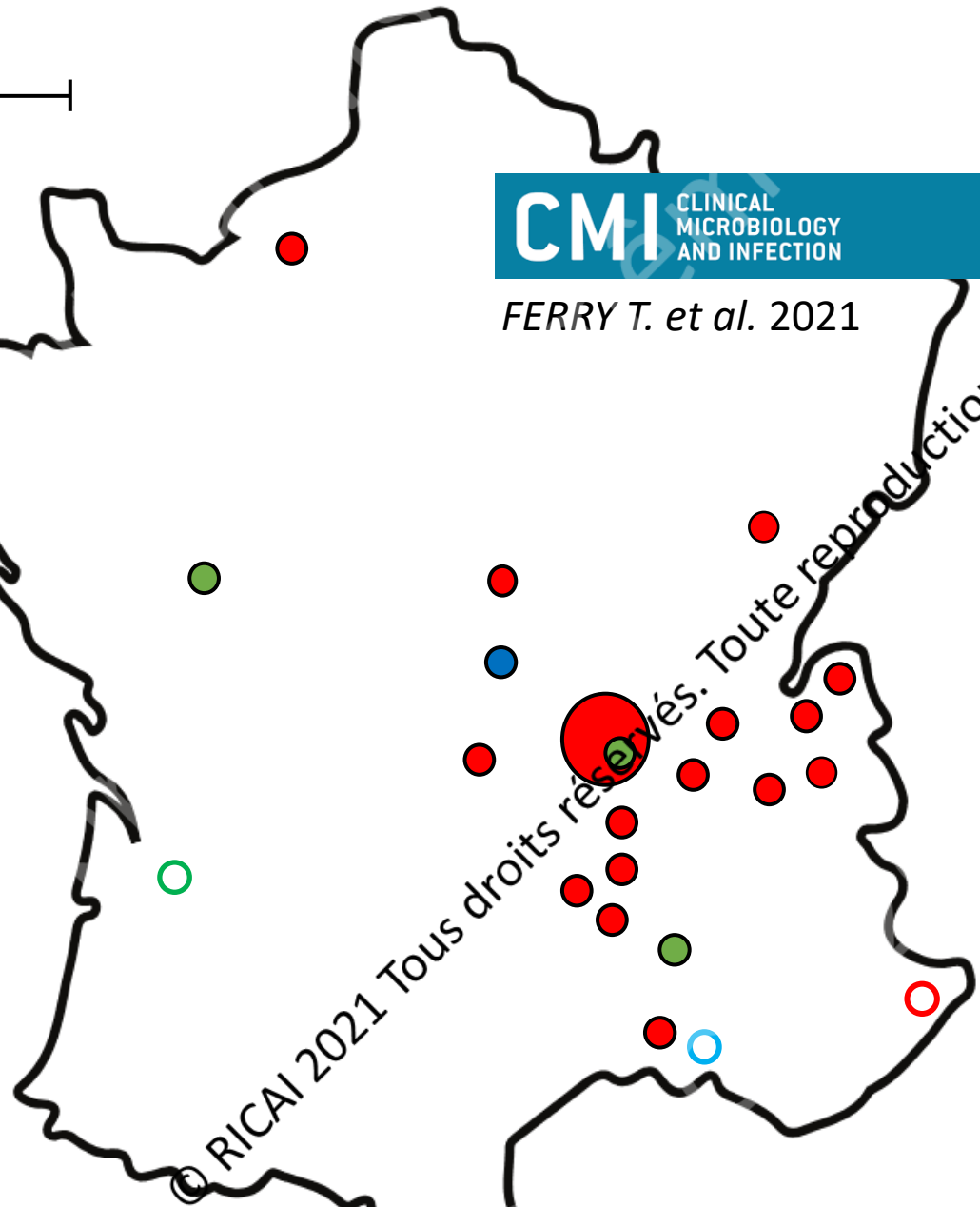
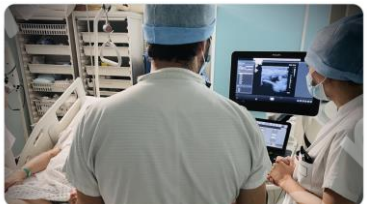


