

38^{ème} Réunion Interdisciplinaire de Chimiothérapie Anti-Infectieuse

Paris - Mardi 18 décembre 2018

Antibiotiques à large spectre : quel impact sur le microbiote intestinal?

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Conflits d'intérêt

MSD

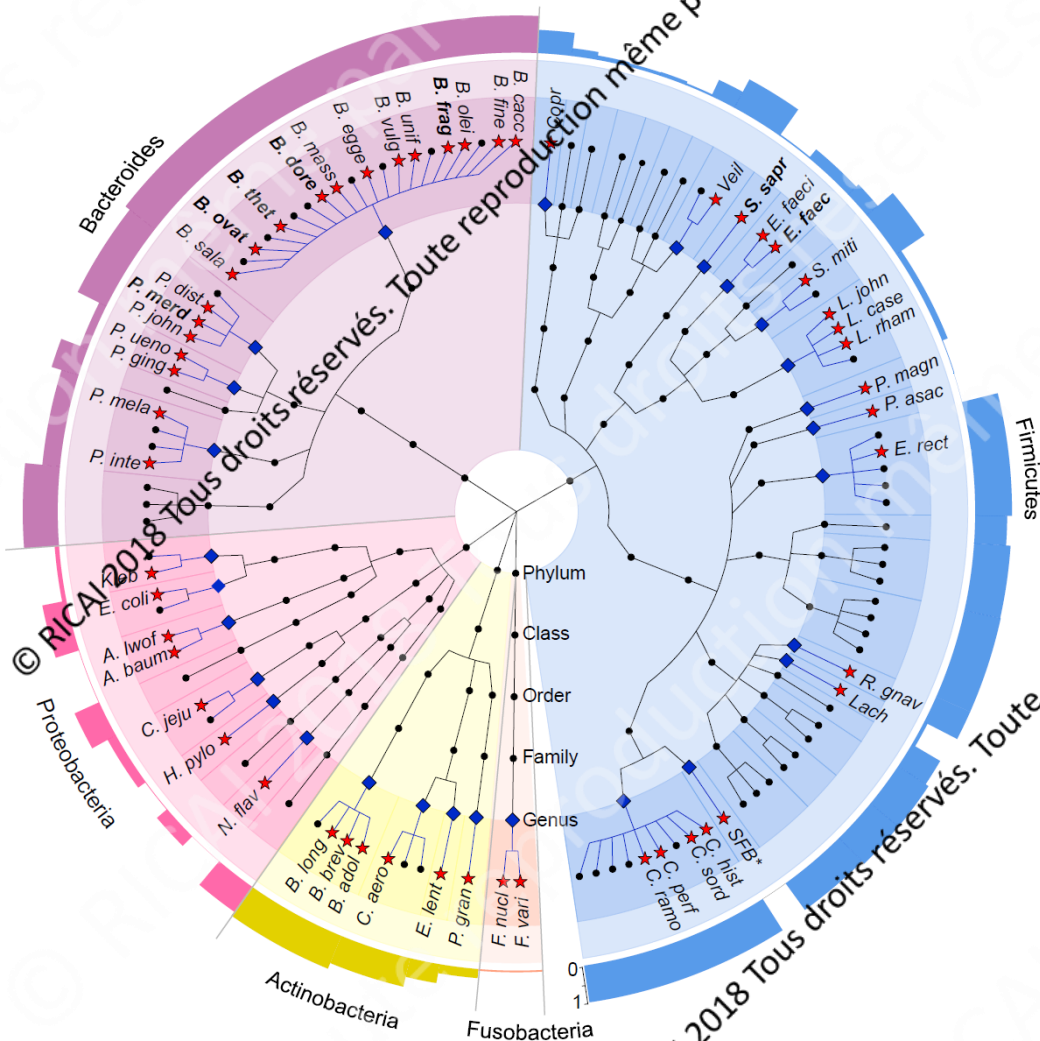
Pfizer

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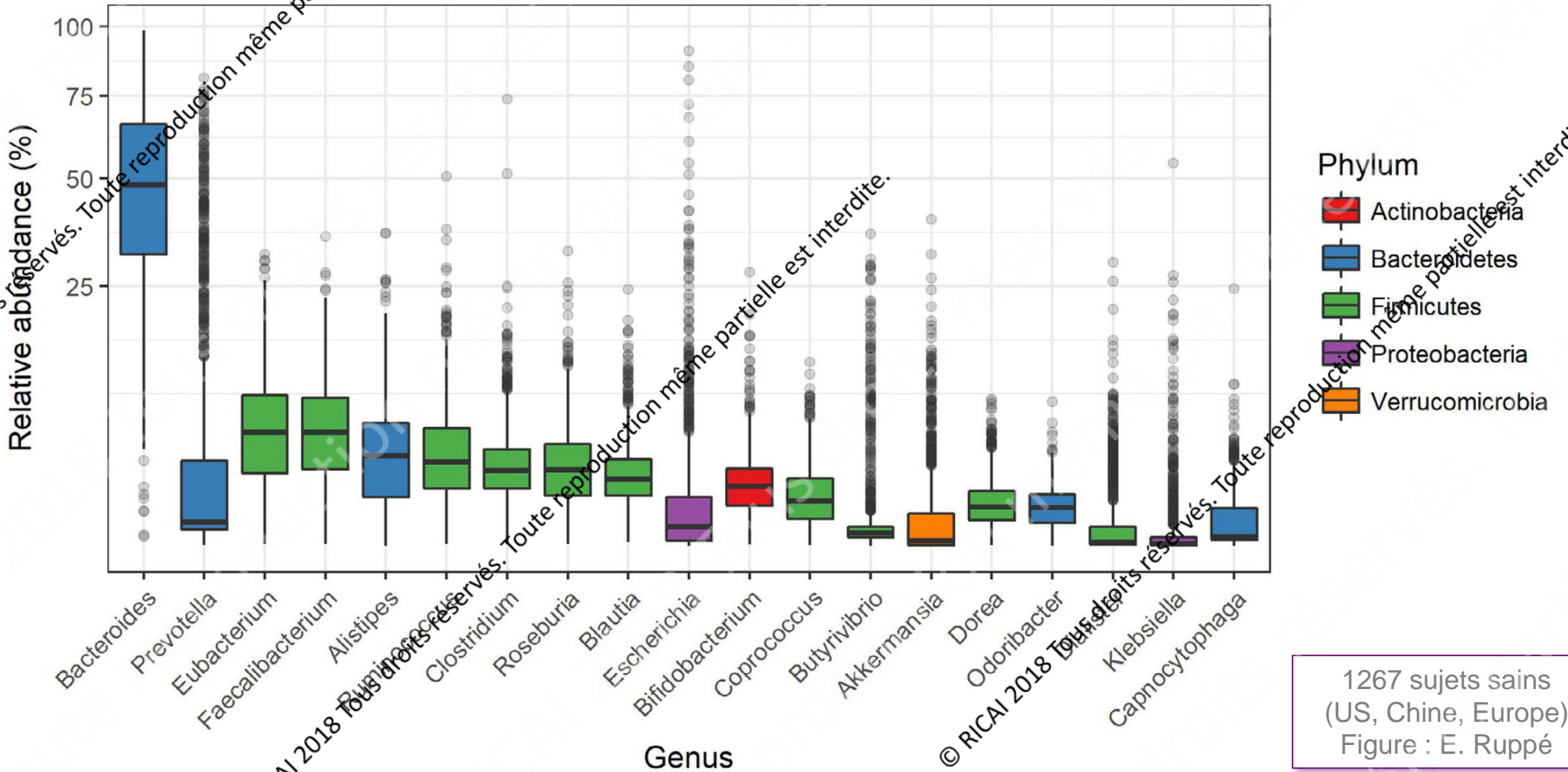
Microbiote intestinal



- Environ 10^{13} microorganismes (bactéries, archéobactéries, virus, levures, protozoaires)
- Rôles physiologiques multiples / fonctions symbiotiques (métaboliques, anti-inflammatoires, immunitaires)
- **Composition : variabilité inter- et intra-individuelle** (facteurs génétiques, environnement, alimentation)
- Côlon : > 90% de bactéries anaérobies non cultivables
- **Phyla prédominants : Firmicutes & Bacteroidetes**

Sender et al. *PLoS Biol* 2016; 14: e1002533
Li et al. *Nat Biotechnol* 2014; 32: 834-841
Feng et al. *Front Microbiol* 2018; 9: 151

Microbiote intestinal



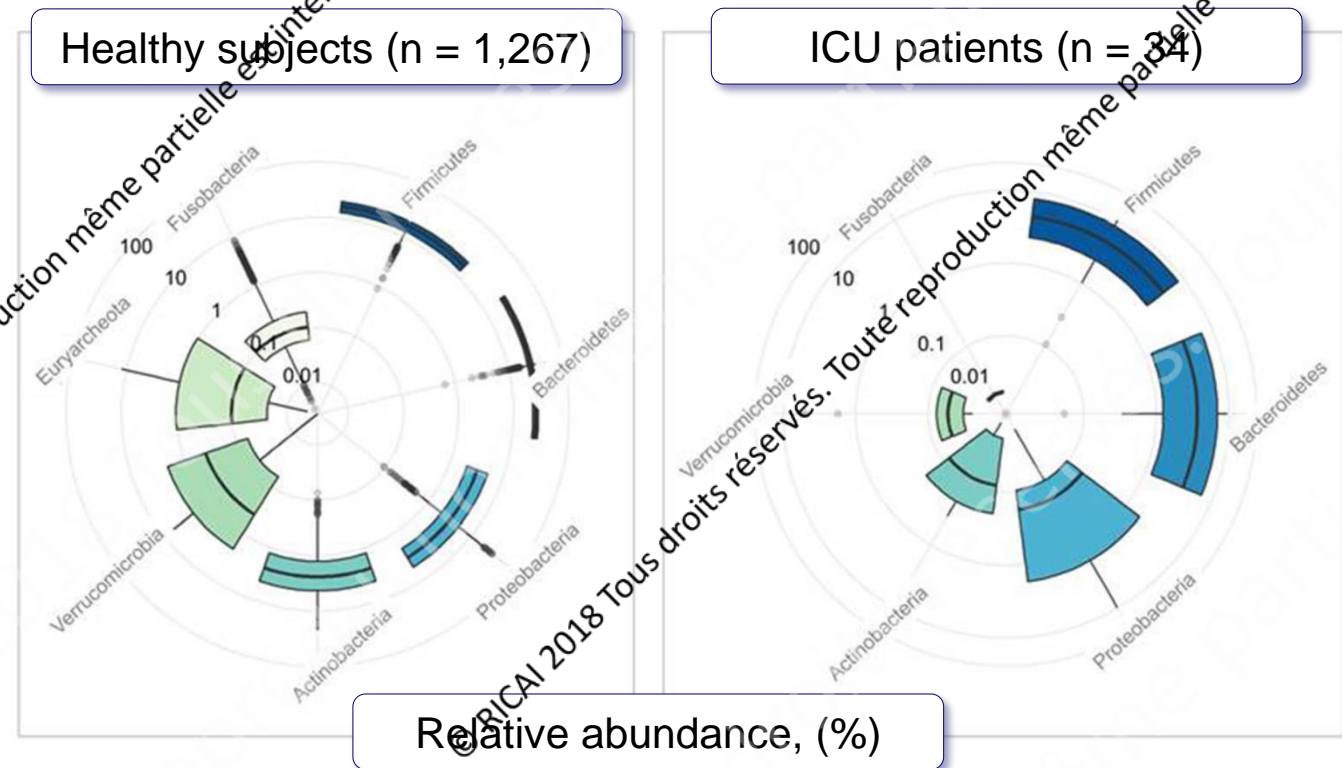
The gut microbiota of critically ill patients: first steps in an unexplored world

Étienne Ruppé^{1,2}, Thiago Lisboa⁴ and François Barbier^{5*}

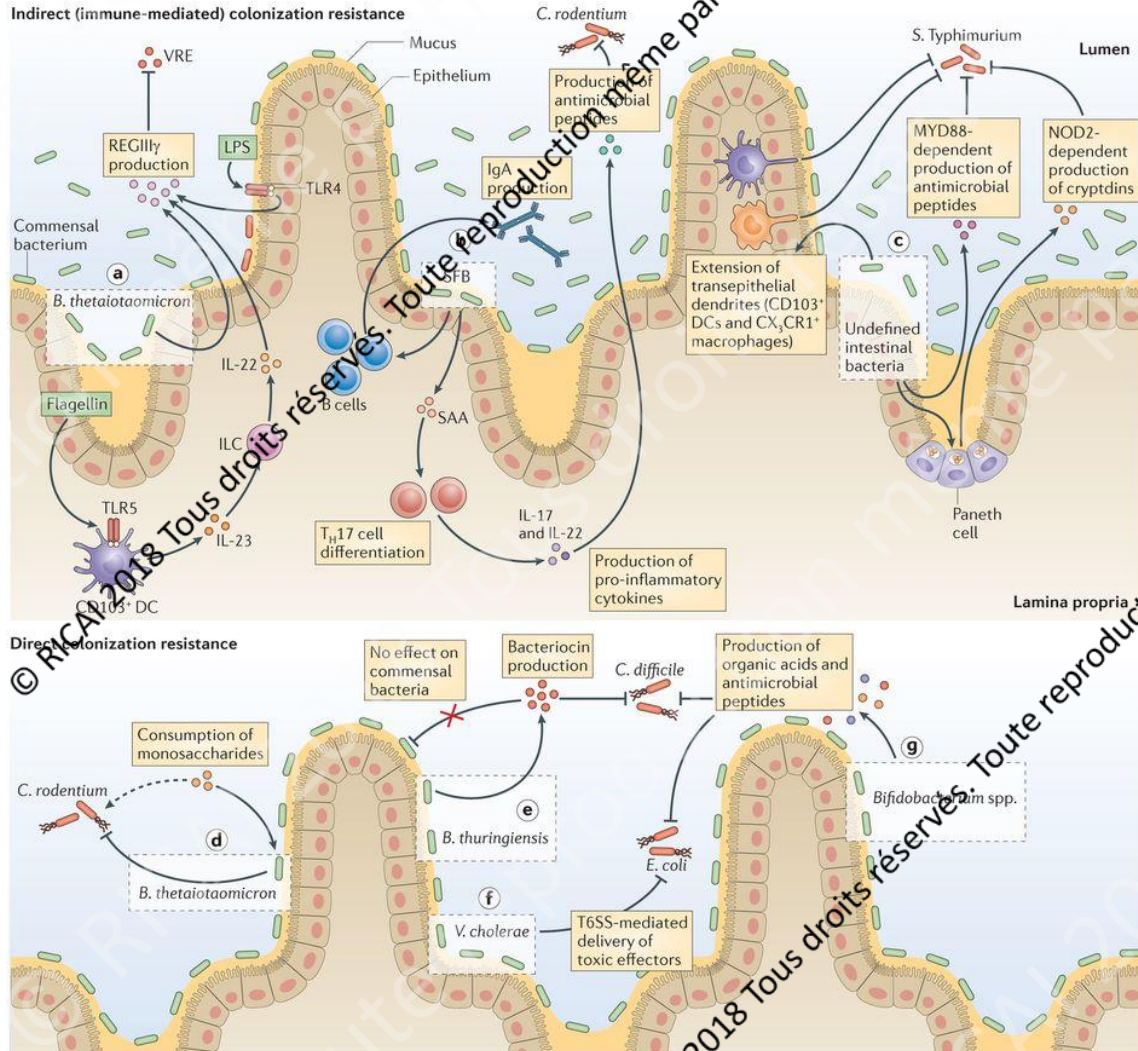
Intensive Care Med (2018) 44:1561–1564

Altérations profondes et précoces du microbiote intestinal chez les patients de réanimation

- **Perte de diversité**
- Raréfaction ou disparition de genres potentiellement « symbiotiques »
(ex: *Faecalibacterium*, *Ruminococcus*)
- Augmentation de l'abondance relative voire dominance de genres pathogènes
(ex : *Enterococcus*, Enterobacteriaceae)
- **Impact pronostique ?**



Microbiote intestinal & résistance à la colonisation



- « Antagonisme » entre flore commensale anaérobie et pathogènes exogènes
- Compétition nutritionnelle ou induction d'une réponse immunitaire ciblée
- Spécificité espèce commensale / pathogène, ex :
 - ✓ *Clostridium bolteae* + *Blautia producta* versus *Enterococcus* / ERV
 - ✓ *Clostridium scindens* versus *Clostridium difficile*

Buffie & Pamer. *Nature Reviews Immunology* 2013; 13: 790-801

Buffie et al. *Nature* 2015; 517:205-208

Caballero et al. *Cell Host Microbe* 2017; 21: 592-602

Long-term impact of oral vancomycin, ciprofloxacin and metronidazole on the gut microbiota in healthy humans

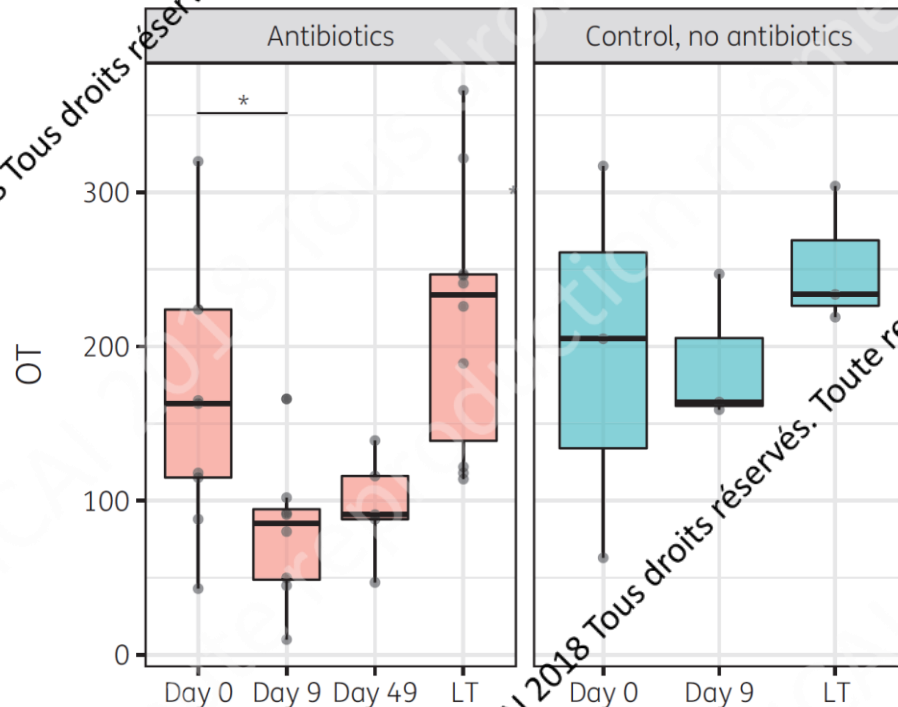


JAC 2018 (e-pub)

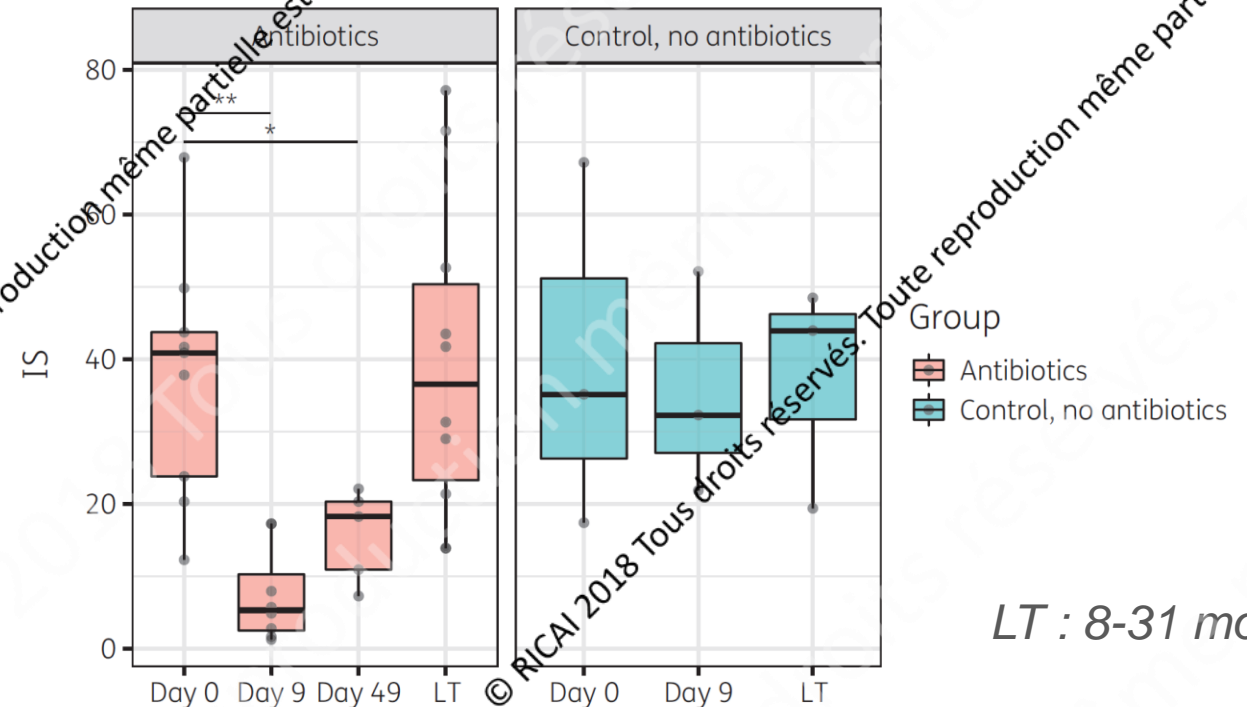
Bastiaan W. Haak^{1*}, Jacqueline M. Lankelma¹, Floor Hugenholtz¹, Clara Belzer²,
Willem M. de Vos³ and W. Joost Wiersinga^{1,4}