- 26 **BJI** (including 22 **PJI**)
- 3 **endocarditis**
- 1 **VAP + bacteremia**

+ 3 patients managed outside Lyon

Under the supervision of **ansm**

●○ **BJI** ●○ **Endocarditis** ●○ **Pneumonia**



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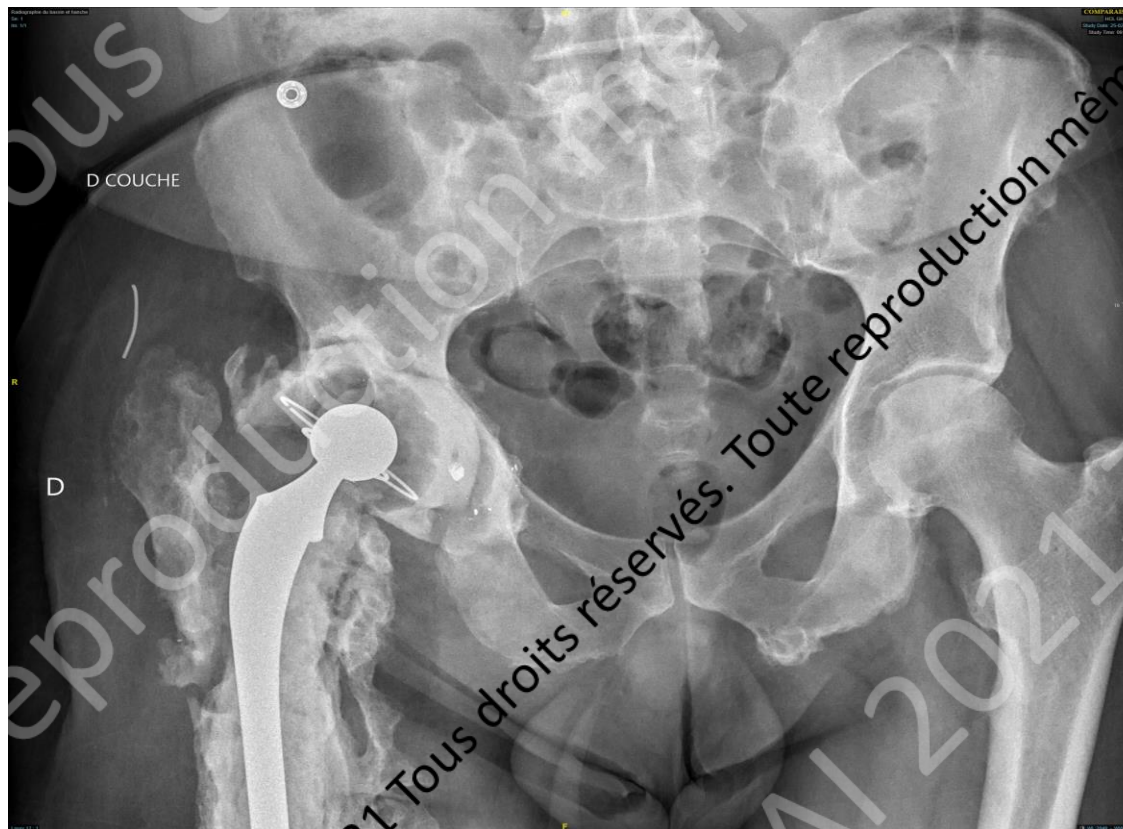
Implementation of a Phage Therapy Center in a CRIOAc

13 patients (43%)