13 volontaires sains traités par vancomycine/ciprofloxacine/métronidazole pendant 7 jours (n = 10)
ou non traité (n = 3) / Analyse du microbiote intestinal par *RNA r16S profiling*

(a) Richness (OT)



(b) Microbiota diversity (IS)



LT : 8-31 months

Long-term impact of oral vancomycin, ciprofloxacin and metronidazole on the gut microbiota in healthy humans

Bastiaan W. Haak^{1*}, Jacqueline M. Lankelma¹, Floor Hugenholtz¹, Clara Belzer², Willem M. de Vos³ and W. Joost Wiersinga^{1,4}

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13 volontaires sains traités par vancomycine/ciprofloxacine/métronidazole pendant 7 jours (n = 10) ou non traité (n = 3) / Analyse du microbiote intestinal par RNA r16S profiling



Broad- and narrow-spectrum antibiotics: an unhelpful categorization

Clin Microbiol Infect 1997; 3: 395–396

« The expression 'broad-spectrum antibiotic' was used in the mid-1950s, when the bacterial spectrum of chloramphenicol and the first tetracyclines could be strikingly opposed to the narrow spectrum of activities of penicillin G, and streptomycin. In the 1960s, aminopenicillins, then ureidopenicillins, became the broad-spectrum penicillins in comparison with penicillin G. Until then, the quality of being broad spectrum or narrow spectrum was given to an antibiotic only when referring to a comparator. Later, the reference to a comparator was omitted, and broad and narrow lost their relativities and became independent characteristics of a compound, used with different meaning and often improperly. Never (...) was any effort made to define those words. »

Jacques Acar

Elaboration of a consensual definition of de-escalation allowing a ranking of β -lactams

Clin Microbiol Infect 2015; **21**: 649.e1–649.e10

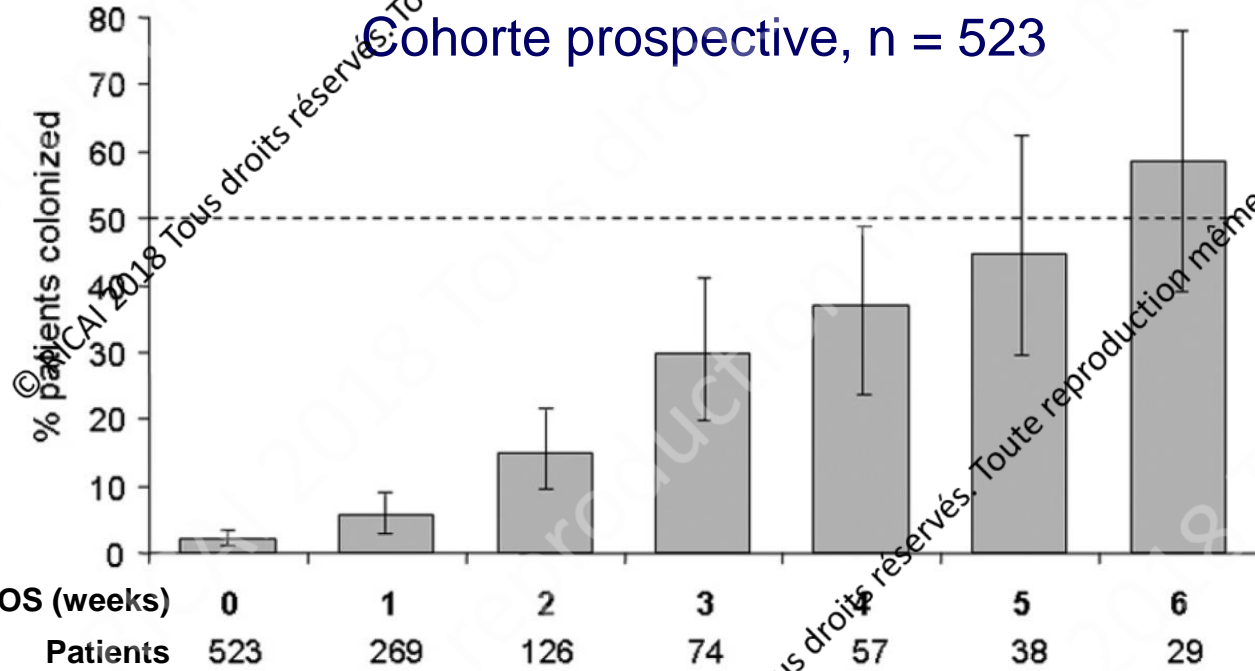
E. Weiss¹, J.-R. Zahar², P. Lesprit³, E. Ruppe⁴, M. Leone⁵, J. Chastre⁶, J.-C. Lucet⁷, C. Paugam-Burtz¹, C. Brun-Buisson⁸ and J.-F. Timsit⁹, on behalf of the 'De-escalation' Study Group

Classification des bêta-lactamines selon leur spectre clinique et **leur impact écologique** (28 experts français, méthode DELPHI)

Rank	Molecule(s)	Similar response rate (%) ^a	Consensus reaching round number ^b
1	Amoxicillin	100	2
2	Amoxicillin + Clavulanic Acid	88	3
3	Third-generation cephalosporin Ureido/carboxy-penicillin	81	3
4	Piperacilin + Tazobactam Ticarcilin + Clavulanic Acid Fourth-generation cephalosporin, Antipseudomonal third-generation cephalosporin	71	4
5	Ertapenem	81	3
6	Imipenem Meropenem Doripenem	85	2

Emergence of Imipenem-Resistant Gram-Negative Bacilli in Intestinal Flora of Intensive Care Patients

Laurence Armand-Lefèvre,^{a,b} Cécile Angebault,^{a,b} François Barbier,^{b,c} Emilie Hamelet,^a Gilles Defrance,^a Etienne Ruppé,^{a,b} Régis Bronchard,^d Raphaël Lepeule,^b Jean-Christophe Lucet,^e Assiya El Mniai,^a Michel Wolff,^c Philippe Montravers,^d Patrick Plésiat,^f Antoine Andremont^{a,b}



Species	No. of strains	Resistance mechanisms ^a	
		Enzymes	Other
<i>P. aeruginosa</i>	19		OprD-
	6	AmpC++	OprD-
	4		OprD- MexAB efflux ++
	2	AmpC++	OprD- MexAB efflux ++
	1	GES-9	OprD-
	4	VIM-2	
<i>Enterobacteriaceae</i>			
<i>K. pneumoniae</i>	2	DHA-1	OMP-
	1	TEM-1 CTX-M15	NP
<i>E. aerogenes</i>	1	TEM-4 AmpC++	OMP-
<i>E. cloacae</i>	1	SHV-12 AmpC++	OMP-
<i>H. alvei</i>	1	AmpC++	NP
<i>A. baumannii</i>	2		
<i>S. maltophilia</i>	12	Wild type	

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TABLE 2 Univariate and multivariate analysis of risk factors associated with intestinal colonization of imipenem-resistant Gram-negative bacilli^a

Characteristic or outcome	No. of individuals or parameter value (%, unless range is specified)		Univariate OR ^b	Univariate P ^c	Multivariate OR ^d
	Carrier patients (n = 36)	Controls (n = 36)			
Antibiotic treatments					
Exposure time to antibiotics, days [median (range)]	11.5 (0–51)	9.0 (0–37)		0.84	
Penicillin exposure	8 (22.2)	16 (44.4)	0.4 (0.1–1.1)	0.08	0.3 (0.1–0.8)
Penicillin and β-lactamase inhibitor exposure	17 (47.2)	20 (55.6)	0.7 (0.3–2.0)	0.64	
Cephalosporin exposure	20 (55.6)	17 (47.2)	1.4 (0.5–3.9)	0.64	
Imipenem exposure	28 (77.8)	14 (38.9)	5.4 (1.8–17.8)	<0.01	
Days of imipenem exposure				<0.01	
0	8 (22.2)	22 (61.1)	1.0		1.0
1 to 3	10 (27.8)	6 (16.7)	4.4 (1.1–20.5)		5.9 (1.5–25.7)
4 to 21	18 (50.0)	8 (22.2)	6.0 (1.7–23.3)		7.8 (2.4–29.8)
Fluoroquinolone exposure	9 (25.0)	8 (22.2)	1.2 (0.3–4.0)	1.00	
Aminoglycoside exposure	25 (69.4)	21 (58.3)	1.6 (0.6–4.8)	0.46	
Glycopeptide exposure	20 (55.6)	11 (30.6)	2.8 (1.0–8.4)	0.06	
Metronidazole exposure	5 (13.9)	6 (16.7)	0.8 (0.2–3.6)	1.00	

Increased GVHD-related mortality with broad-spectrum antibiotic use after allogeneic hematopoietic stem cell transplantation in human patients and mice

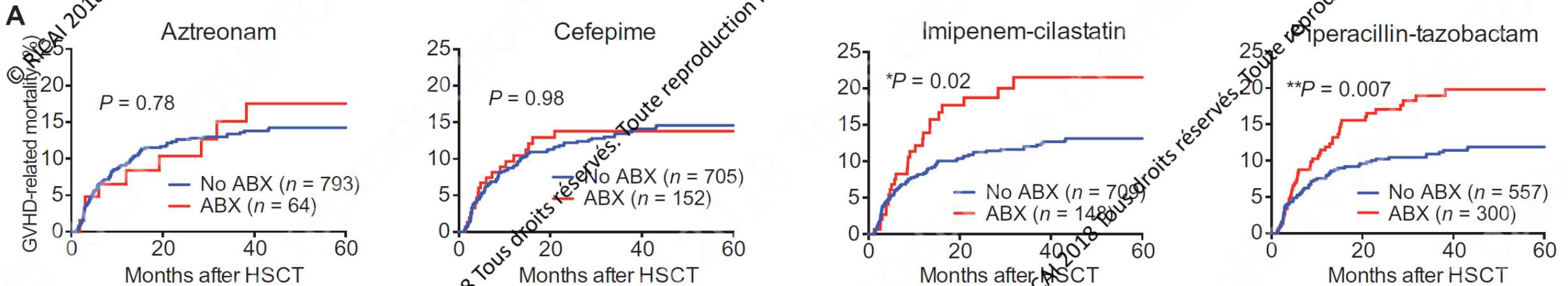


www.ScienceTranslationalMedicine.org 18 May 2016 Vol 8 Issue 339

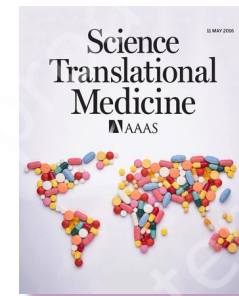
Retrospective cohort of 857 allo-HSCT recipients

Treatment of neutropenic fever with imipenem or piperacillin-tazobactam : association with increased GVHD-related mortality at 5 years

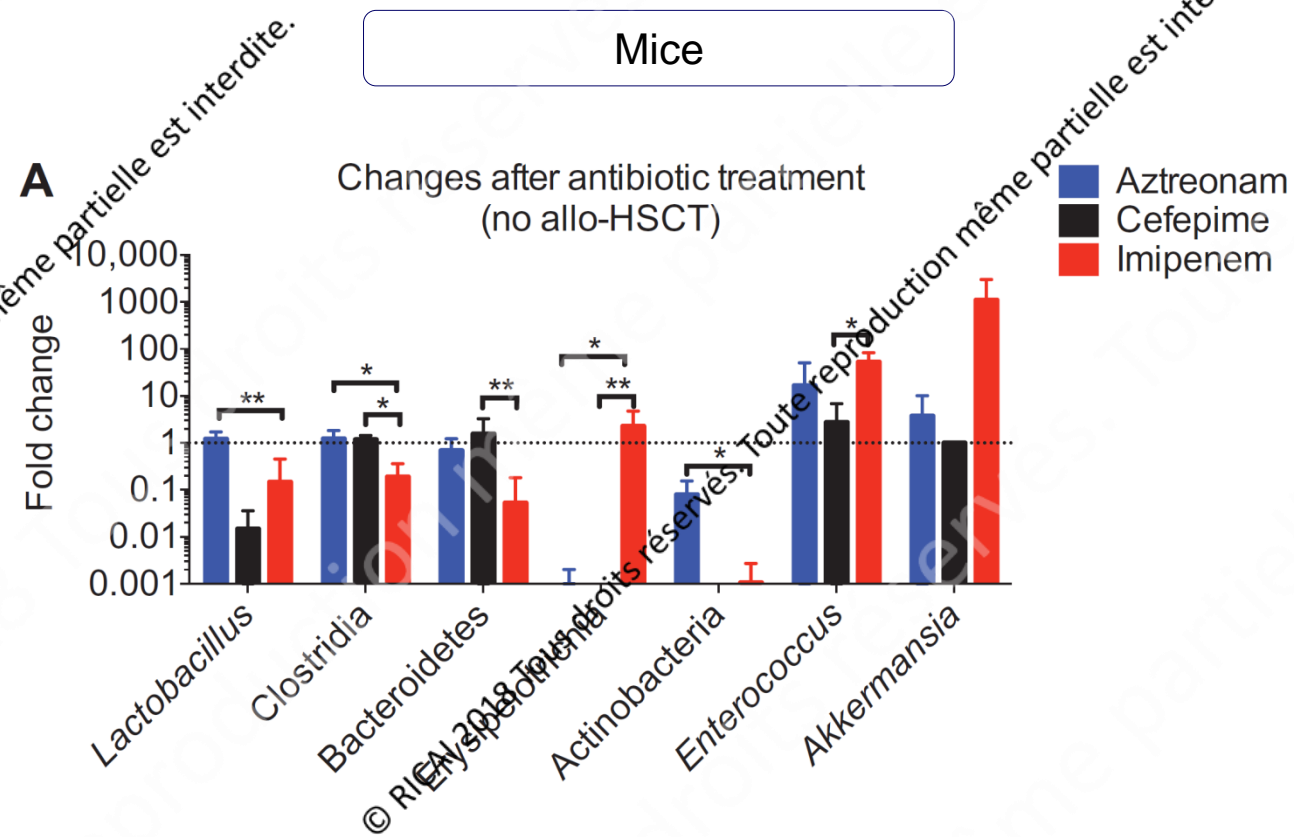
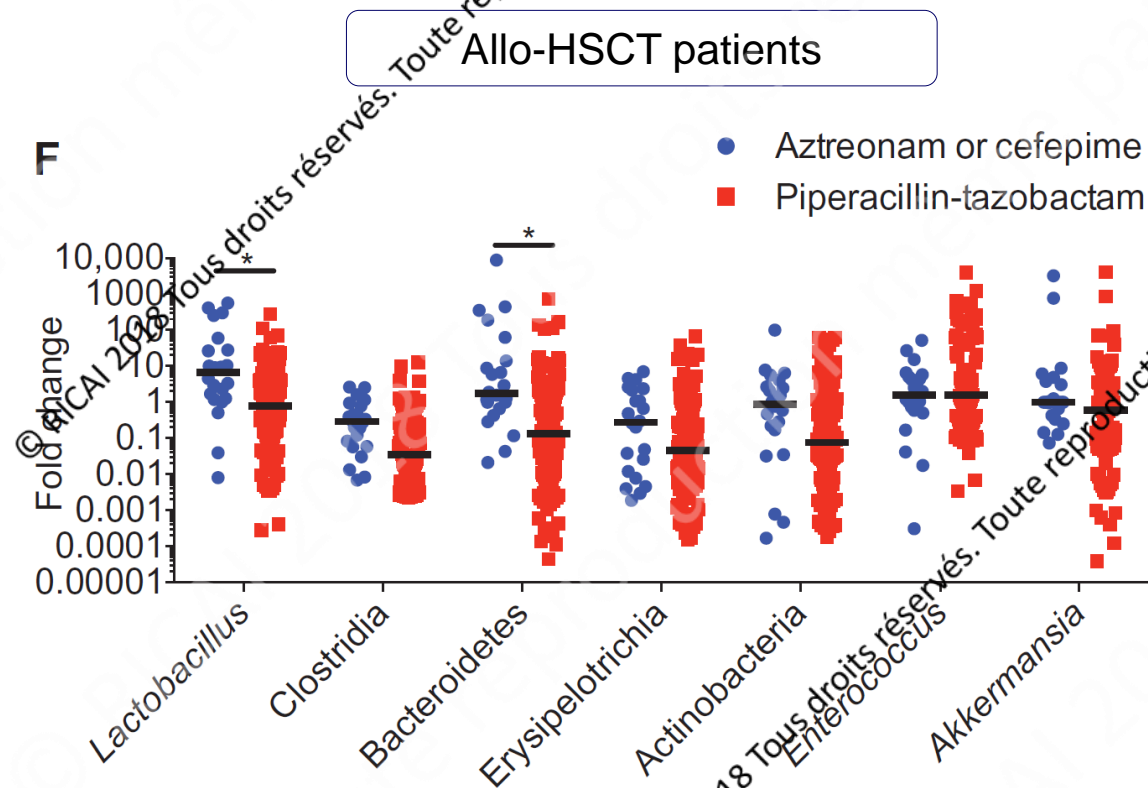
(use of aztreonam or cefepime were not associated with GVHD-related mortality)



Increased GVHD-related mortality with broad-spectrum antibiotic use after allogeneic hematopoietic stem cell transplantation in human patients and mice