Created for *P. aeruginosa* severe infection



PHAGEin LYON



- 11 with phages from



- 2 with phages from



- 10 **BJI** (including 8 **PJI**)
- 2 **endocarditis**
- 1 **VAP + bacteremia**

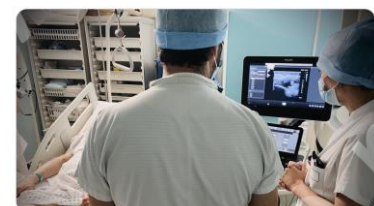
Under the supervision of ANSM

●○ **BJI**

●○ **Endocarditis**

●○ **Pneumonia**

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Implementation of a Phage Therapy Center in a CRIOAc

13 patients (43%)

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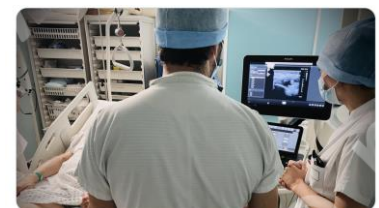


- 10 **BJI** (including 8 **PJI**)
- 2 **endocarditis**
- 1 **VAP + bacteremia**

Under the supervision of  **ansm**
Agence nationale de sécurité des médicaments et des produits de santé

●○ **BJI** ●○ **Endocarditis** ●○ **Pneumonia**

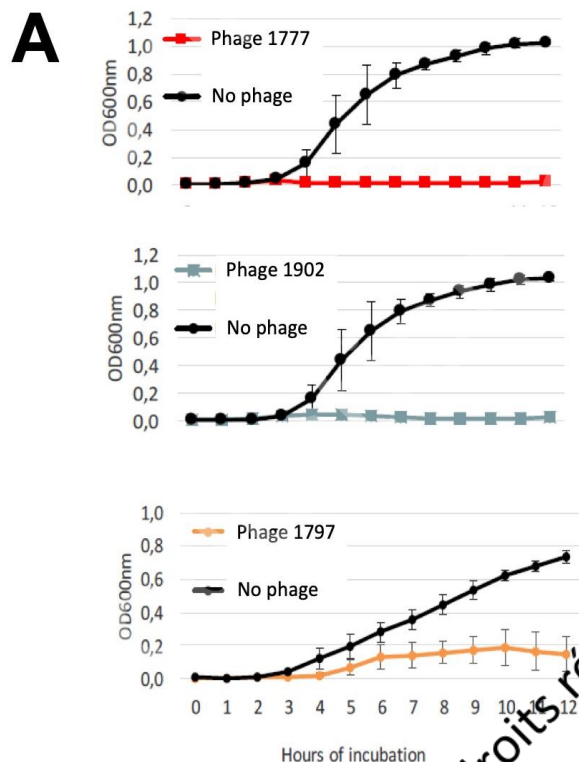
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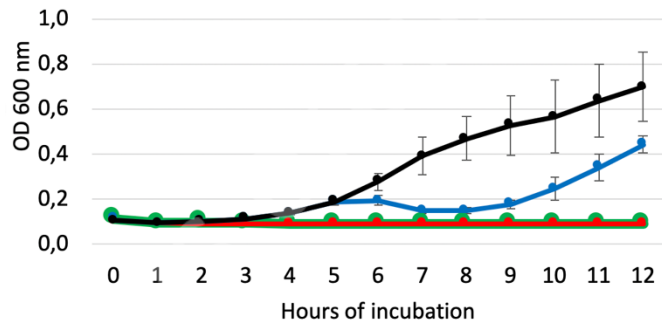
Intravenous administration of personalized cocktail of bacteriophages as salvage therapy in combination with ceftazidime/avibactam in patients with relapsing *P. aeruginosa* bacteremia: Lesson learned from two cases



Conclusions: The type of filter used for the magistral preparation and the duration of the perfusion influenced the phage titer, as the titer in the patient's blood. Personalized GMP bacteriophage therapy has the potential to be used as salvage therapy of *P. aeruginosa* intravascular implant infections.