www.ScienceTranslationalMedicine.org 18 May 2016 Vol 8 Issue 339



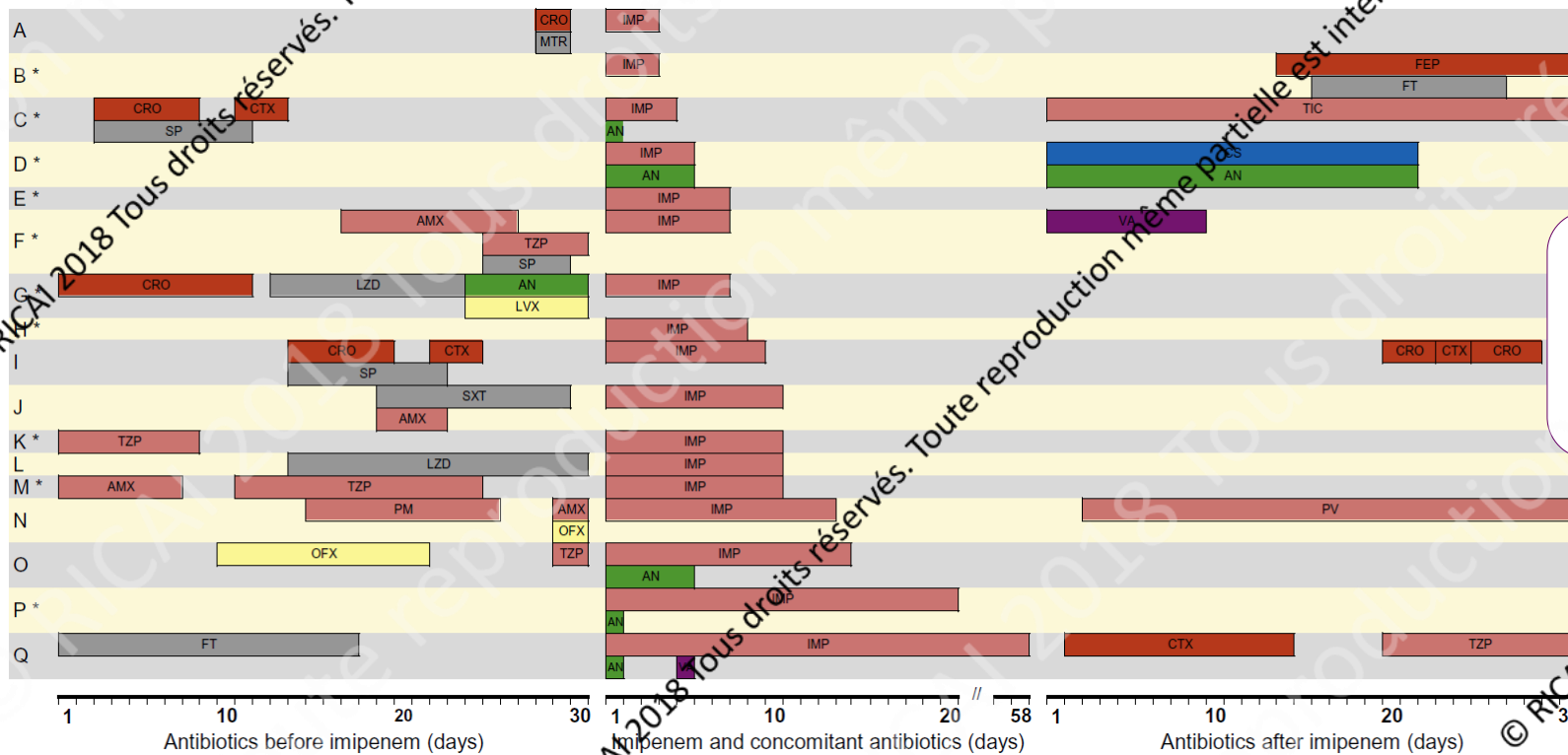
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Unexpected persistence of extended-spectrum β -lactamase-producing Enterobacteriaceae in the faecal microbiota of hospitalised patients treated with imipenem

N. Grall ^{a,b,c,*}, V. Lazarevic ^{d,1}, N. Gaïa ^d, C. Couignal ^{a,b,e}, C. Laouénan ^{a,b,e}, E. Ilic-Habensus ^f, I. Wieder ^c, P. Plesiat ^g, C. Angebault ^{h,i}, M.E. Bougnoux ^{h,i}, L. Armand-Lefevre ^{a,b,c}, A. Andremont ^{a,b,c}, X. Duval ^{a,b,f}, J. Schrenzel ^{d,j}

International Journal of Antimicrobial Agents 50 (2017) 81–87

17 patients traités par imipénème, dont 12 porteurs d'EBLSE (CMI d'imipénème, 0,09-1 mg/l)



Prélèvement de flore colique :

J0, J3, Jfin, J15, J30

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17 patients traités par imipénème, dont 12 porteurs d'EBLSE (CMI d'imipénème, 0,09-1 mg/l)

Colonisation rates by imipenem-susceptible ESBL-E remained stable over time, suggesting that **imipenem intestinal concentrations were very low.**

Carriage rates of carbapenem-resistant GNB were also stable over time.

Metagenomics showed no global effect of imipenem on bacterial taxonomic profiles at the sequencing depth.

IR-GNB

Candida spp

D0

D3

Dend

D15

D30

Bowel colonization with resistant gram-negative bacilli after antimicrobial therapy of intra-abdominal infections: observations from two randomized comparative clinical trials of ertapenem therapy

OASIS-I : ertapénème *versus* pipéracilline-tazobactam (durée médiane de traitement : 6 jours)

DiNubile et al. *Eur J Clin Microbiol Infect Dis* 2005; 24: 443-449

Table 2 Proportion of assessable patients with resistant gram-negative bacilli isolated from rectal swabs at different time points during OASIS I by treatment group

Organism recovered from assessable patients ^a	Ertapenem group			Piperacillin-tazobactam group		
	Baseline (n=162)	End of therapy (n=155)	2 weeks post-therapy (n=133)	Baseline (n=160)	End of therapy (n=156)	2 weeks post-therapy (n=133)
Piperacillin-tazobactam-resistant <i>Enterobacteriaceae</i> , n (%)	1 (0.6)	2 (1.3)	3 (2.3)	1 (0.6)	19 (12.2) ^b	6 (4.5)
Ertapenem-resistant <i>Enterobacteriaceae</i> , n (%)	0 (0)	1 (0.6)	0 (0)	0 (0)	3 (1.9)	0 (0)
ESBL-producing <i>Enterobacteriaceae</i> , n (%)	1 (0.6)	0 (0)	1 (0.8) ^[1]	1 (0.6)	4 (2.6)	1 (0.8)
Imipenem-resistant <i>P. aeruginosa</i> , n (%)	0 (0)	0 (0)	0 (0)	2 (1.3)	2 (1.3) ^[1]	0 (0)
Piperacillin-tazobactam-resistant <i>P. aeruginosa</i> , n (%)	1 (0.6)	3 (1.9) ^[1]	0 (0)	0 (0)	1 (0.6)	0 (0)

Bowel colonization with resistant gram-negative bacilli after antimicrobial therapy of intra-abdominal infections: observations from two randomized comparative clinical trials of ertapenem therapy

OASIS-II : ertapénème *versus* ceftriaxone/métronidazole (durée médiane de traitement : 6 jours)

DiNubile et al. *Eur J Clin Microbiol Infect Dis* 2005; 24: 443-449

Table 3 Proportion of assessable patients with resistant gram-negative bacilli isolated from rectal swabs at different time points during OASIS II by treatment group

Organism recovered from assessable patients ^a	Ertapenem treatment			Ceftriaxone/metronidazole group		
	Baseline (n=201)	End of therapy (n=196)	2 weeks post-therapy (n=182)	Baseline (n=195)	End of therapy (n=193)	2 weeks post-therapy (n=174)
Ceftriaxone-resistant <i>Enterobacteriaceae</i> ^b , n (%)	9 (4.5)	3 (1.5) ^[1]	5 (2.7)	5 (2.6)	33 (17.1) ^{[3]*}	39 (22.4) ^{†, [1]}
Ertapenem-resistant <i>Enterobacteriaceae</i> , n (%)	1 (0.5)	1 (0.5) ^{[1]*}	0 (0) [†]	0 (0)	1 (0.5)	0 (0)
ESBL-producing <i>Enterobacteriaceae</i> , n (%)	8 (4.0)	0 (0) ^{**}	4 (2.2) ^{††}	4 (2.1)	18 (9.3) ^{[3]**}	30 (17.2) ^{††, [1]}
Imipenem-resistant <i>P. aeruginosa</i> , n (%)	0 (0)	2 (1.0)	1 (0.5)	0 (0)	0 (0)	0 (0)

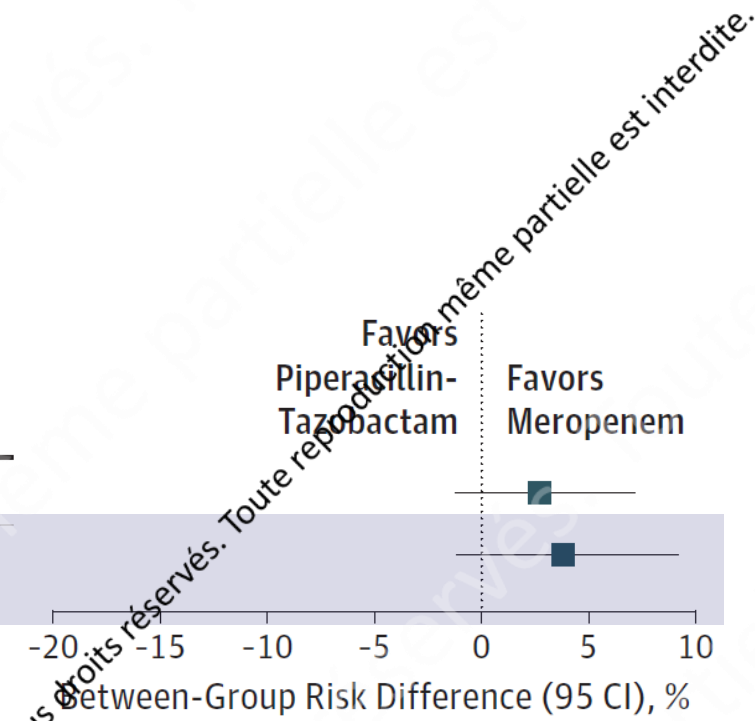
Effect of Piperacillin-Tazobactam vs Meropenem on 30-Day Mortality for Patients With *E coli* or *Klebsiella pneumoniae* Bloodstream Infection and Ceftriaxone Resistance

A Randomized Clinical Trial



JAMA. 2018;320(10):984-994.

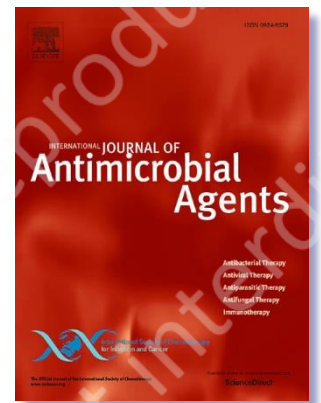
Measure of Failure	Patients Meeting End Point, No./Total No. (%)		Between-Group Difference (95% CI)
	Piperacillin-Tazobactam	Meropenem	
Microbiological relapse	9/187 (4.8)	4/191 (2.1)	2.7 (-1.1 to 7.1)
Secondary infection with multiresistant organism or <i>Clostridium difficile</i>	15/187 (8.0) ^b	8/191 (4.2) ^c	3.8 (-1.1 to 9.1)



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Carbapenems and alternative β -lactams for the treatment of infections due to extended-spectrum β -lactamase-producing Enterobacteriaceae: What impact on intestinal colonisation resistance?

Paul-Louis Woerther^{a,b,*}, Raphaël Lepeule^{a,1}, Charles Burdet^{c,d,e,1}, Jean-Winoc Decousser^{a,b}, Étienne Ruppé^{c,d,f}, François Barbier^g



Int J Antimicrob Agents 2018 (e-pub)

Facteurs de risque d'acquisition d'un portage intestinal d'EPC (cohortes prospectives & études cas-témoins)

Exposition aux carbapénèmes

- Expositions aux pénicillines, C3G/C4G, BL-BLI, fluoroquinolones, glycopeptides
- Durée de séjour
- Procédures invasives
- Pression de colonisation

Papadimitriou-Olivgeris et al. *J Antimicrob Chemother* 2012; 67: 2976-81

Schwartz-Neiderman et al. *Infect Control Hosp Epidemiol* 2016; 37: 1219-25

Schwaber et al. *Antimicrob Agents Chemother* 2008; 52: 1028-33

Madueño et al. *Am J Infect Control* 2017; 45: 77-9

Ling et al. *Antimicrob Resist Infect Control* 2015; 4: 26

Zhao et al. *Am J Infect Control* 2014; 42: e61-4

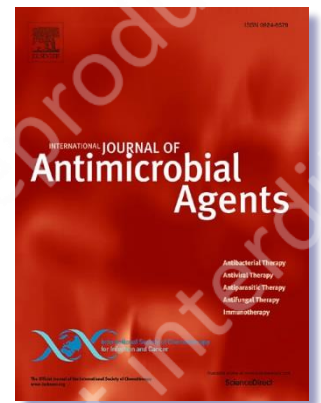
Gasink et al. *Infect Control Hosp Epidemiol* 2009; 30: 1180-5

Muggeo et al. *Antimicrob Chemother* 2017; 72: 1496-501

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- Expositions aux pénicillines, C3G/C4G, BL-BLI, fluoroquinolones, glycopeptides
- Durée de séjour
- Procédures invasives
- **Pression de colonisation**

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Muggeo et al. *J Antimicrob Chemother* 2017; 72: 1496-501

Keyvan Razazi
Lennie P. G. Derde
Marine Verachten
Patrick Legrand
Philippe Lesprit
Christian Brun-Buisson

Clinical impact and risk factors for colonization with extended-spectrum β -lactamase-producing bacteria in the intensive care unit

**Cohorte prospective, France,
2010-2011**

610 patients

Portage d'EBLSE à l'admission
en réanimation = 15%

Table 4 Adjusted odds ratio for ESBL acquisition among 212 patients staying in ICU for 5 days or more

Predictor	Odds ratio	[95 % CI]
Age \geq 75 years	6.3	[2.17–18.6]
Male gender	3.5	[1.03–11.7]
Colonization pressure ^a	1.3	[1.18–1.49]
3GC within past 3 months	4.8	[1.52–15.0]
B-lactam + inhibitor within past 3 months	3.5	[1.22–10.1]

Risk Factors and Outcomes for Intestinal Carriage of AmpC-Hyperproducing *Enterobacteriaceae* in Intensive Care Unit Patients

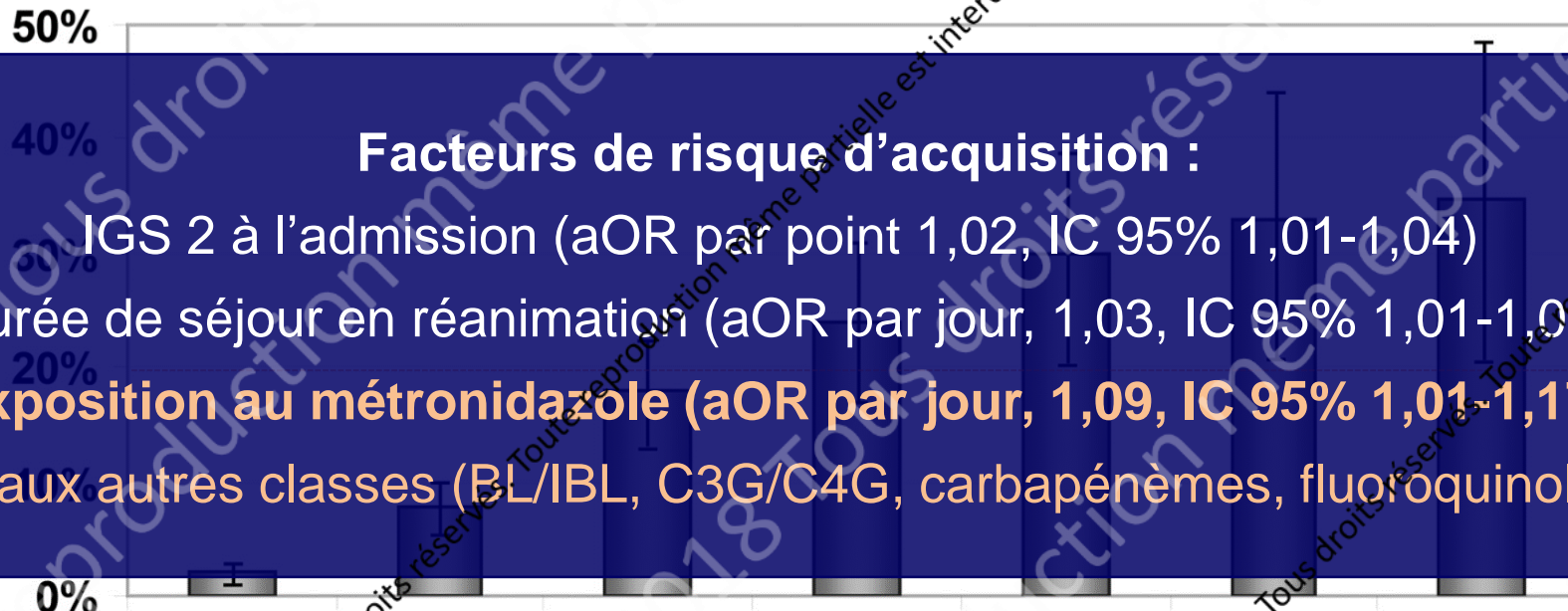


Simon Poignant,^a Jérôme Guinard,^b Aurélie Guignon,^b Laurent Bret,^b Didier-Marc Poisson,^b Thierry Boulain,^a François Barbier^a
 Medical Intensive Care Unit^a and Department of Microbiology,^b La Source Hospital, CHR Orléans, Orléans, France

March 2016 Volume 60 Number 3

Portage intestinal d'entérobactéries hyper-productrices de céphalosporinase AmpC

Patients de réanimation (n = 1209), dépistage par ER admission / hebdomadaire



Facteurs de risque d'acquisition :

IGS 2 à l'admission (aOR par point 1,02, IC 95% 1,01-1,04)

Durée de séjour en réanimation (aOR par jour, 1,03, IC 95% 1,01-1,04)

Exposition au métronidazole (aOR par jour, 1,09, IC 95% 1,01-1,17)

Exposition aux autres classes (BL/IBL, C3G/C4G, carbapénèmes, fluoroquinolones) : NS

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Patients de réanimation (n = 1209), dépistage par ER admission / hebdomadaire

Association entre consommation de métronidazole et incidence des infections acquises en réanimation à entérobactéries hyper-productrices d'AmpC

Fihman et al. *Int J Antimicrob Agents* 2015; 46: 518-25

Association entre exposition au métronidazole et acquisition d'un portage de *Pseudomonas aeruginosa* MR

Paranythiotou et al. *Clin Infect Dis* 2004; 38: 670-7

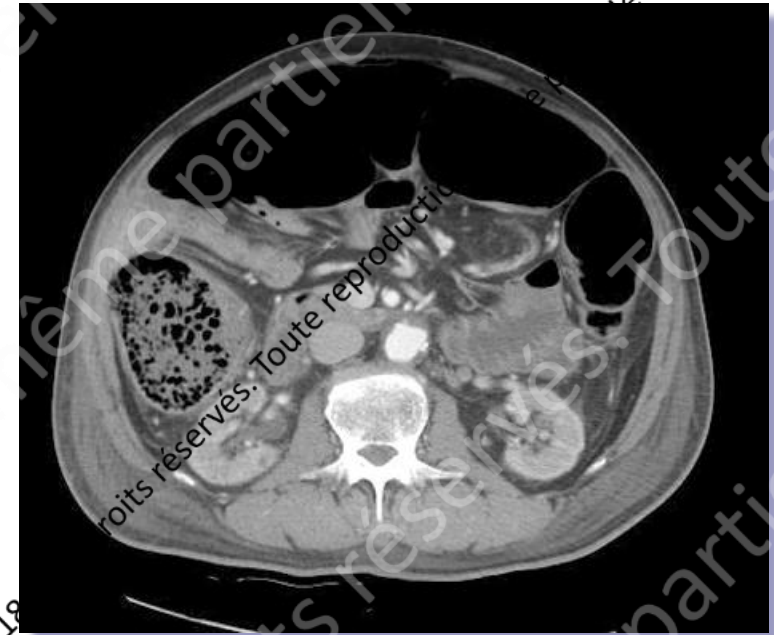
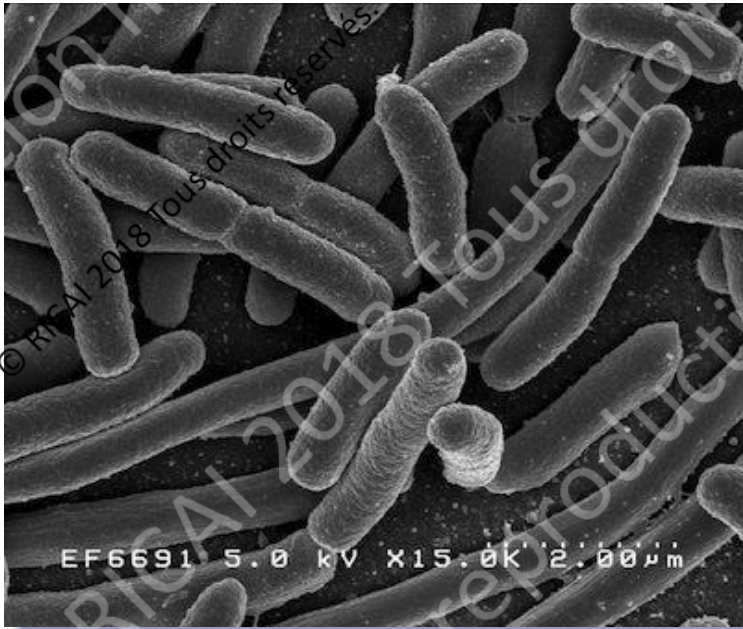
Cipriano Souza et al. *J Hosp Infect* 2008; 69: 402-3