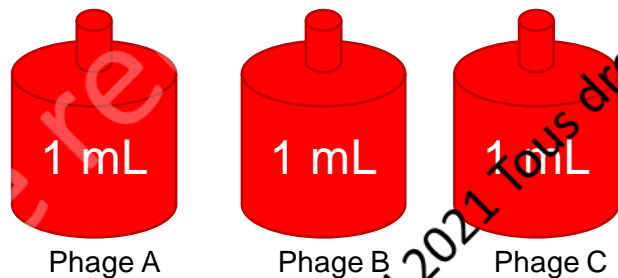
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PHAGE_{in}LYON team



Phagogram
Selection of active bacteriophages

Active GMP *S. aureus* Bactériophages



ID Clinic



CRIAAC LYON



Surgery



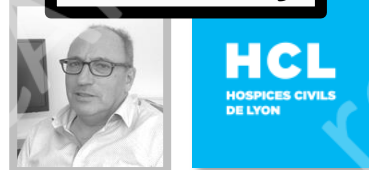
Lab



Under the supervision of



Pharmacy



Extemporaneous magistral preparation of the mix of bacteriophages



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PHAG-ONE

Reunion de Lancement Mercredi 8 Septembre 2021

7 Workpackages – 10 partenaires



www.chu-lyon.fr

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PHAG-ONE

Reunion de Lancement
Mercredi 8 Septembre 2021

7 Workpackages – 10 partenaires

Isoler, tester, caractériser, produire, purifier et libérer
des lots pharmaceutiques
de phages thérapeutiques académiques
à coût maîtrisé utilisables en clinique chez l'Homme

3 pathogènes cible

Staphylococcus aureus / Staphylococcus epidermidis / Escherichia coli



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Organisation du projet PHAG-ONE

WP7 : Etude socio-antropologique

- Epistémologie / Suivi de l'émergence d'une innovation biomédicale
- Perception/Suivi/Accompagnement chez grand public-décideurs-politiques

WP1 : Phage discovery

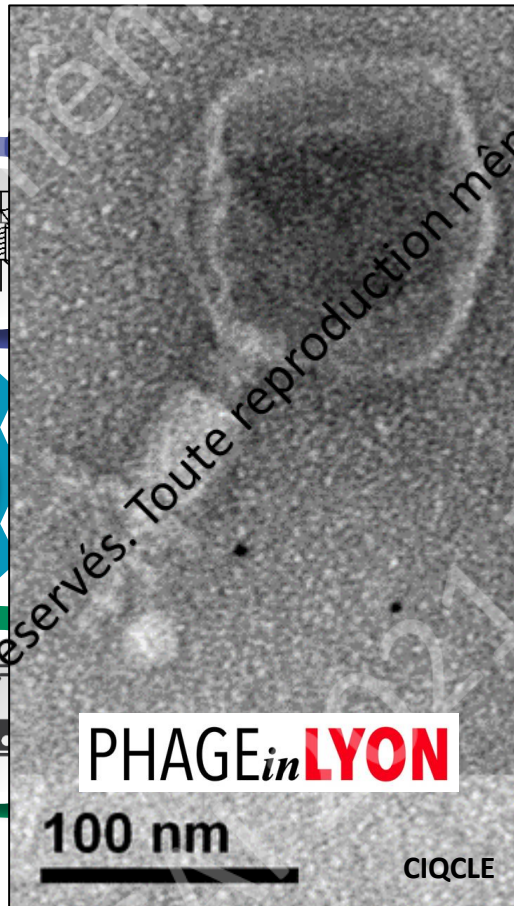
- Isolement
- Caractérisation
- Entraînement

WP2 : USP

- Sélection souche de production
- Développement et validation process amplification phagique
- Contrôles qualité

WP3 : DSP / FF

- Développement et validation process de purification
- Contrôles qualité
- Libération de lots
- Formulation / Stabilité



Safety

- Tolerance
- Toxicité

WP6 : Utilisation clinique

- Référentiels
- Etudes de cohorte
- Essais cliniques

WP5 : Diagnostic

- Titration
- Phagogramme

WP4 : Efficacité / PK-PD

- Modèle Lapin - Tissu Cage
- Modèle Lapin - Infection sur prothèse
- Etude PK/PD *in vitro* et *in vivo*



PRODUCTION de médicaments hospitaliers, non disponibles sur le marché, ou pour le traitement de maladies orphelines ; production de médicaments expérimentaux adaptés au protocole de recherche clinique (mise en insu ...)



CONTRÔLES QUALITÉ des matières premières à usage pharmaceutique et des préparations hospitalières. Centre de référence pour contrôles des eaux de dialyse (endotoxines). **ETUDES DE STABILITÉ** selon la méthodologie ICH.



RECHERCHE ET DEVELOPPEMENT de nouvelles préparations adaptées à des besoins cliniques spécifiques ou adaptées aux exigences du protocole de recherche clinique.



EXPERTISE TECHNIQUE et REGLEMENTAIRE de la gestion des essais cliniques associée à la fabrication/contrôles des médicaments expérimentaux (essais institutionnels).

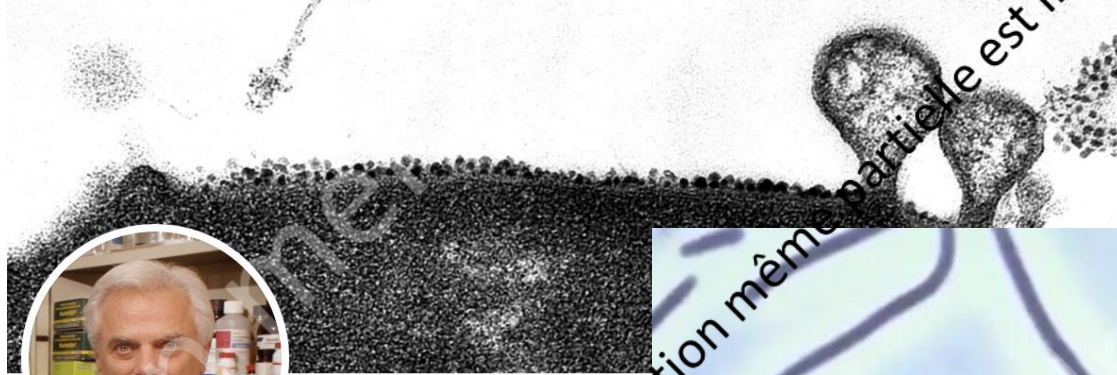


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Lysines de bactériophages



Vincent A Fischetti
@microbephage



Tristan Ferry Lyon University Hospitals
@FerryLyon

Incredible talk of Pr. Vincent A. Fischetti @microbephage @IDWeek2019 about the great potential of #bacteriophage #lysins to induce bacterial explosion... and disappearance! It's good to hear that he discovered lysins that are active against #multidrugresistant #ESKAPE pathogens!