	D0	D7	D14	D21	D28	D35	D42
Total, N	1,209	610	273	151	97	64	46
Carriers, n (%)	24 (2.0)	48 (7.9)	49 (17.9)	36 (23.8)	29 (29.9)	21 (32.8)	16 (34.8)

Antibiotics and hospital-acquired *Clostridium difficile* infection: update of systematic review and meta-analysis

Claudia Slimings^{1*} and Thomas V. Riley^{1,2}

J Antimicrob Chemother 2014; **69**: 881 – 891



Antibiotics and hospital-acquired *Clostridium difficile* infection: update of systematic review and meta-analysis

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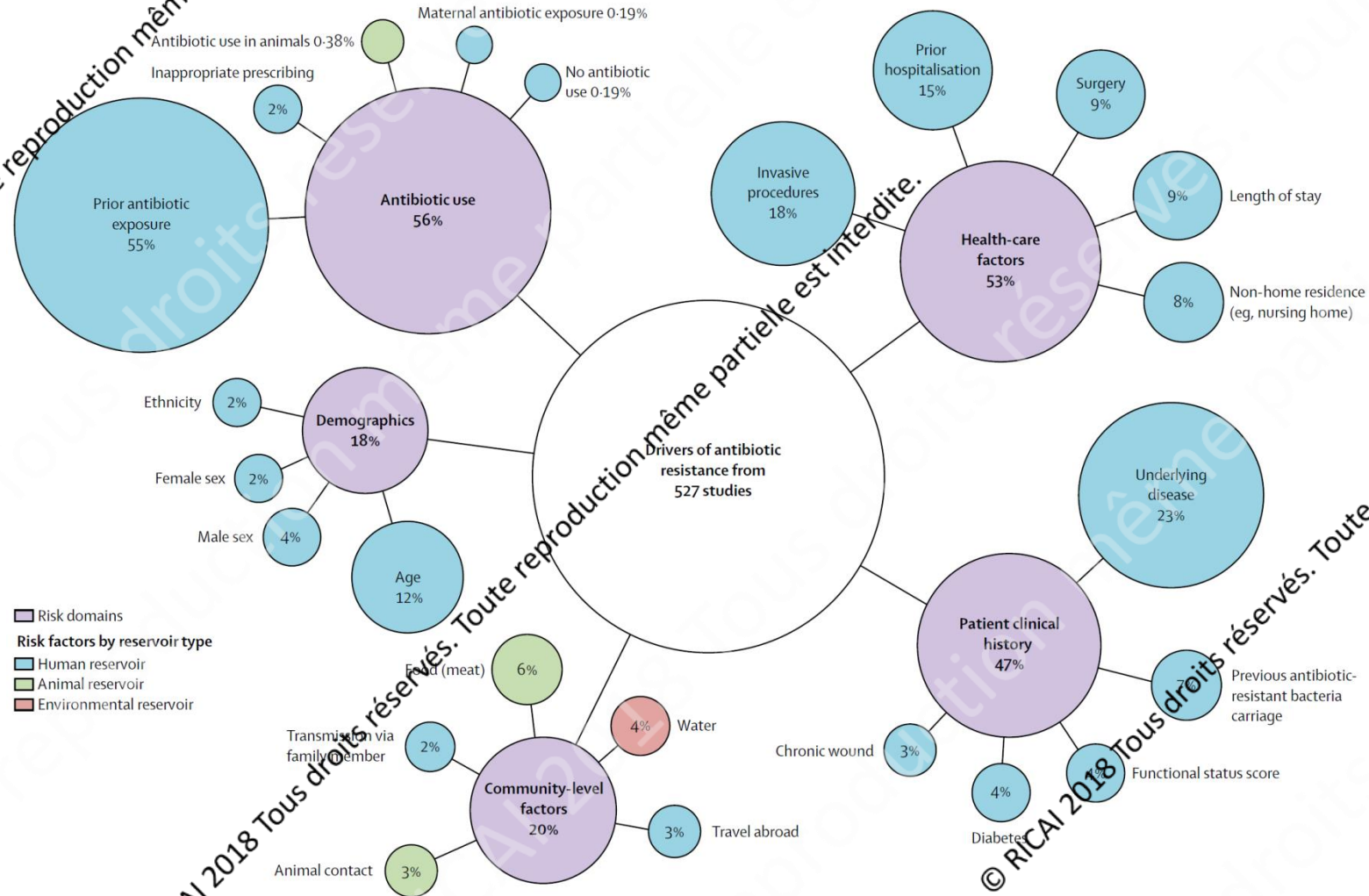
Impact de l'exposition aux antibiotiques sur le risque d'infection à *C. difficile*

Antibiotiques/classes	aOR (IC 95%)	Nb d'études	I ²
C3G	3,20 (1,80-5,71)	6	79%
Clindamycine	2,86 (2,04-4,02)	6	28%
C2G	2,23 (1,47-3,37)	6	48%
C4G	2,14 (1,30-3,52)	6	48%
Carbapénèmes	1,84 (1,26-2,68)	6	0%
Cotrimoxazole	1,78 (1,04-3,05)	5	70%
Fluoroquinolones	1,66 (1,17-2,35)	10	64%
Pénicillines (associations)	1,45 (1,05-2,02)	6	54%

Quantifying drivers of antibiotic resistance in humans: a systematic review

Anuja Chatterjee, Maryam Modarai, Nichola R Naylor, Sara E Boyd, Prat Atun, James Barlow, Alison H Holmes, Alan Johnson, Julie V Robotham