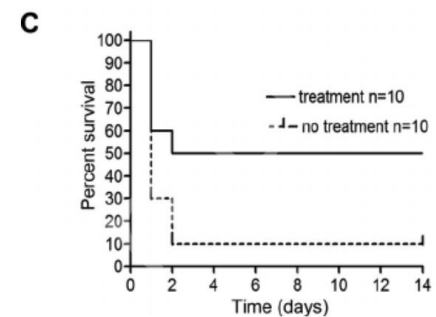
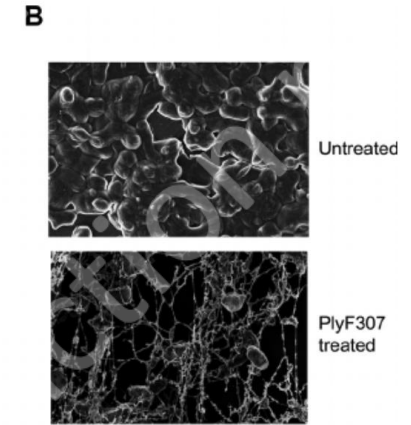
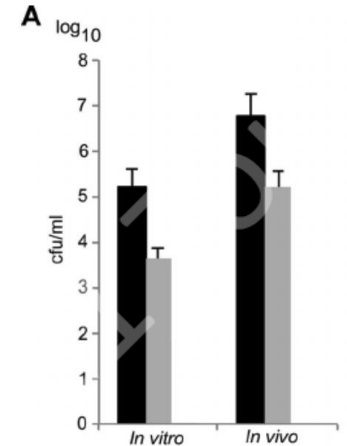
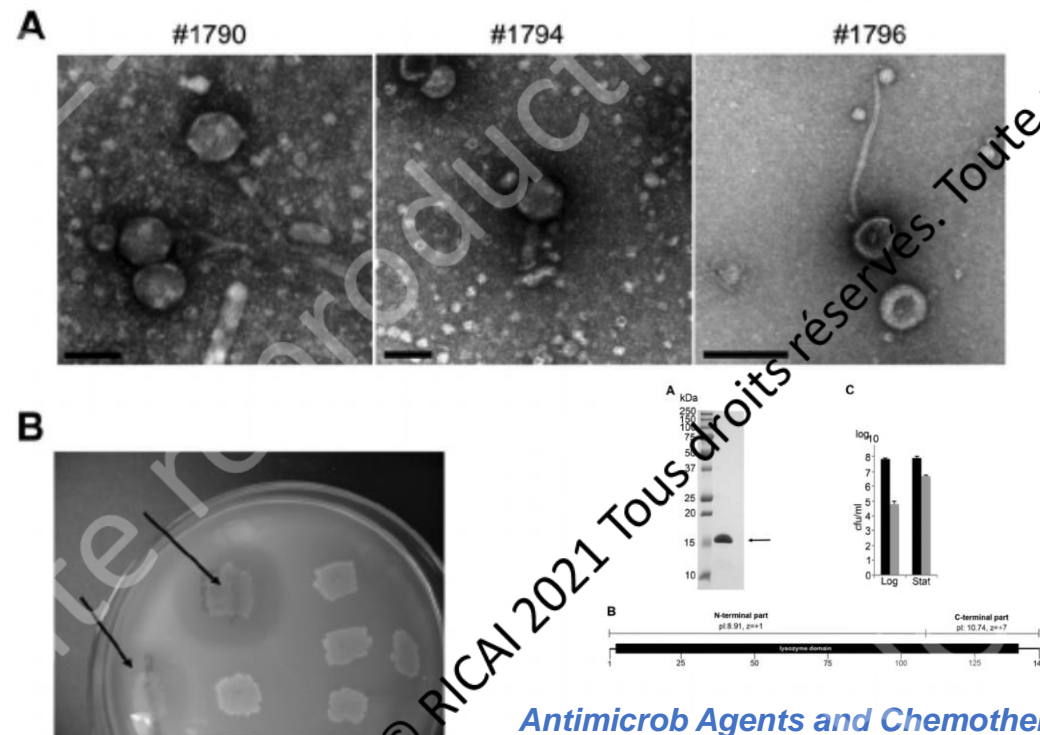


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Novel Phage Lysin Capable of Killing the Multidrug-Resistant Gram-Negative Bacterium *Acinetobacter baumannii* in a Mouse Bacteremia Model

Rolf Lood,^a Benjamin Y. Winer,^{a*} Adam J. Pelzek,^{a*} Roberto Diez-Martinez,^{a,b} Mya Thandar,^a Chad W. Euler,^{a,c} Raymond Schuch,^{a*} Vincent A. Fischetti^a

Laboratory of Bacterial Pathogenesis and Immunology, The Rockefeller University, New York, New York, USA^a; Molecular Microbiology and Infection Biology Department, Host-Parasite Interplay in Pneumococcal Infection Group, Centro de Investigaciones Biológicas Madrid, Spain^b; Department of Medical Laboratory Sciences, School of Arts and Sciences, Hunter College, CUNY, New York, New York, USA^c



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Conclusion



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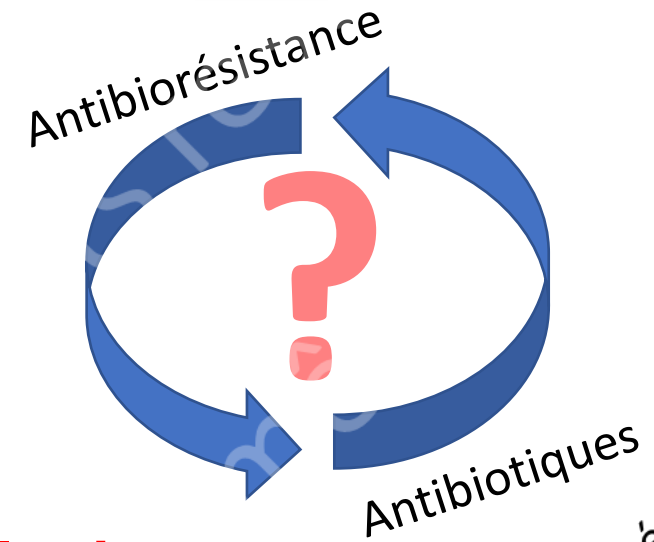
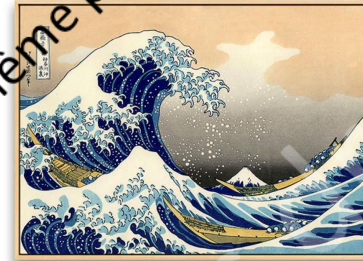


Nouveaux inhibiteurs de β -lactamase à disposition

- En combinaison avec des “vieux” antibiotiques

Nouveaux antibiotiques

- En combinaison avec des “vieux” inhibiteurs
- Céphalosporine siderophore

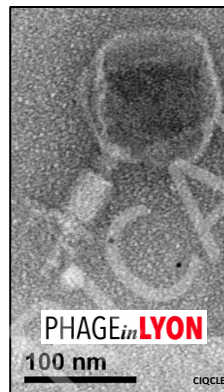


Evaluation clinique, AMM

Positionnement en fonction des pathogènes et des pathologies

Développement des thérapies non traditionnelles (definition OMS)

- Bactériophages
 - Industriels
 - Académiques
- Lysines de phages
- Groupe ESCMID



ESGNTA

European Society of Clinical Microbiology and Infectious Diseases

ESCMID STUDY GROUP FOR NON-TRADITIONAL ANTIBACTERIAL THERAPY



PHAGEin LYON



Centre International de Recherche en Infectiologie

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Lyon BJI Study group

Coordinator: Tristan Ferry

Infectious Diseases Specialists – Tristan Ferry, Florent Valour, Thomas Perpoint, Florence Ader, Sandrine Roux, Agathe Becker, Claire Triffault-Fillit, Anne Conrad, Cécile Poudroux, Pierre Chauvelot, Paul Chabert, Johanna Lippman, Evelyne Braun

Surgeons – Sébastien Lustig, Elvire Servien, Cécile Batailler, Stanislas Gunst, Axel Schmidt, Elliot Sappey-Marinier, Quentin Ode, Michel-Henry Fessy, Anthony Viste, Jean-Luc Besse, Philippe Chaudier, Lucie Louboutin, Adrien Van Haecke, Marcelle Mercier, Vincent Belgaid, Aram Gazarian, Arnaud Walch, Antoine Bertani, Frédéric Rongieras, Sébastien Martres, Franck Trouillet, Cédric Barrey, Ali Mojallal, Sophie Brosset, Camille Hanriat, Hélène Person, Philippe Céruse, Carine Fuchsmann, Arnaud Gleizer,