Lancet Infect Dis 2018;
18: e368-78

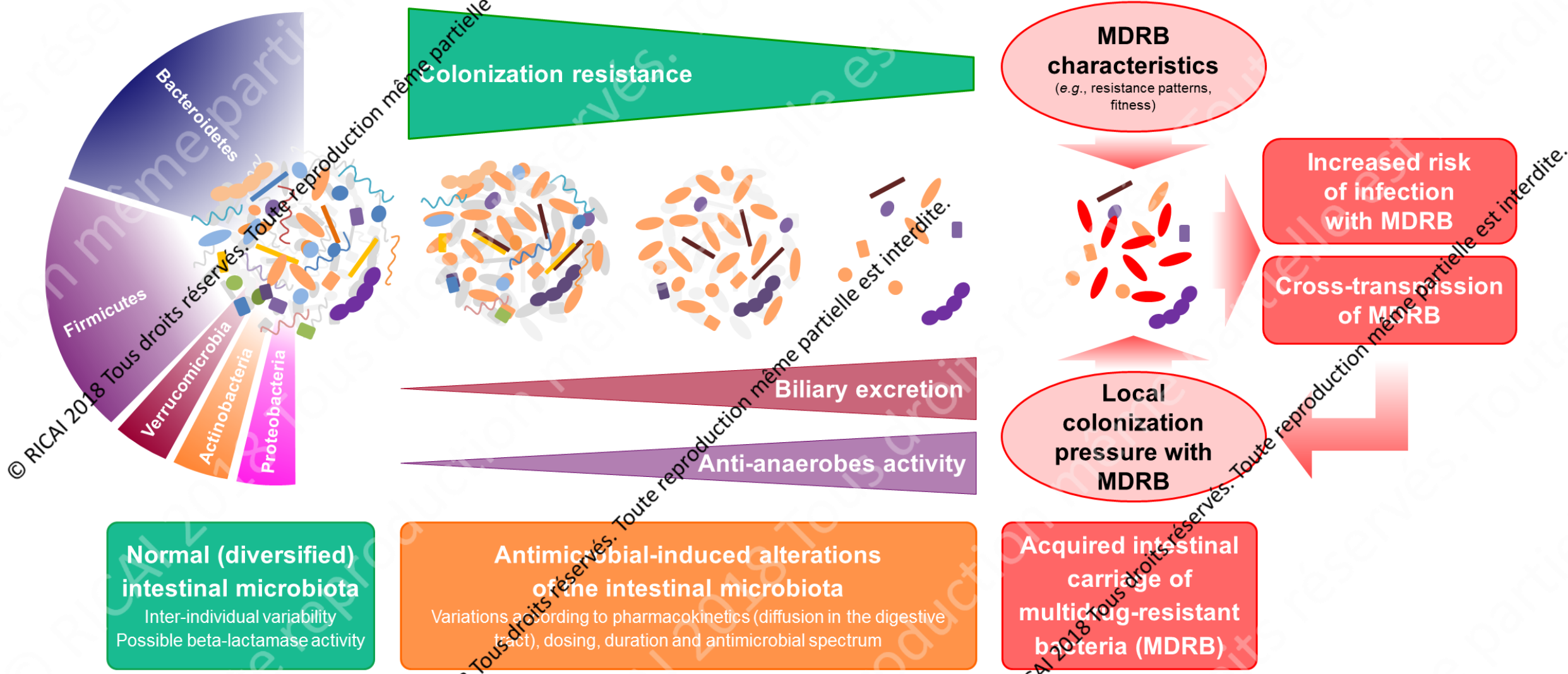


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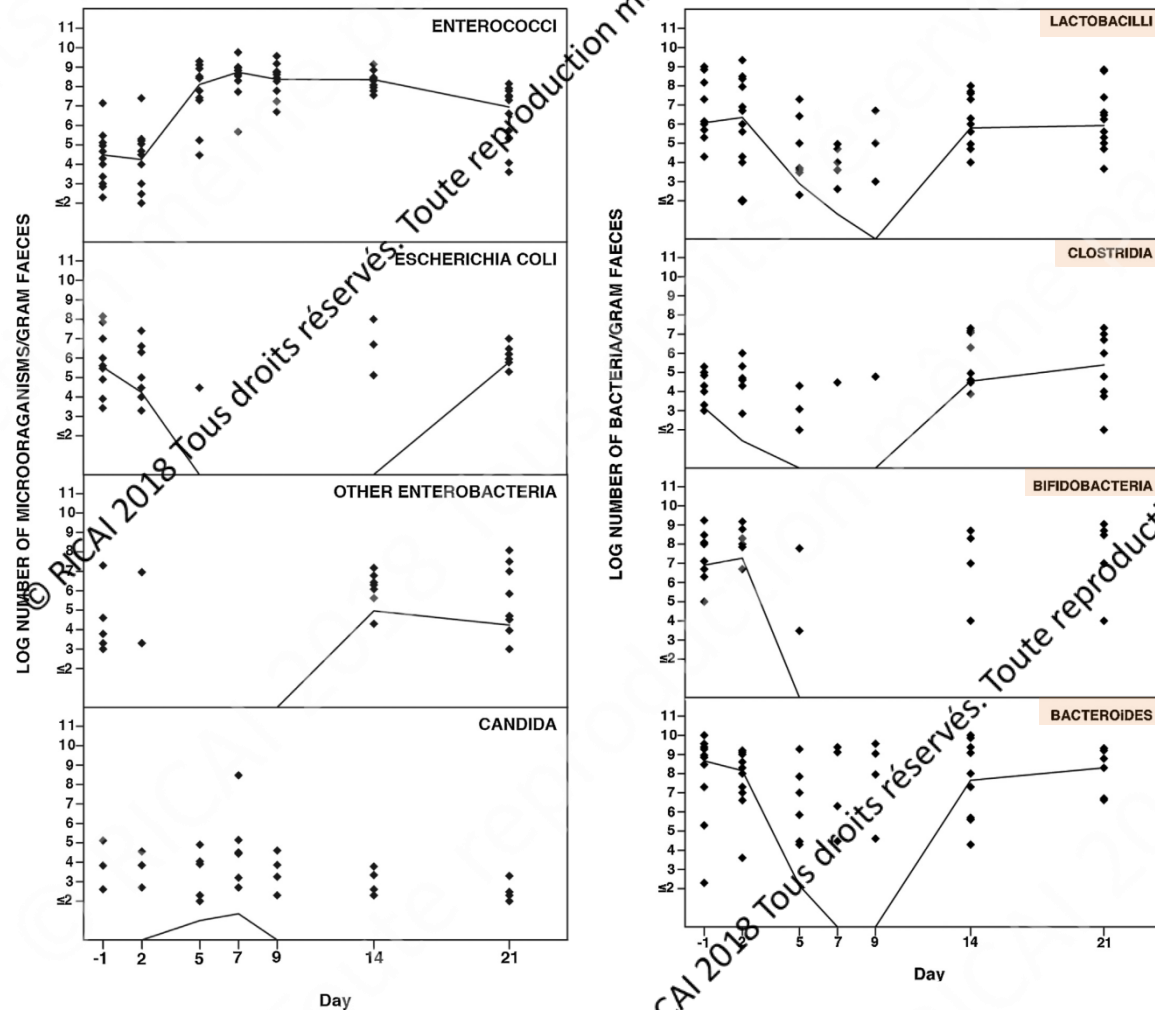
Impact of antibiotic exposure on intestinal colonization resistance



Ecological effect of ceftazidime/avibactam on the normal human intestinal microbiota

International Journal of Antimicrobial Agents 46 (2015) 60–65

Mamun-Ur Rashid^a, Staffan Rosenborg^{b,c}, Georgios Panagiotidis^{b,c}, Karin Söderberg Löfdal^{b,c}, Andrej Weintraub^a, Carl Erik Nord^{a,*}



- 12 volontaires sains
- Ceftazidime 2 gr + avibactam 500 mg/8h x 7 jours
- Cultures (pas de BM)
- **Concentrations dans les selles**
 - ✓ Ceftazidime 0-468 mg/kg
 - ✓ Avibactam 0-146 mg/kg
- ***Clostridium difficile* toxigène** : acquisition sous traitement chez 5/12 volontaires

Nouveaux antibiotiques anti-BGNMR

	Spectre d'activité						Impact écologique
	ESBLE	EPC KPC	EPC OXA48	EPC MBL	MDR- <i>Pa</i>	CR- <i>Ab</i>	
Ceftolozane-tazobactam							
Ceftazidime-avibactam							
Méropénème-vaborbactam							
Sulopenem							
Céfépime/AAI101							
Imipénème/relebactam							
Aztreonam/avibactam							
Céfidérol							
Plazomicine							
Eravacycline							

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Nouveaux antibiotiques anti-BGNMR

	Spectre d'activité						Impact écologique
	ESBLE	EPC KPC	EPC OXA48	EPC MBL	MDR- <i>Pa</i>	CR- <i>Ab</i>	
Ceftolozane-tazobactam	++	-	-	-	+++	-	
Ceftazidime-avibactam	+++	+++	+++	-	++	+	
Méropénème-vaborbactam	+++	+++	-	-	+/-	-	
Sulopenem	+++	-	-	-	-	-	
Céfépime/AAI101	++	+/-	-	+/-	+/-	-	
Imipénème/relebactam	+++	++	+/-	-	++	+/-	
Aztreonam/avibactam	+++	+++	+++	+++	+/-	-	
Céfidérol	+++	+++	+++	+++	+++	+++	
Plazomicine	+++	+++	+++	+/-	-	-	
Eravacycline	+++	++	++	++	-	+++	

Nouveaux antibiotiques anti-BGNMR

	Spectre d'activité						Impact écologique
	ESBLE	EPC KPC	EPC OXA48	EPC MBL	MDR- <i>Pa</i>	CR- <i>Ab</i>	
Ceftolozane-tazobactam	++	-	-	-	+++	-	?
Ceftazidime-avibactam	+++	+++	+++	-	++	+	(?)
Méropénème-vaborbactam	+++	+++	-	-	+/-	-	?
Sulopenem	+++	-	-	-	-	-	?
Céfépime/AAI101	++	+/-	-	+/-	+/-	-	?
Imipénème/relebactam	+++	++	+/-	-	++	+/-	?
Aztreonam/avibactam	+++	+++	+++	+++	+/-	-	?
Céfidérocol	+++	+++	+++	+++	+++	+++	?
Plazomicine	+++	+++	+++	+/-	-	-	?
Eravacycline	+++	++	++	++	-	+++	?

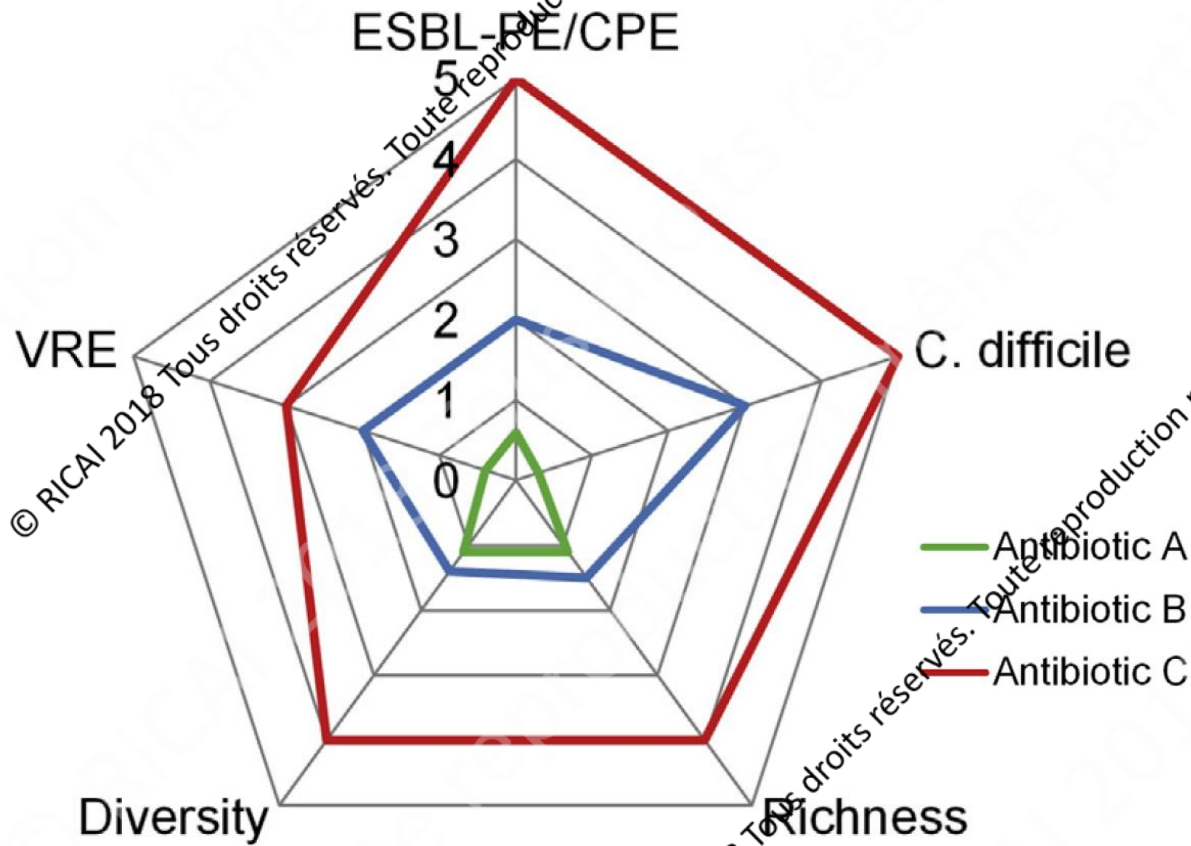
Impact des antibiotiques sur le microbiote intestinal : *take-home messages*

- L'impact écologique d'un antibiotique ne peut se résumer à son spectre clinique
- Implication probable de multiples paramètres, modifiables ou non :
 - ✓ Activité anti-anaérobie, biodisponibilité intestinale, durée de traitement
 - ✓ Cofacteurs possibles de dysbiose intestinale
 - ✓ Type de BMR et pression de colonisation (mesures d'hygiène)
 - ✓ Microbiote « basal » (résistance à la colonisation)
- **Préserver ou restaurer la résistance à la colonisation :**
 - ✓ *Antibiotic stewardship*
 - ✓ Absorption des antibiotiques dans la lumière intestinale (ex: DAV 132)
 - ✓ Probiotiques, FMT

Impact of antibiotics on the intestinal microbiota needs to be re-defined to optimize antibiotic usage

Clinical Microbiology and Infection 24 (2018) 3–5

E. Ruppé^{1,2,3,*}, C. Burdet^{1,2,4}, N. Grall^{1,2,3}, V. de Lastours^{1,2,5}, F.-X. Lescure^{1,2,6},
A. Andremont^{1,2,3}, L. Armand-Lefèvre^{1,2,3}



Nécessité d'études comparatives dans des environnements à forte pression de colonisation :
Carbapénèmes vs alternatives
Nouveaux antibiotiques anti-BGNMR