Anesthesiologists – Frédéric Aubrun, Mikhail Dziadzko, Caroline Macabéo, Dana Patrascu;

Microbiologists – Frederic Laurent, Laetitia Beraud, Tiphaine Roussel-Gaillard, Céline Dupieux, Camille Kolenda, Jérôme Josse;

Imaging – Fabien Craighero, Loic Bousel, Jean-Baptiste Pialat, Isabelle Morelec;

PK/PD specialists – Michel Tod, Marie-Claude Gagnieu, Sylvain Goutelle;

Clinical research assistant and database manager – Eugénie Mabrut

PHAGEⁱⁿ LYON

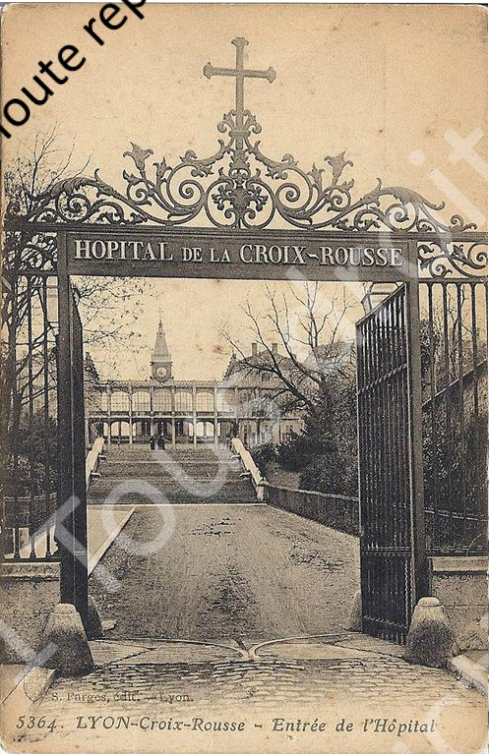


Coordinators: Tristan Ferry and Frédéric Laurent

Mathieu Medina, Camille kolenda, Floriane Launay, Melanie Bonhomme, Leslie Blazere, Tiphaine Legendre, Eline Terrazzoni, Fabrice Pirot, Camille Merienne, Samira Filali, Benjamine Lapras, Gilles Leboucher, Thomas Briot.



Croix-Rousse Hospital



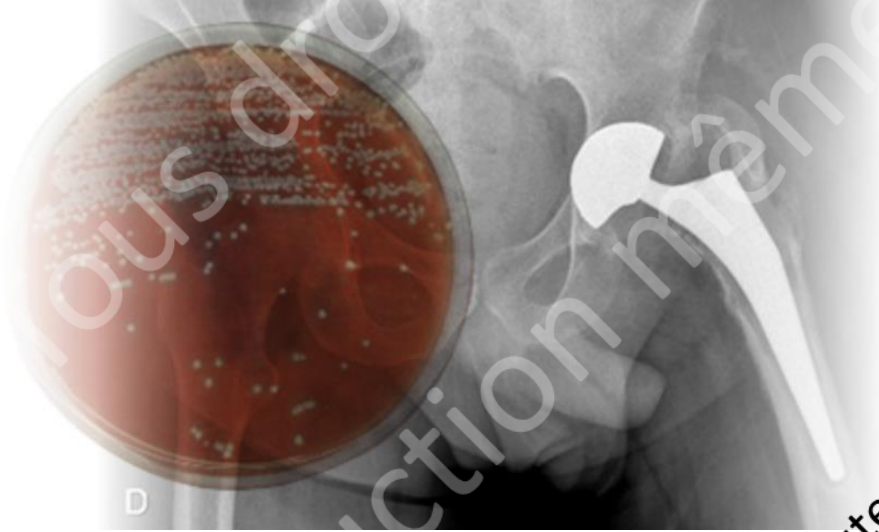
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Lyon 1



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